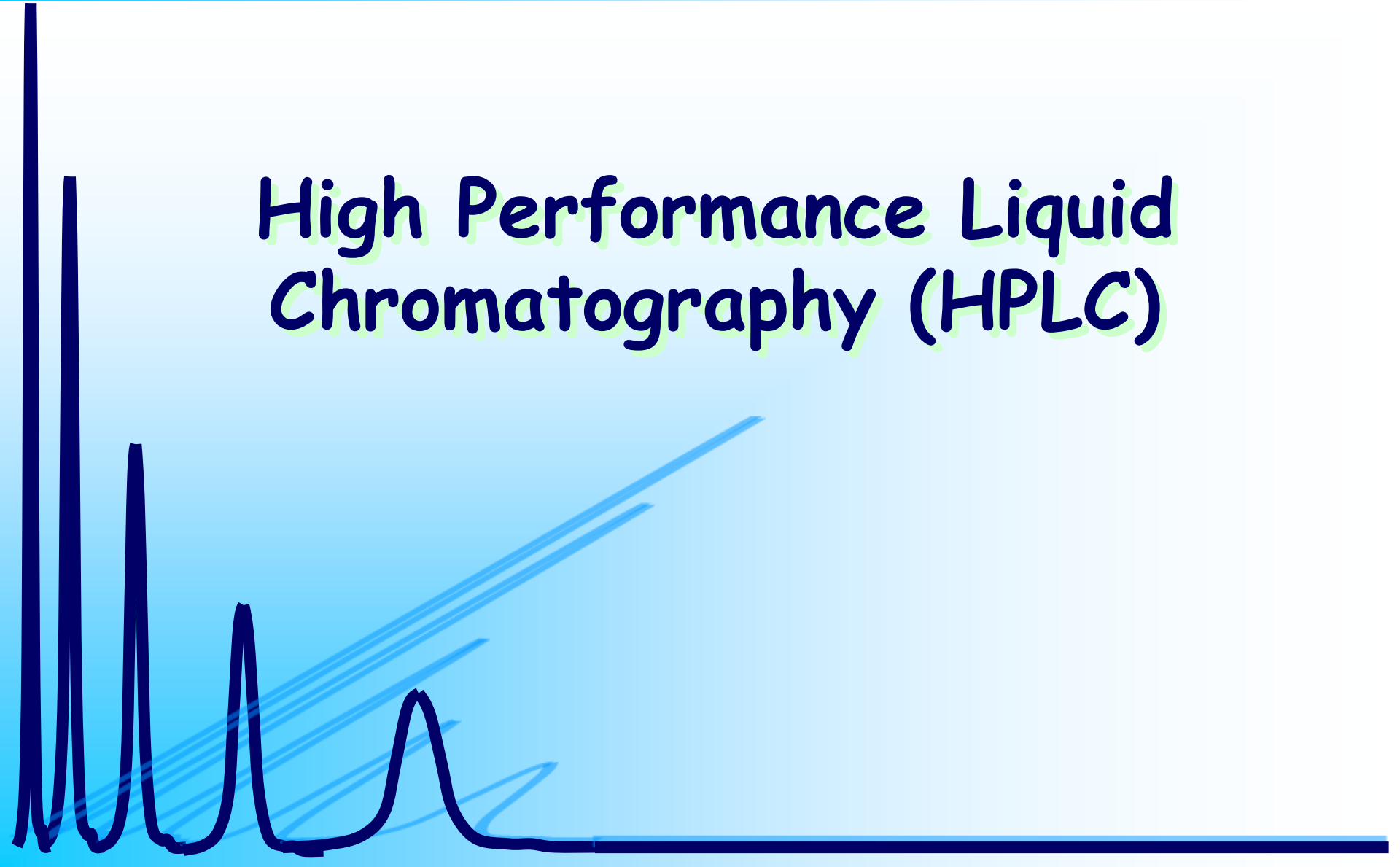


A stylized chromatogram is shown on the left side of the slide. It features a dark blue baseline with five distinct peaks of decreasing height from left to right. A light blue diagonal line with a dashed effect extends from the bottom left towards the top right, passing behind the peaks. The title text is positioned to the right of the chromatogram.

High Performance Liquid Chromatography (HPLC)

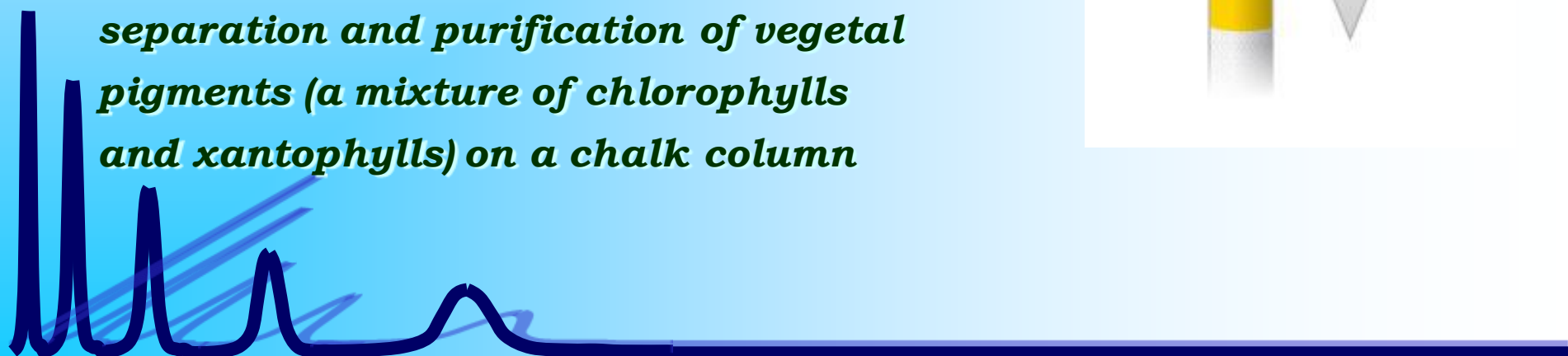
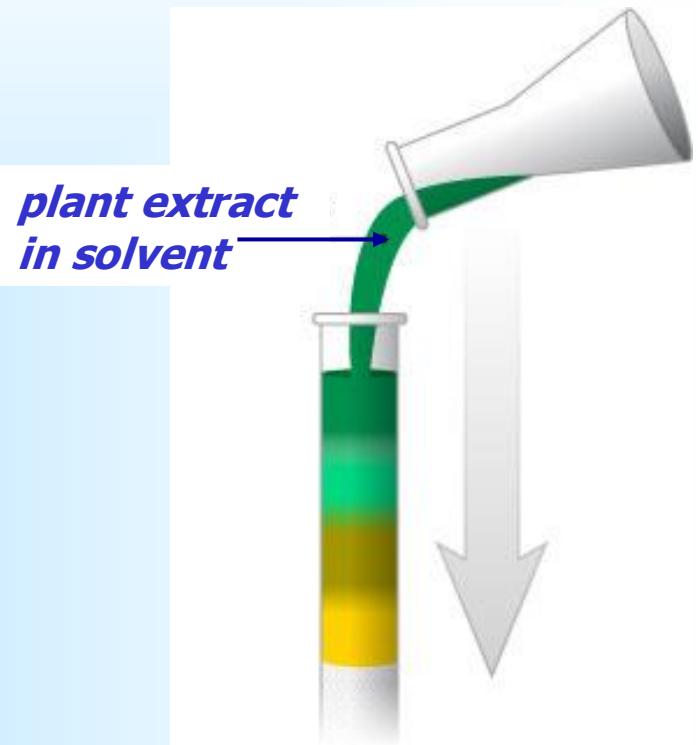
Liquid chromatography

- It is the first ever described chromatographic method (by Tswett in 1903)
- Unlike gas chromatography, the sample in liquid chromatography must not be vaporized so, almost all kinds of compounds can be analysed by liquid chromatography
- The development of instrumental liquid chromatography was later than for gas chromatography because of the higher pressure needed for the former
- HPLC is considered to be the major chromatographic technique available today for non-volatile or heat-sensitive substances.



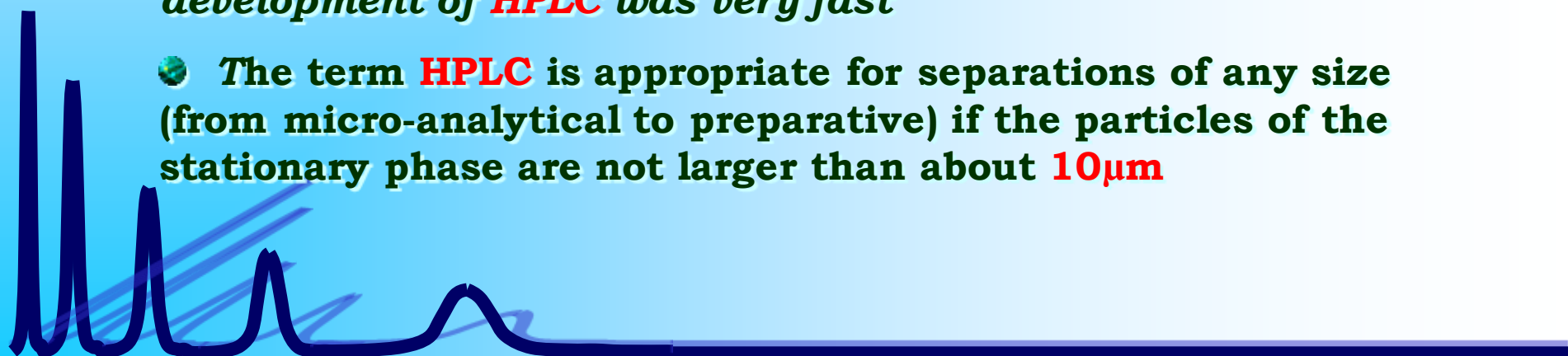
A little history:

● The official date of birth of chromatography is the **21 March 1903** in Warsaw when **Mikhail Semenovitch TSWETT** has presented at the Congress of the **Polish Natural Sciences Society** a communication entitled: « **A new class of adsorption phenomena and their applications in biochemical analysis** » about the separation and purification of vegetal pigments (a mixture of chlorophylls and xanthophylls) on a chalk column



A little history:

- **1938** : **REICHSTEIN** proposes a theory for the **elution** and **separation** of solutes on a column
- **1952** : application of gradient elution
- **1967** : beginning of HPLC after the works of **HUBER** and **HUZSMAN**, this technique was first named « **High Speed Liquid Chromatography** » then « **High Pressure Liquid Chromatography** » and finally « **High Performance Liquid Chromatography** »
- **1969** : after the 5th International Symposium International « **Advances in Chromatography** » the development of **HPLC** was very fast
- The term **HPLC** is appropriate for separations of any size (from micro-analytical to preparative) if the particles of the stationary phase are not larger than about **10µm**



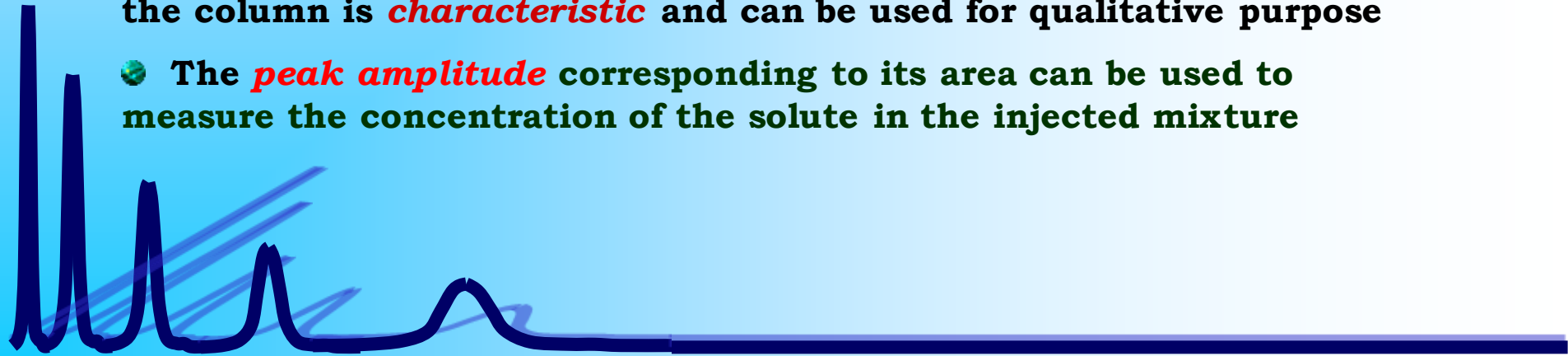
Fundamental definitions (to IUPAC nomenclature)

- **Chromatography:** a **physical** method of **separation** in which the components to be separated are distributed between **two phases**, one of which is stationary (**stationary phase**) while the other (the **mobile phase**) moves in a definite direction
- **Chromatogram:** a **graphical** or other presentation of **detector response**, concentration of analyte in the effluent or other quantity used as a measure of effluent concentration versus effluent volume or time
- **Stationary Phase:** one of the two phases forming a chromatographic system. It may be a **solid**, a **gel** or a **liquid**. If a liquid, it may be **distributed** on a solid. This solid may or may not contribute to the separation process. The liquid may also be **chemically bonded** to the solid (**bonded phase**: covalently bonded to the support particles or to the inside wall of the column tubing) or immobilized onto it (**immobilized phase**)
- **Mobile Phase:** a **fluid** which percolates through or along the stationary bed, in a definite direction. It may be a **liquid** (**liquid chromatography**) or a **gas** (**gas chromatography**) or a **supercritical fluid** (**supercritical-fluid chromatography**)



Principle of liquid chromatography

- A liquid used as **mobile phase** moves along a tube used as **column**. This column is packed with a solid support which plays the role of **stationary phase**
- If the **stationary phase** and the **mobile phase** were correctly selected, the constituents of mixture are unequally retained along the column
- This phenomenon called **retention** means that the injected solutes move **slower** than the mobile phase with **different speeds**. They are thus **eluted** successively from the column and **separated**
- The **detector** connected at the column outlet gives a signal corresponding to each solute which is recorded as the **chromatogram**
- In the working conditions, the **retention time** spent by each solute in the column is **characteristic** and can be used for qualitative purpose
- The **peak amplitude** corresponding to its area can be used to measure the concentration of the solute in the injected mixture



Simplified scheme of the chromatographic separation process

injector —————→ flow of mobile phase —————→ detector

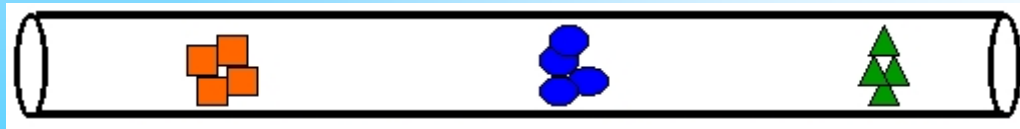
$t = 0$



$t = 5\text{mn}$



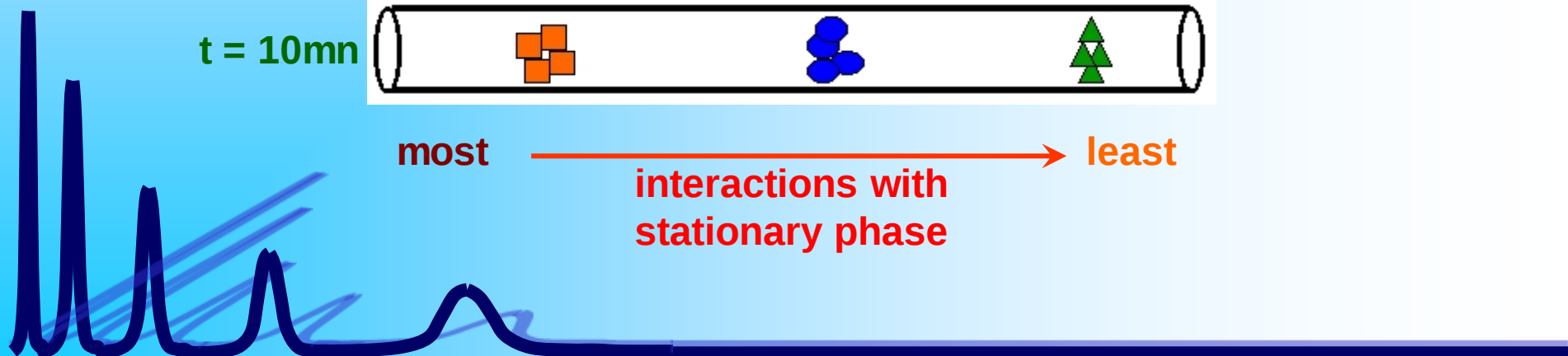
$t = 10\text{mn}$



most

—————→
interactions with
stationary phase

least



Main modes in liquid chromatography

There are several modes in high performance liquid chromatography

They are classified according to the **mechanism of separation**

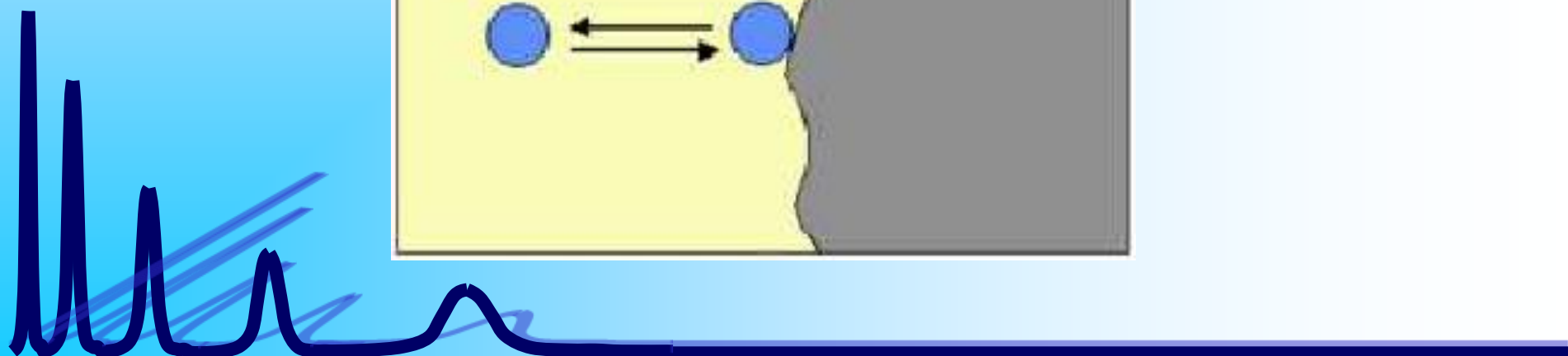
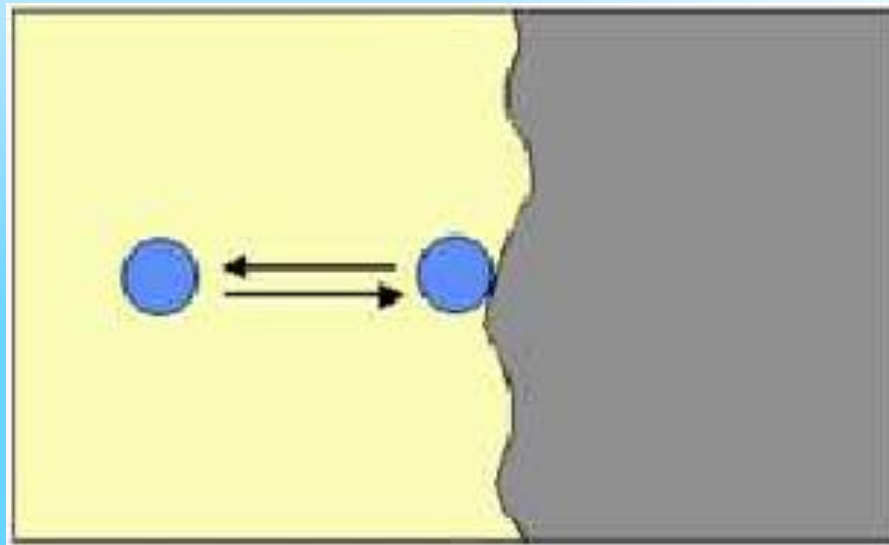
Each mode corresponds to a given kind of ***interaction***:

- ***surface adsorption***
- ***solvent partitioning***
- ***ion exchange***
- ***size exclusion***



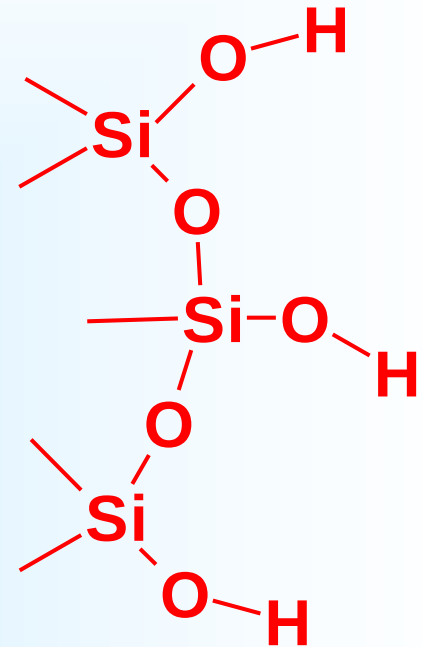
Adsorption chromatography

- the stationary phase is a **solid adsorbant**
- retention is due to a series of **adsorption / desorption** steps
- separation is based mainly on differences between the **adsorption affinities** of the sample components for the surface of the active solid (**liquid solid chromatography**)

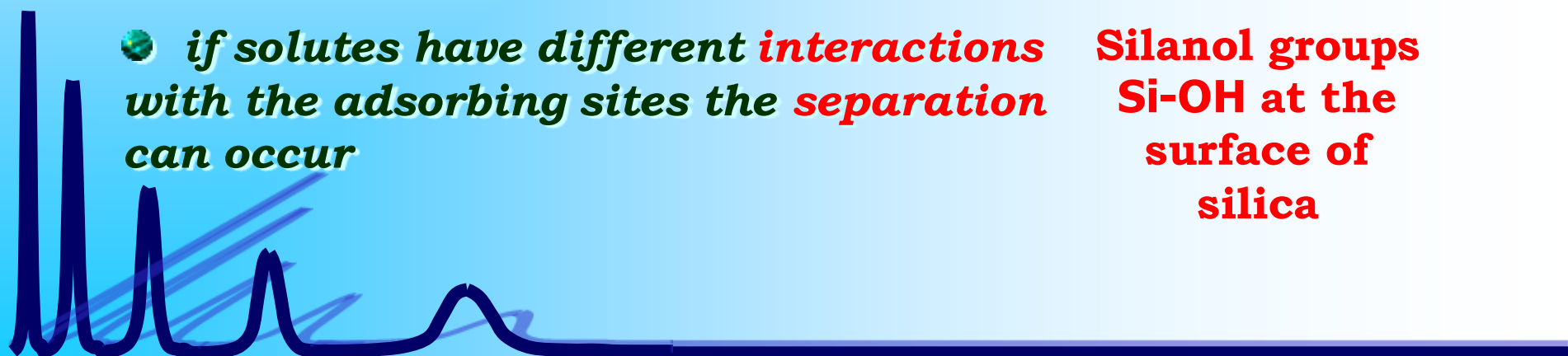


Adsorption chromatography

- **silica** and **alumina** are the most used stationary phases
- both solute and solvent can be **attracted** by the active sites at the **surface** of the stationary phase
- the molecules are retained by the **interaction** of their polar functional groups with the **surface functional groups** such as **silanols** of silica
- if solutes have different **interactions** with the adsorbing sites the **separation** can occur

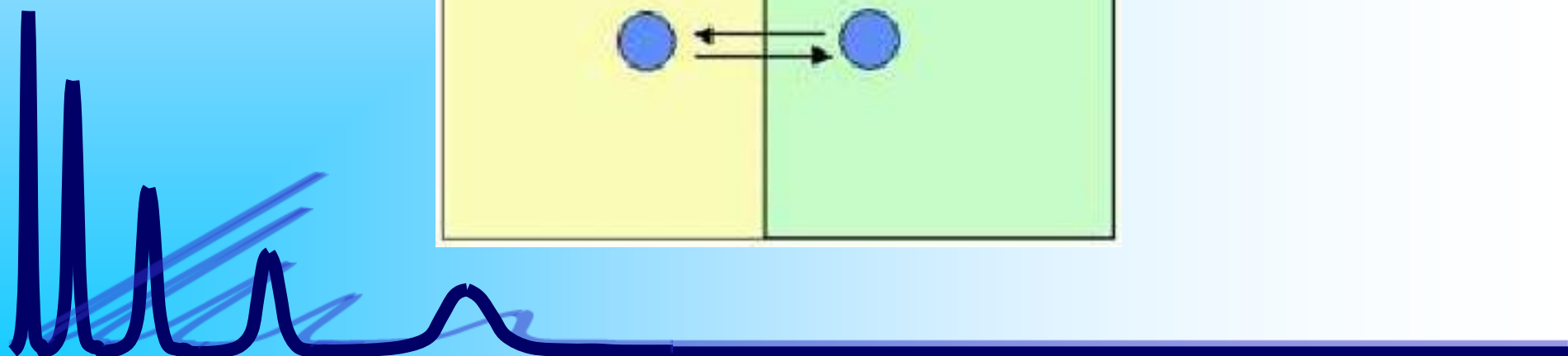
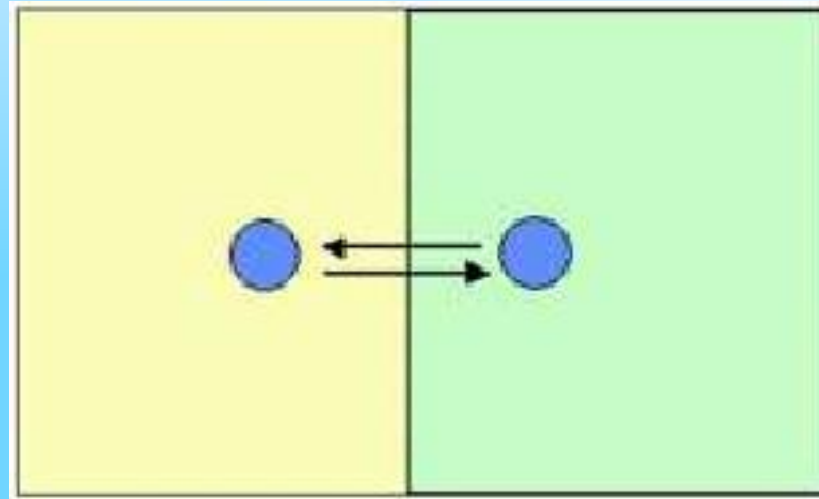


**Silanol groups
Si-OH at the
surface of
silica**



Partition chromatography

- the stationary phase is a **liquid coated** or **linked** to a **solid support**
- retention is due to the **partitioning** of the solute between the two **liquid** phases (**relative solubility**)
- separation is based mainly on differences between the **solubilities** of the components in the mobile and stationary phases (**liquid liquid chromatography**)



Partition chromatography

- the most retained species is that having the highest **affinity** (**solubility**) for the liquid stationary phase, relatively to the mobile phase (**eluent**)
- separation is based on the differences in **relative solubility**

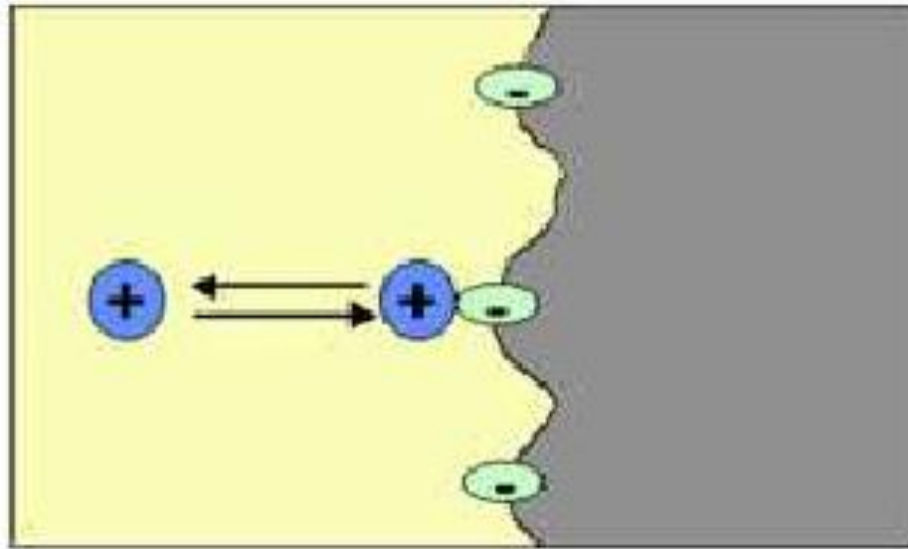
There are two modes in liquid chromatography

- « **normal** » mode : **polar stationary phase** and **non-polar mobile phase** (the first mode described). In this procedure, the stationary phase is more polar than the mobile phase. This term is used in liquid chromatography to emphasize the contrast to reversed-phase chromatography
- « **reversed-phase** » mode: **non-polar stationary phase** and **polar mobile phase** (the most used mode). In this procedure the mobile phase is significantly more polar than the stationary phase, e.g., a microporous silica-based material with chemically bonded alkyl chains



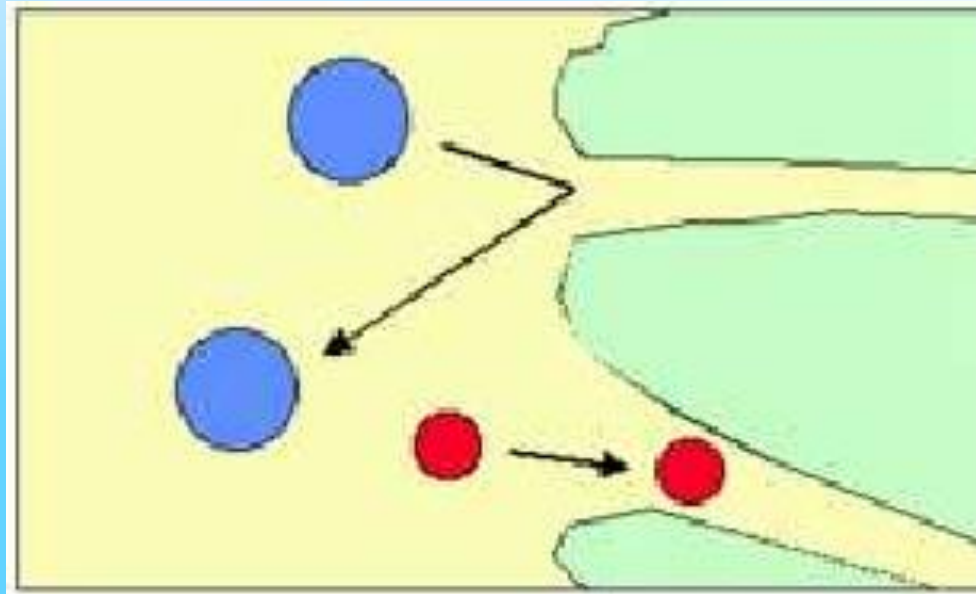
Ion exchange chromatography (IEC)

- the stationary phase has **ionically charged groups** at the surface
- the retention is due to the **attractive interactions** between **ionic solutes** and the opposite **charged** stationary phase
- separation is based mainly on differences in the **ion exchange affinities** of the sample components
- this technique is now often referred to as **Ion Chromatography (IC)**



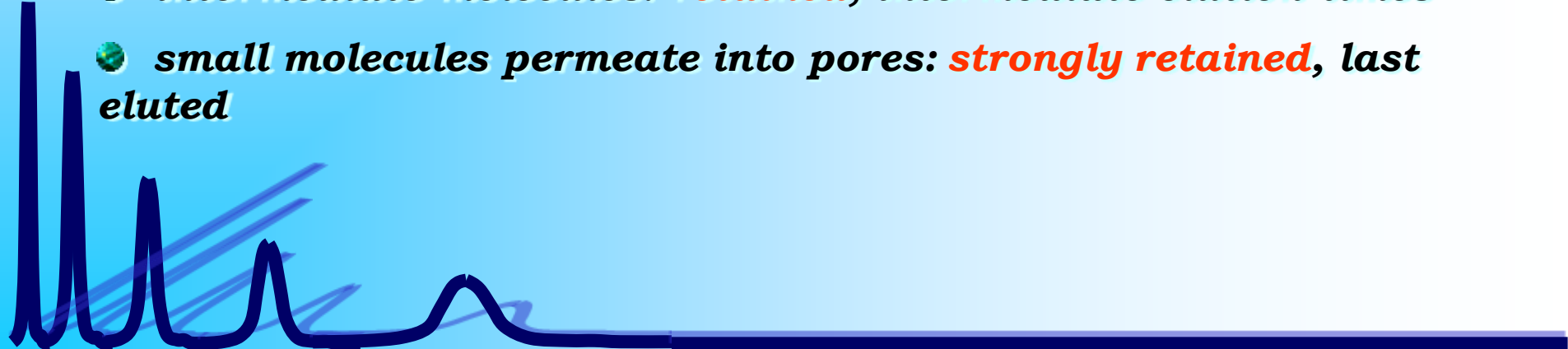
Size exclusion chromatography (SEC)

- the stationary phase is a **porous** material having controlled **pore size**
- separation is based mainly on **exclusion effects**, such as differences in **molecular size** and/or **shape**
- the terms **Gel Filtration** and **Gel-Permeation Chromatography** (GPC) were used earlier to describe this process



Size exclusion chromatography (SEC)

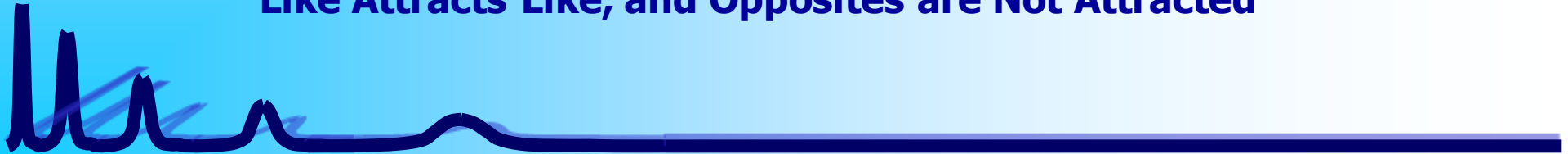
- In this mode, each column can separate solutes having **specific size range**
- separation mechanism is **sieving**
- the **larger** species cannot enter all the pores and will **elute first** because they have a **shorter** path in the column
- this mode is very useful for the **determination of molecular size** of macromolecules (polymers, proteins,...)
- large molecules **excluded** from pores - not retained, first eluted
- intermediate molecules: **retained**, intermediate elution times
- small molecules permeate into pores: **strongly retained**, last eluted



Importance of polarity in HPLC

- the notion of **polarity** plays a fundamental role in HPLC
- all chemicals have a unique and characteristic behaviour related to their molecular structure and electron charge distribution
- they can be described as being “**polar**” or “**non-polar**”, with a range of polarities between the most polar and most non-polar
- **water** is a good example of a very polar liquid, and **paraffin** based oil is a good example of a very non-polar liquid
- this “**polarity**” characteristic of chemicals allows to explain the chromatographic “**retention mechanisms**” that are used to create many HPLC separations
- a simple rule describes this behavior for polarity-based retention mechanisms:

“Like Attracts Like, and Opposites are Not Attracted”



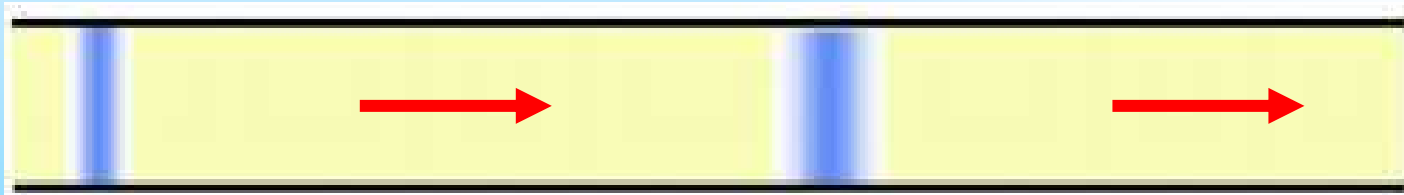
Mobile phase composition:

- ***isocratic analysis***: in this procedure the composition of the mobile phase remains **constant** during the elution process
- ***gradient elution***: in this procedure the composition of the mobile phase is **changed continuously** or **stepwise** during the elution process

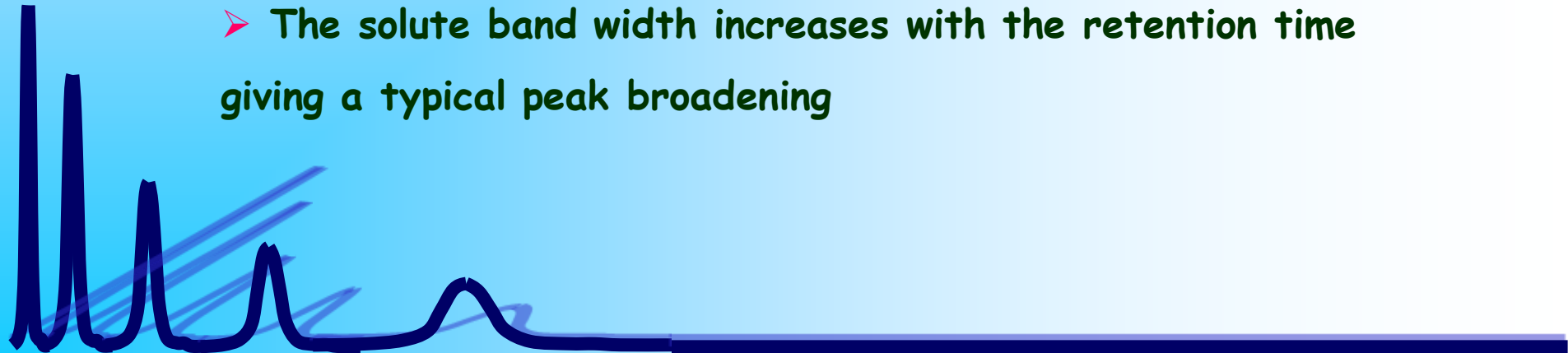


Elution process:

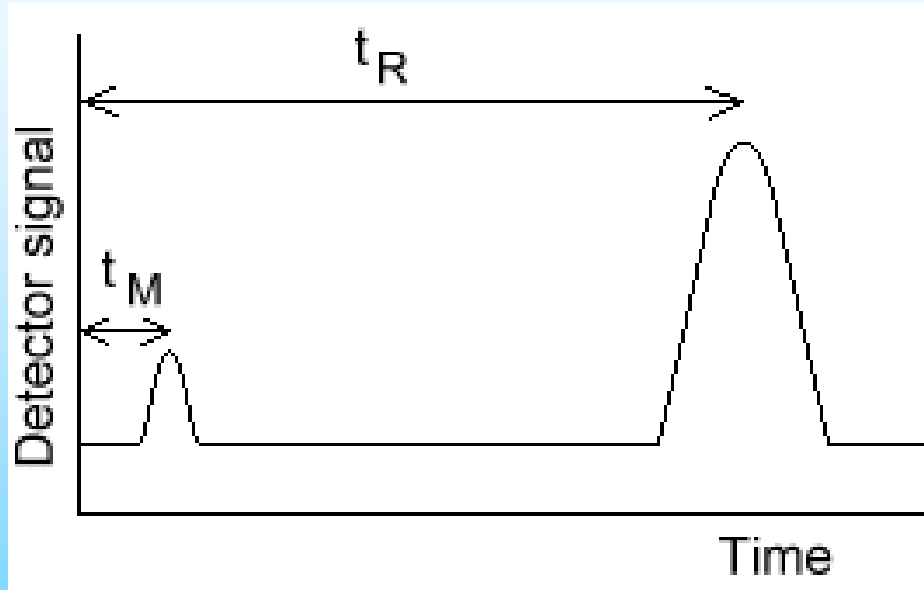
- During the solute transfer in the column, it shows a typical broadening due to the diffusion phenomena (transversal and longitudinal)



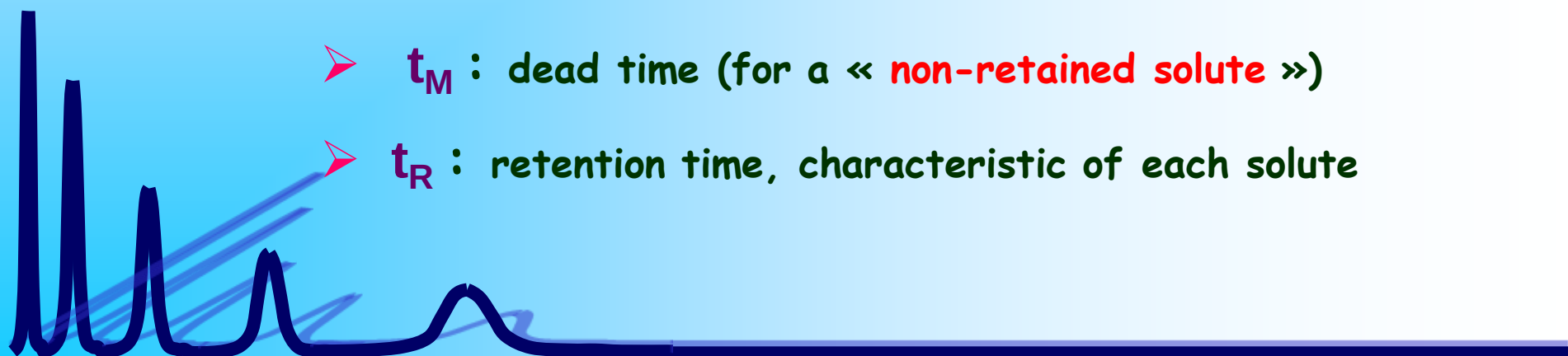
- The solute band width increases with the retention time giving a typical peak broadening



The chromatogram: characteristic parameters



- t_M : dead time (for a « non-retained solute »)
- t_R : retention time, characteristic of each solute



Some fundamental equations:

- Corrected retention time:

$$t'_R = t_R - t_M$$

- average linear velocity ($\bar{u}$) is measured from the retention time of an unretained substance (t_M) which moves at the same velocity as the mobile phase:

$$\bar{u} = L / t_M$$

- Retention factor (or capacity ratio) k : corresponds to a relative retention:

$$k = t'_R / t_M = (t_R - t_M) / t_M$$

- since:

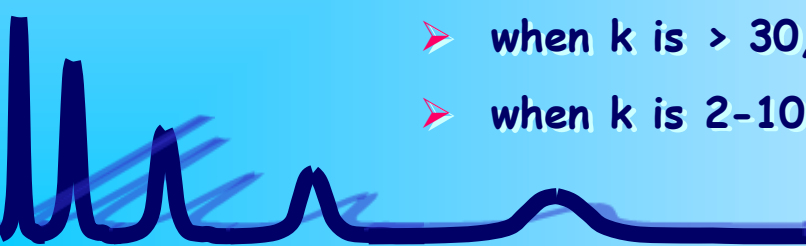
$$t_M = L / \bar{u}$$

we can write:

$$t_R = (1 + k) \cdot t_M = (1 + k) \cdot L / \bar{u}$$

Hence the retention time is directly proportional to the column length L and inversely proportional to the linear flow rate of the mobile phase $\bar{u}$

- when k is ≤ 1.0 , separation is **poor**
- when k is > 30 , separation is **slow**
- when k is 2-10, separation is **optimum**



Some fundamental equations:

➤ average linear velocity ($\bar{u}$) is measured from the retention time of an unretained substance (t_M) which moves at the same velocity as the mobile phase:

$$\bar{u} = L / t_M = F / q$$

where: F is the mobile phase flowrate

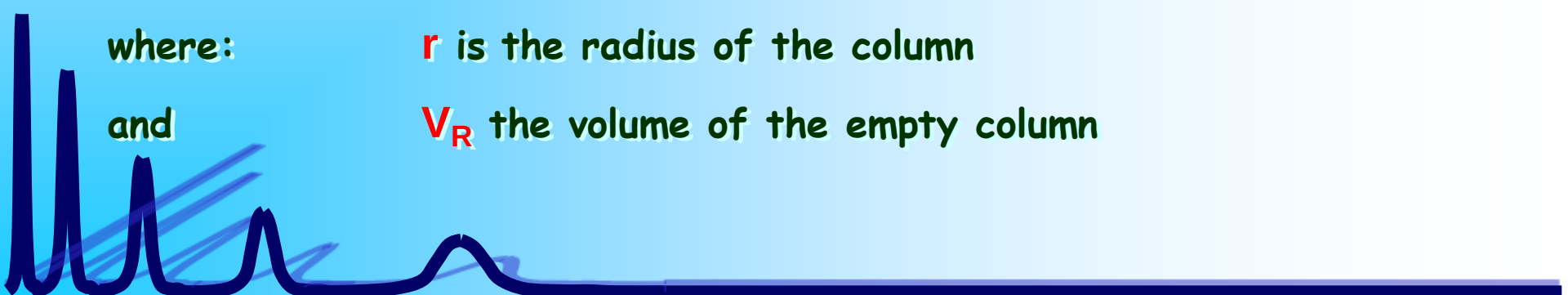
and q the free cross-sectional area of the column

➤ the column porosity ϵ_T is:

$$\epsilon_T = q / \pi \cdot r^2 = F \cdot t_M / \pi \cdot r^2 \cdot L = F \cdot t_M / V_R$$

where: r is the radius of the column

and V_R the volume of the empty column



Column efficiency:

The chromatographic peaks being supposed **gaussian**, the peak broadening can be related to the separation and the column efficiency which is evaluated by the **number of theoretical plates** of the column **N** (similarly to distillation process) which is a number indicative of column performance.

For a **gaussian peak**, **N** can be calculated by one of the following equations:

$$N = (t_R / \sigma)^2 \quad (\sigma: \text{standard deviation of the peak})$$

$$N = 16 (t_R / \omega)^2 \quad (\omega: \text{width at baseline})$$

$$N = 5,54 (t_R / \delta)^2 \quad (\delta: \text{width at half- height})$$

In order to compare columns having different lengths, one calculate the **plate height** or **height equivalent to a theoretical plate HETP**:

$$H = L/N \quad (L : \text{column length})$$

H may vary from centimeters (**packed columns**) to several microns (**high resolution capillary columns**)



Column efficiency:

➤ Column selectivity:

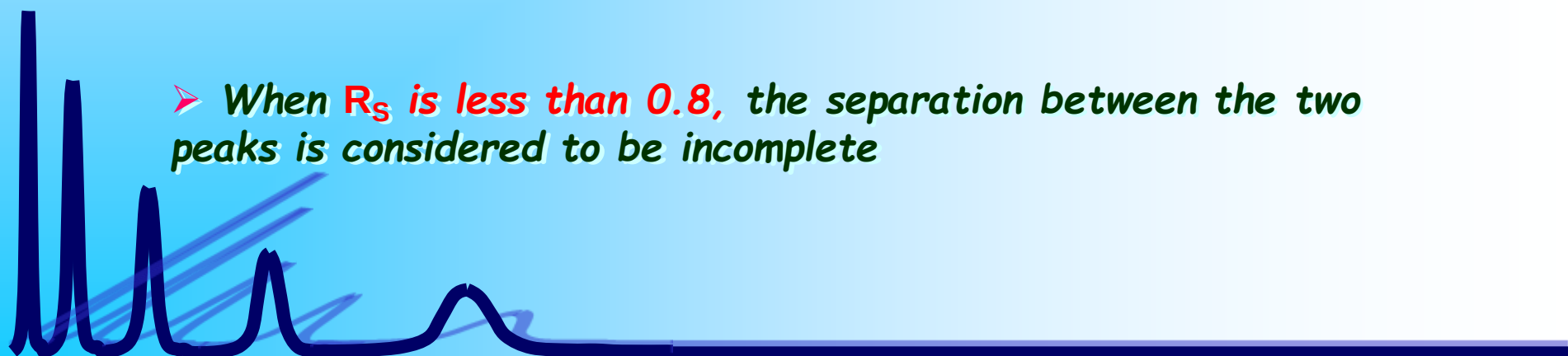
$$\alpha = t'_{R2} / t'_{R1} = k_2 / k_1 \quad (\text{separation occurs only if } \alpha > 1)$$

➤ Resolution between two neighboring peaks:

$$R_S = 2 (t_{R2} - t_{R1}) / (\omega_2 + \omega_1) = 1,18 (t_{R2} - t_{R1}) / (\delta_2 + \delta_1)$$

➤ For two neighboring peaks, a resolution R_S higher than 1 means a complete separation (for $R_S = 1$, the overlapping peak surface is 2%)

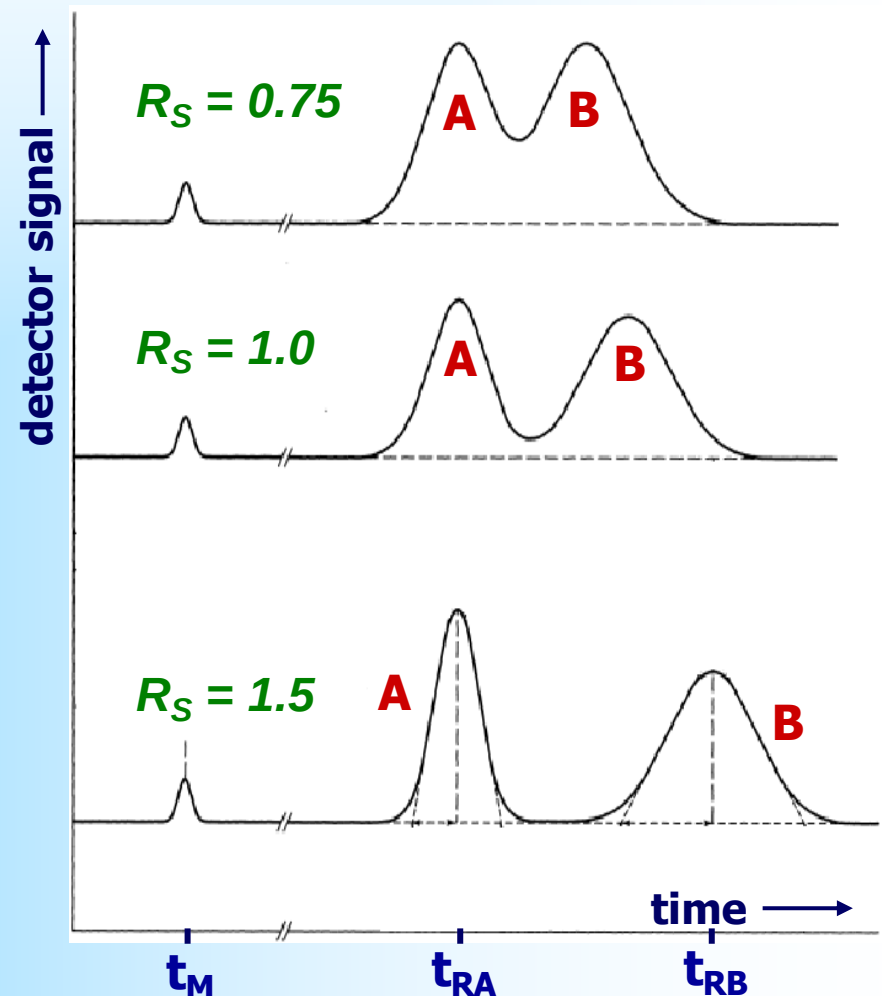
➤ When R_S is less than 0.8, the separation between the two peaks is considered to be incomplete



Optimization of column efficiency and resolution

Resolution (and zone broadening) depends on:

- $\bar{u}$ (linear flow rate): low flow favors increased resolution
- H (plate height) (or N number of plates): use smaller particles, lengthen column, viscosity of mobile phase (diffusion)
- α (selectivity factor): vary temperature, composition of column/mobile phase
- k (capacity factor): vary temperature, composition of column/mobile phase



Optimization of chromatographic separation

From the equations of:

- resolution: $R = 2 (t_{r,B} - t_{r,A}) / (\omega_B + \omega_A)$
 - and column efficiency: $N_B = 16 (t_{r,B} / \omega_B)^2$
 - if we assume that: $\omega_B \approx \omega_A$
- } we can deduce that the resolution is given by:

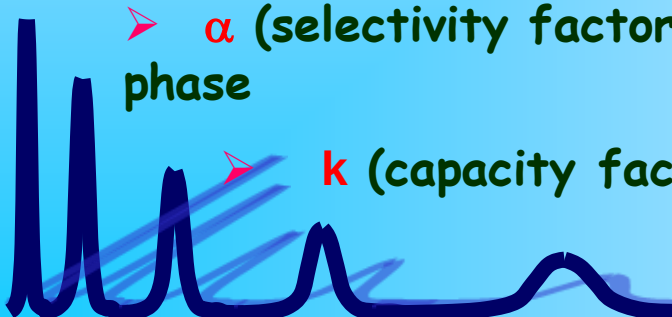
$$R = \frac{1}{4} \sqrt{N_B} \left(\frac{t_{r,B} - t_{r,A}}{t_{r,B}} \right)$$

then

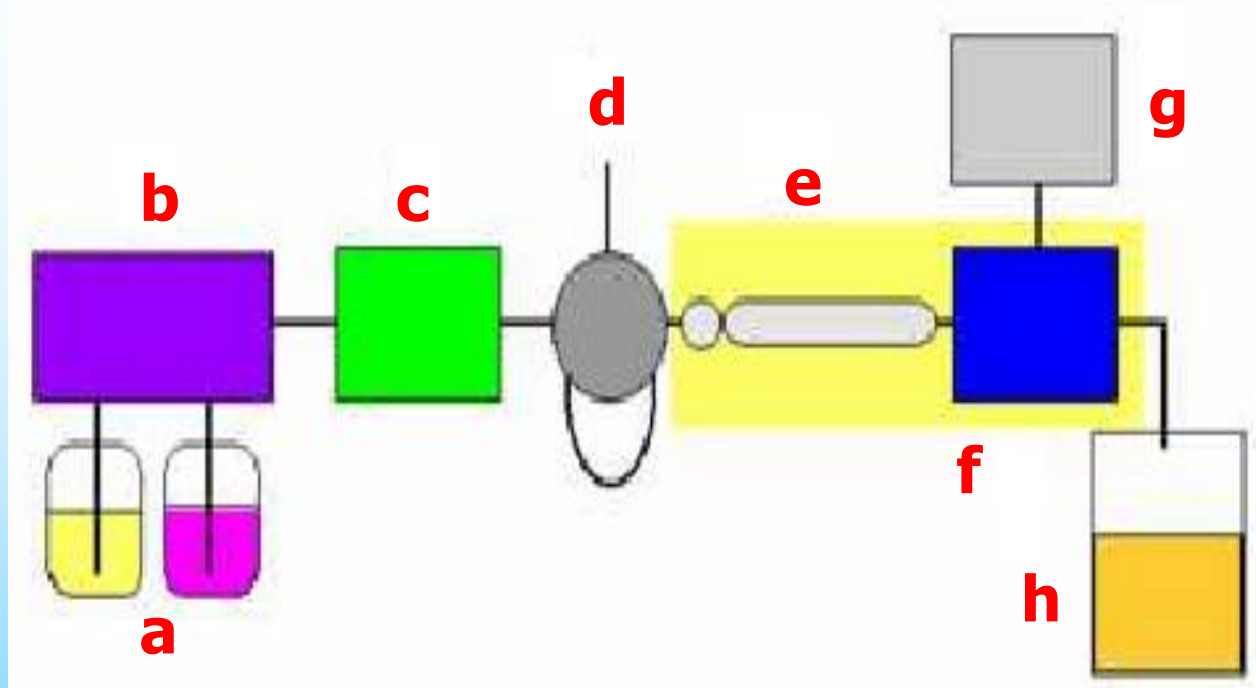
$$R = \frac{1}{4} \sqrt{N_B} \left(\frac{\alpha - 1}{\alpha} \right) \left(\frac{k'_B}{1 + k'_B} \right)$$

To obtain the desired resolution, we can control:

- **N** number of plates (or plate height **H**): use smaller particles, lengthen column, change viscosity of mobile phase (diffusion)
- **α** (selectivity factor): vary temperature, composition of column/mobile phase
- **k** (capacity factor): vary temperature, composition of column/mobile phase



Main parts of HPLC equipment



a- solvent reservoirs

b- gradient elution controller

c- pumping system

d- sample injection

e- precolumn and column

f- detector

g- data processing

h- solvent waste



Agilent HPLC equipment

mobile phase storage

filtration and degassing

preparation of eluent mixtures

HPLC pump

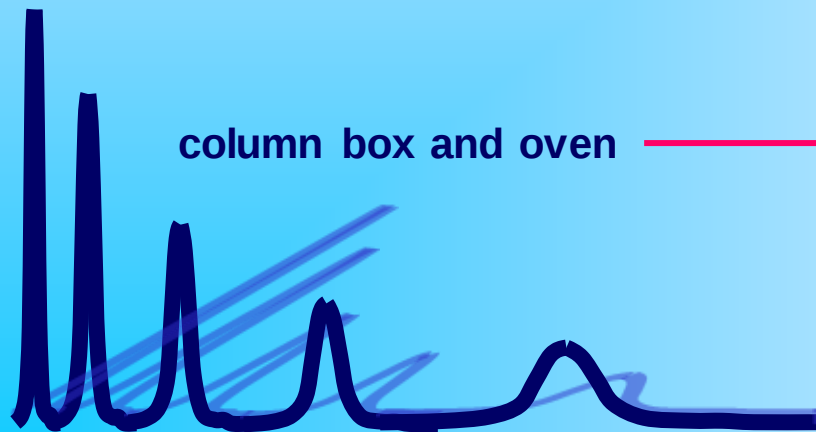
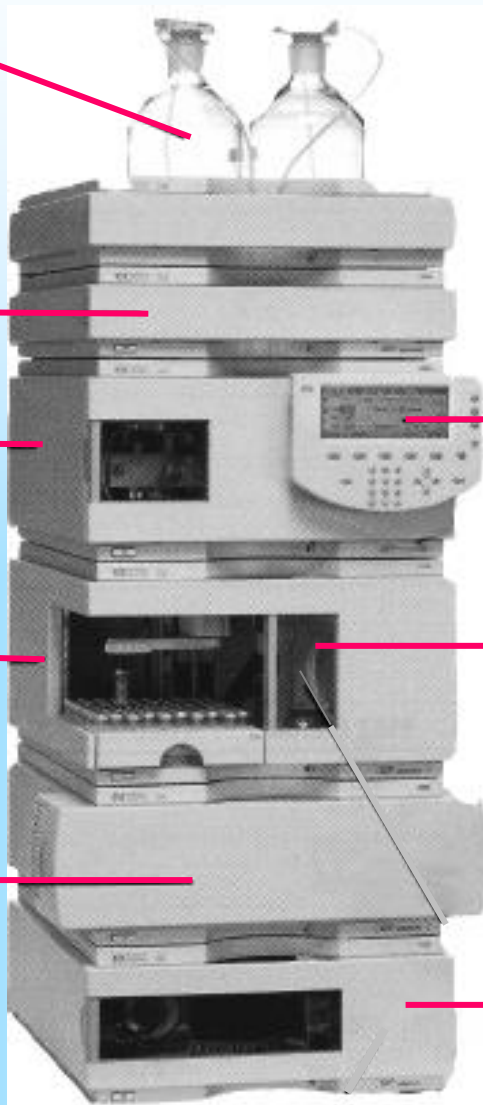
autosampler

column box and oven

control unit

injection valve

detection unit with
UV-visible lamp



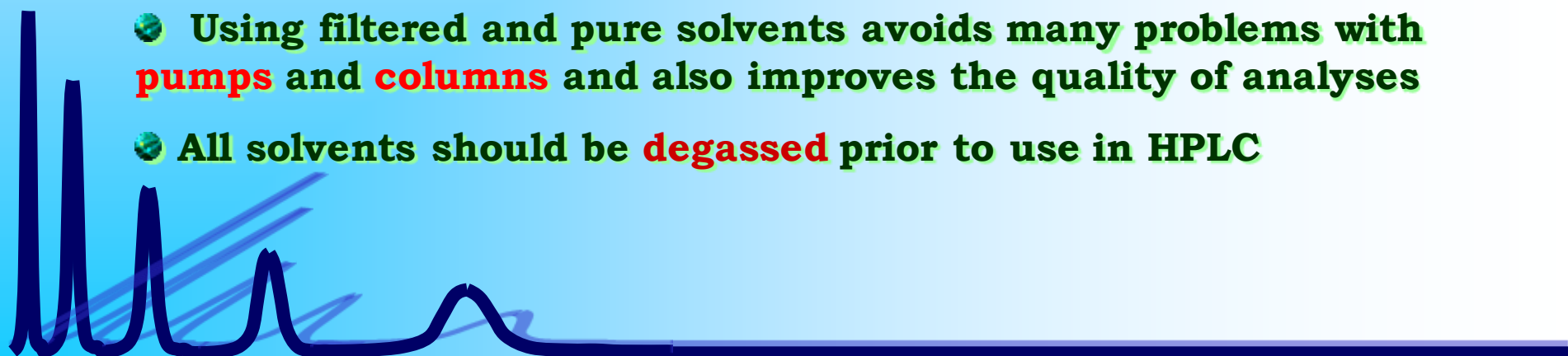
High performance liquid chromatography (HPLC) animation

<https://www.youtube.com/watch?v=ZN7euA1fS4Y>



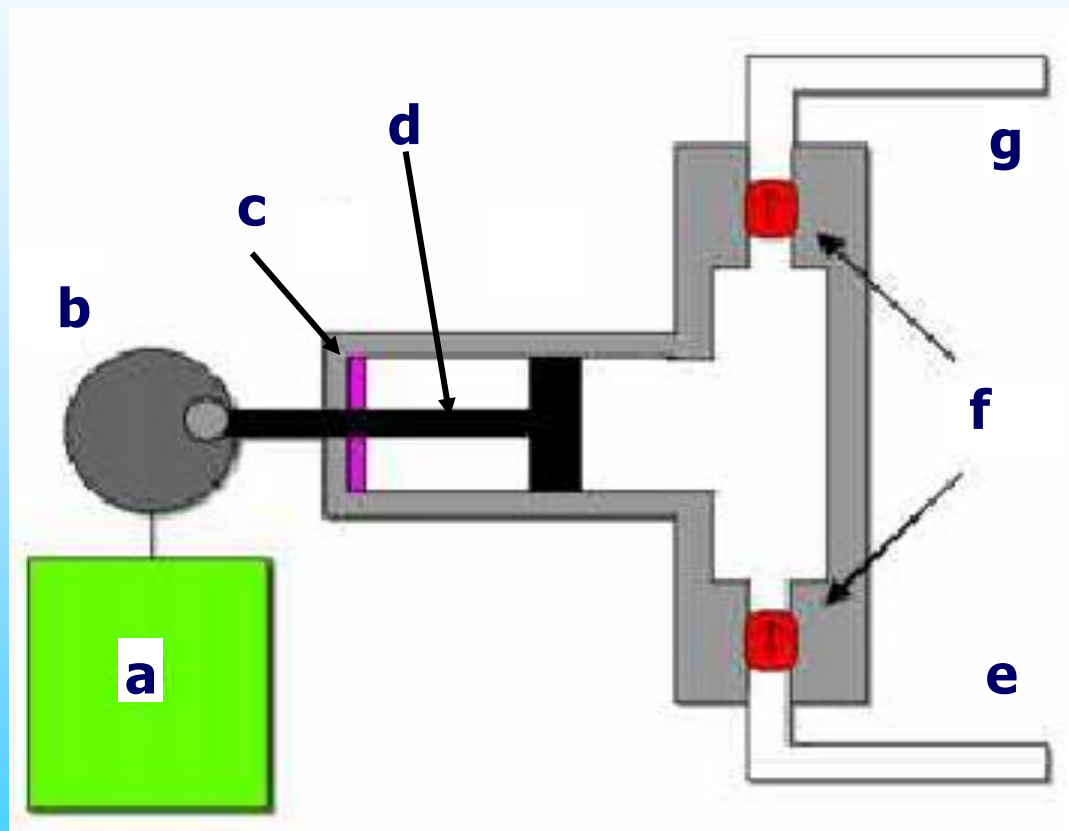
The mobile phase

- In HPLC the quality of the mobile phase is crucial
- The solvent is either **water** (preferred) or **organic**
- All solvents should be of '**HPLC grade**'
- This means:
 - a high **purity**
 - a prior **filtration** on a 0.2µm filter
- The '**HPLC grade**' solvent can be either purchased or produced using a convenient filtration device
- Using filtered and pure solvents avoids many problems with **pumps** and **columns** and also improves the quality of analyses
- All solvents should be **degassed** prior to use in HPLC



Reciprocating piston pump

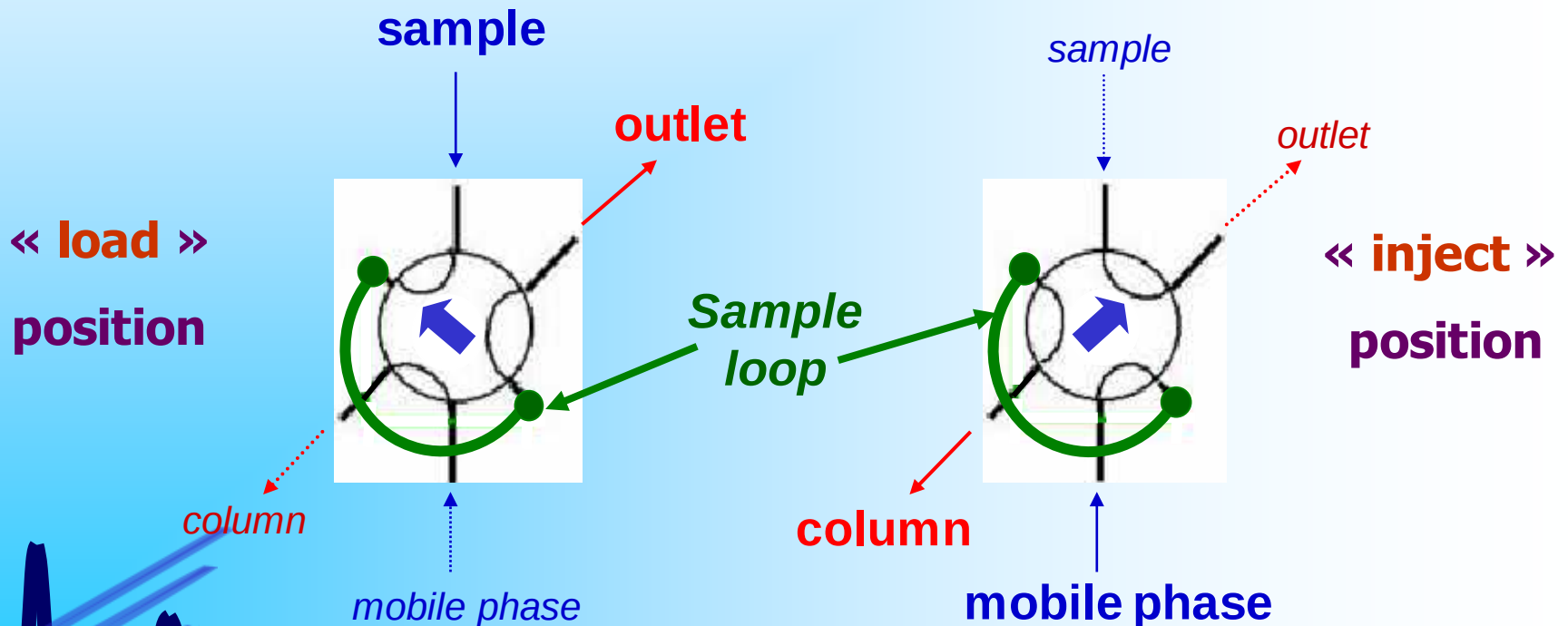
It is the most widespread pumping system used in HPLC:



- a- motor**
- b- transmission**
- c- tightness seal**
- d- piston**
- e- solvent inlet**
- f- inlet and outlet checkvalves**
- g- solvent outlet**

Injection system in HPLC

- The best injection device uses a switching **six-port valve** with a **sampling loop**
- This system allows an **easy** and **reproducible injection** of liquid samples
- It operates in two steps: **load** and **inject**



The HPLC column

- Most stationary phases are **chemically bonded** to the solid support which means a greater stability
- The stationary phase parameters are accurately controlled to assure **constant** and **reproducible** performances
- Packing techniques have also improved in order to obtain **smaller** but **more efficient** columns

A typical HPLC column



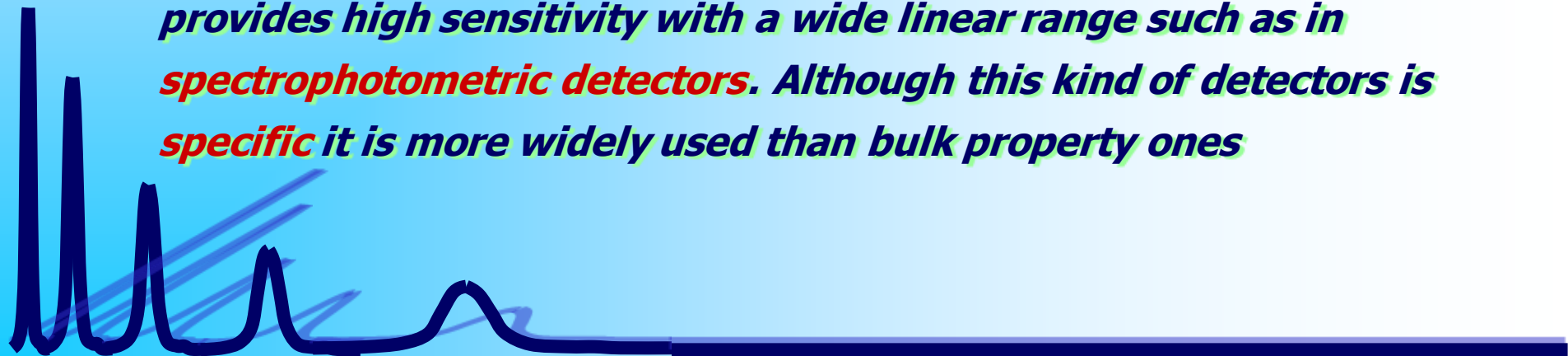
End fittings



Detection in HPLC

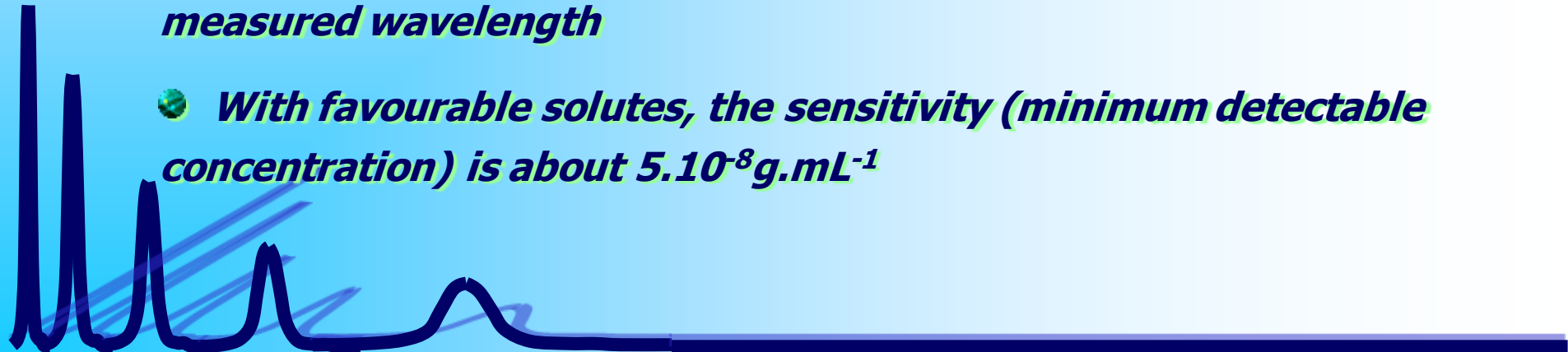
HPLC detectors are commonly classified as:

- **general (or bulk property detector)**: it measures the difference in some physical property between the chromatographic effluent and the pure mobile phase. This kind of detector, such as **refractive index detector**, is considered as **universal** and has a general use but a low sensitivity and a limited dynamic range
- **specific (or solute property detector)**: responds to a physical or chemical property characteristic of the solute. It provides high sensitivity with a wide linear range such as in **spectrophotometric detectors**. Although this kind of detectors is **specific** it is more widely used than bulk property ones



Spectrophotometric UV/visible detector

- *It is a simple and cheap detector*
- *and the most widely used detector in HPLC*
- *It is a solute property detector*
- *It is specific to solutes which exhibit an absorption in the UV/visible range (generally unsaturated compounds)*
- *The mobile phase must not significantly absorb at the measured wavelength*
- *With favourable solutes, the sensitivity (minimum detectable concentration) is about $5 \cdot 10^{-8} \text{g.mL}^{-1}$*



UV detector (with photodiode array)

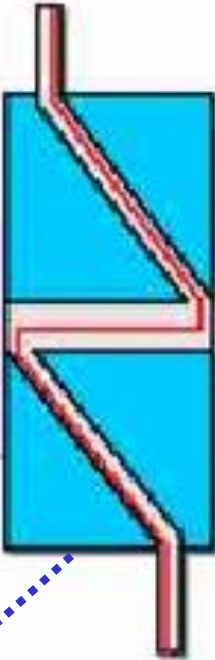
deuterium lamp



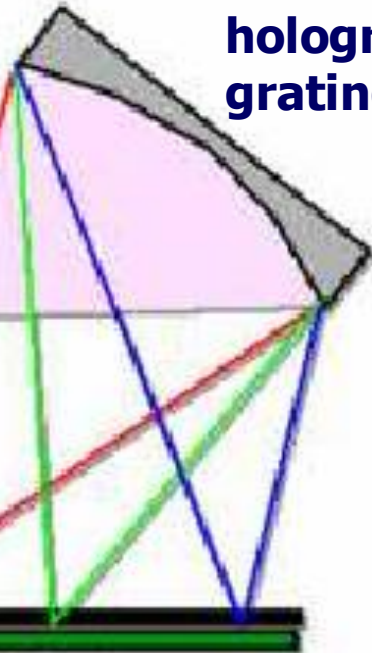
quartz lens



sample cell



holographic grating



photodiode array

