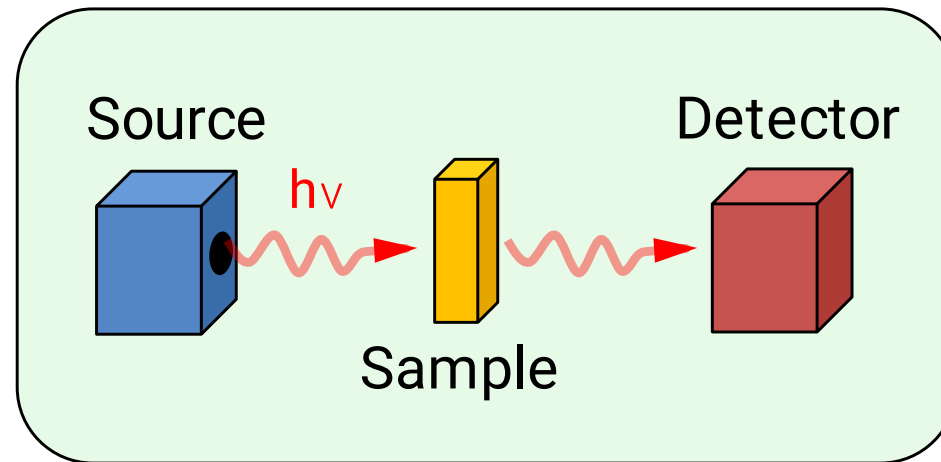
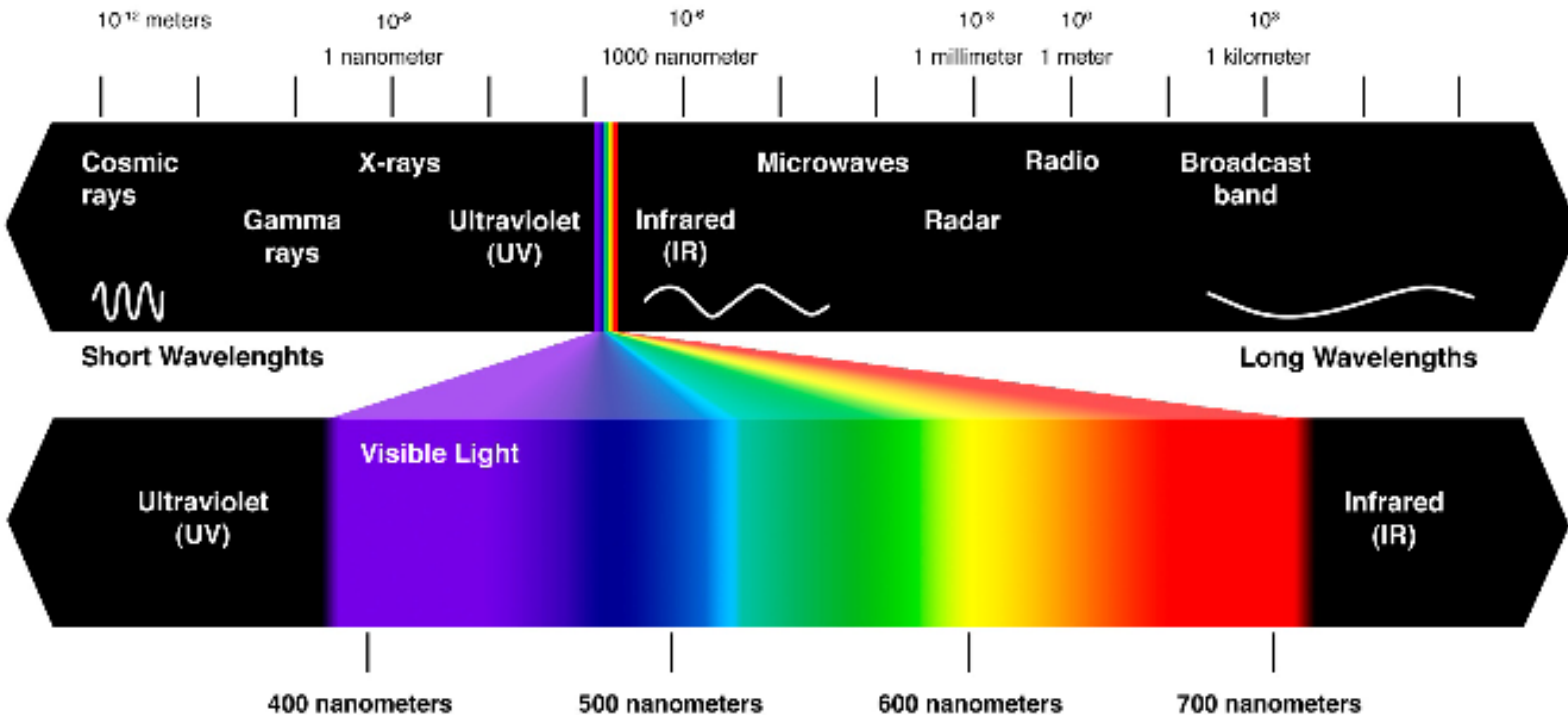
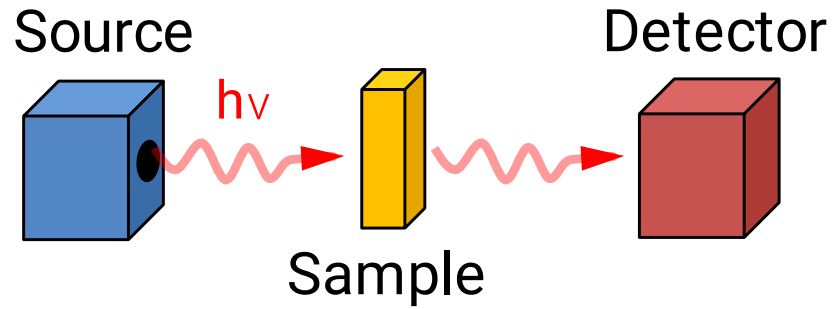


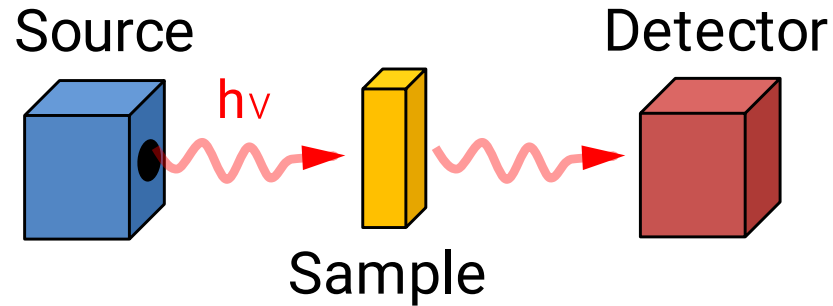
Absorption Spectroscopy



Absorption Spectroscopy



Why Absorption Spectroscopy?



- Color is ubiquitous to humans
- 1000 x more sensitive than NMR
- Qualitative technique (what is in the solution)
- Quantitative technique (concentrations, ratios, etc.)
- Its easy
- It is inexpensive
- Numerous applications

Absorption Spectroscopy in Action

HPLC

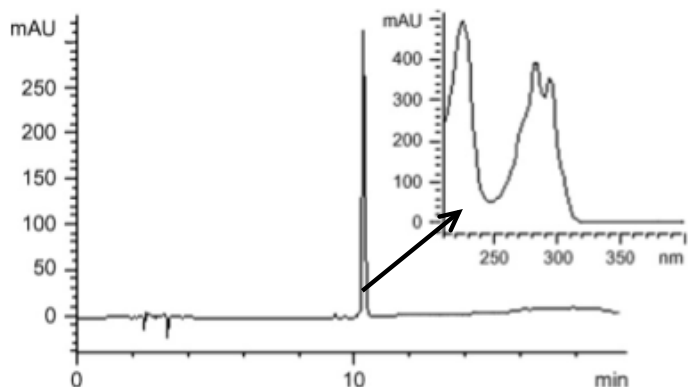
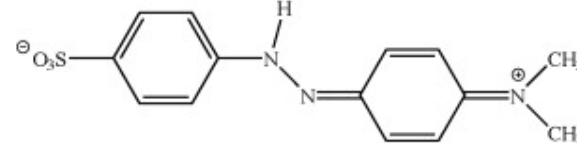
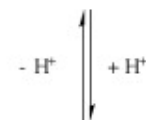
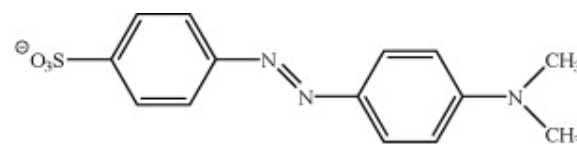


Fig. 3. HPLC chromatogram and UV-vis spectra (inset) of Bromo-Dragonfly. The HPLC trace was detected at 210 nm.

pK_a Determination

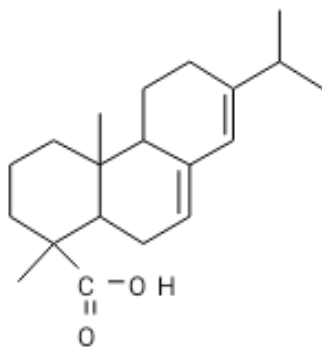
Yellow (pH > 4.4)



Red (pH > 3.2)

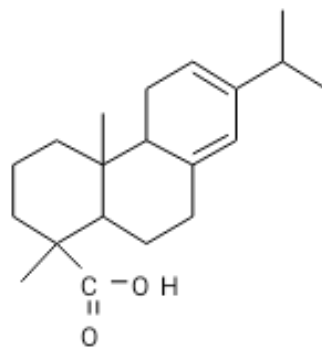
Structure Differentiation

Abietic Acid



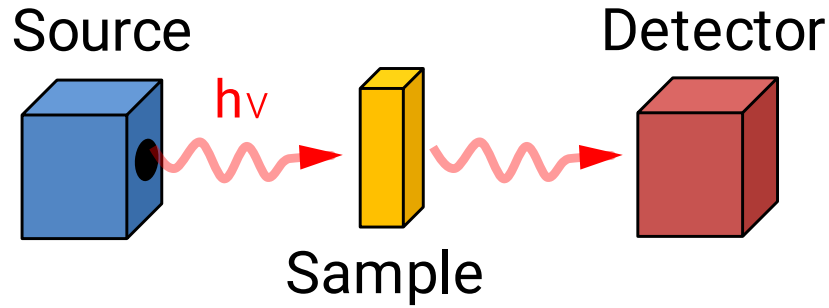
214 nm

Levopimaric acid

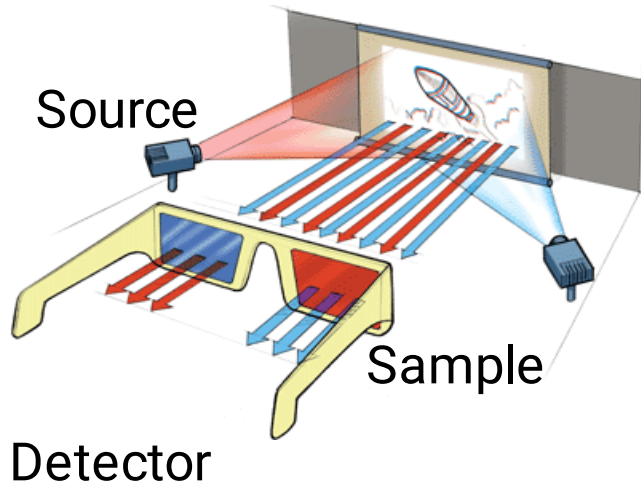


253 nm

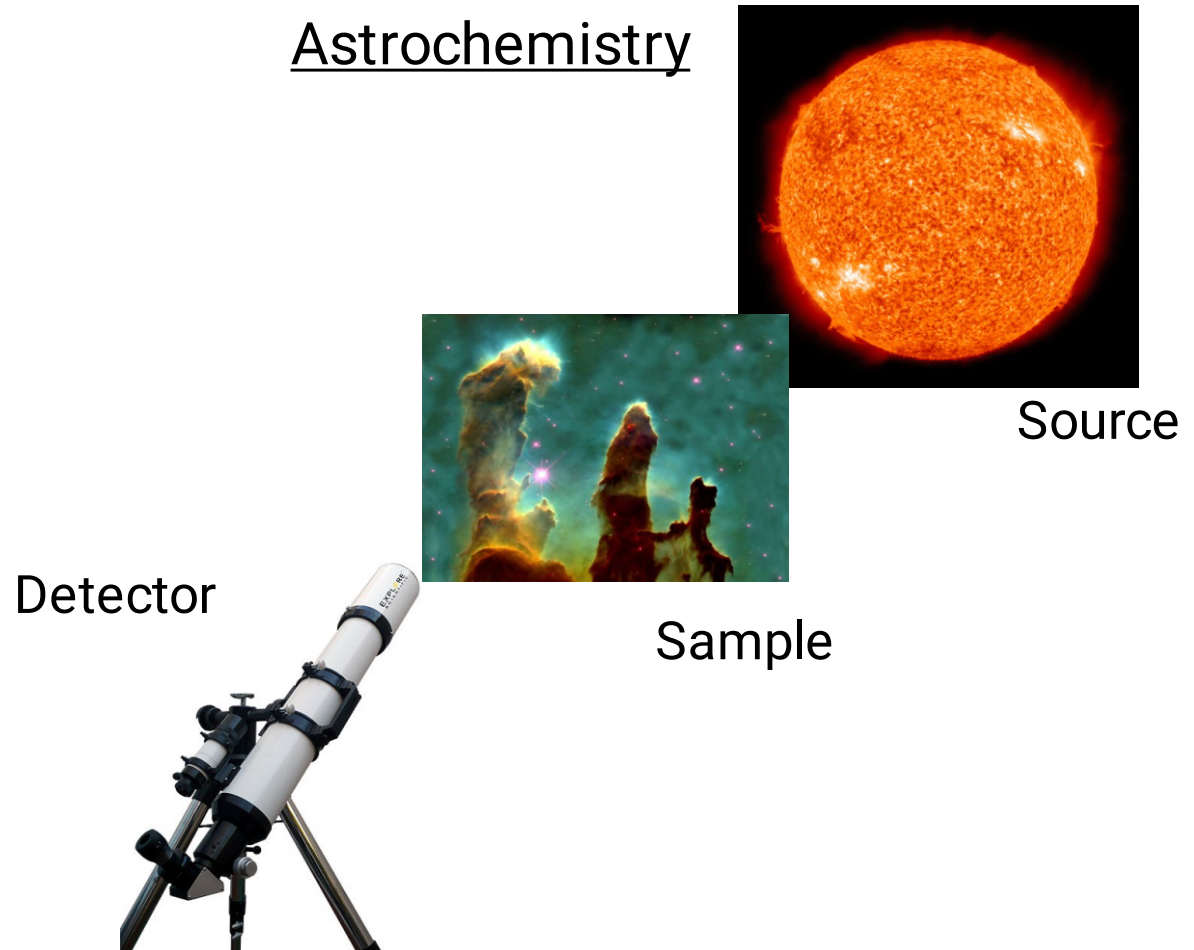
Examples



3D Glasses



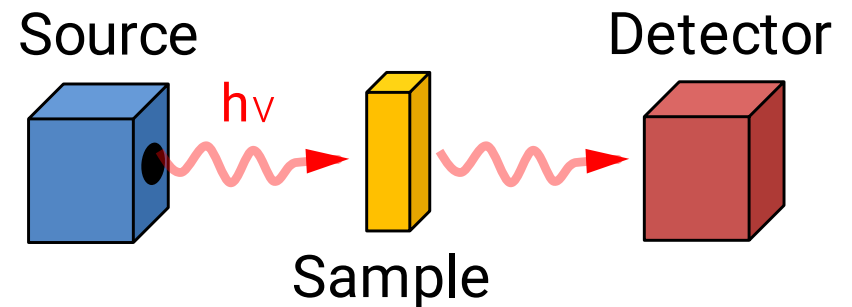
Astrochemistry



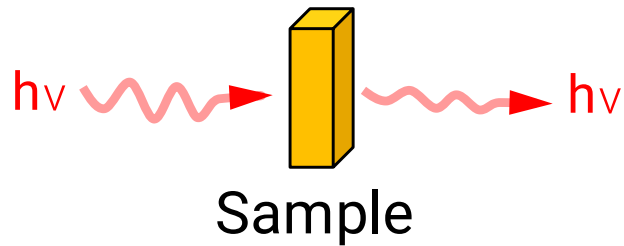
First homework (not really): Think of examples of absorption spectroscopy

Outline

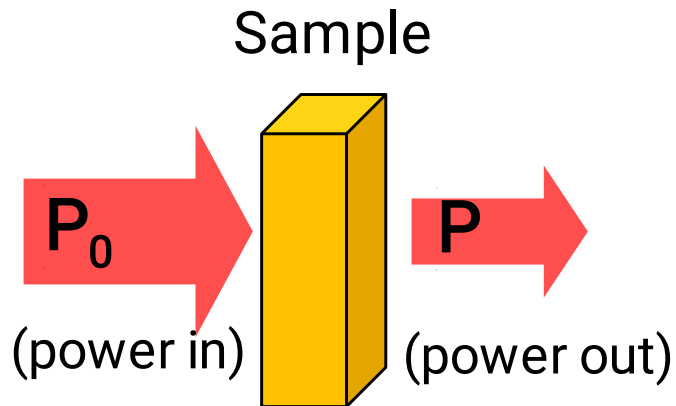
- 1) Absorption
- 2) Spectrum Beer's Law
- 3) Instrument Components
 - Light sources
 - Monochrometers
 - Detectors
 - Other components
 - The sample
- 4) Instrument Architectures
- 5) UV-Vis in Action
- 6) Potential Complications



Absorption by the Numbers



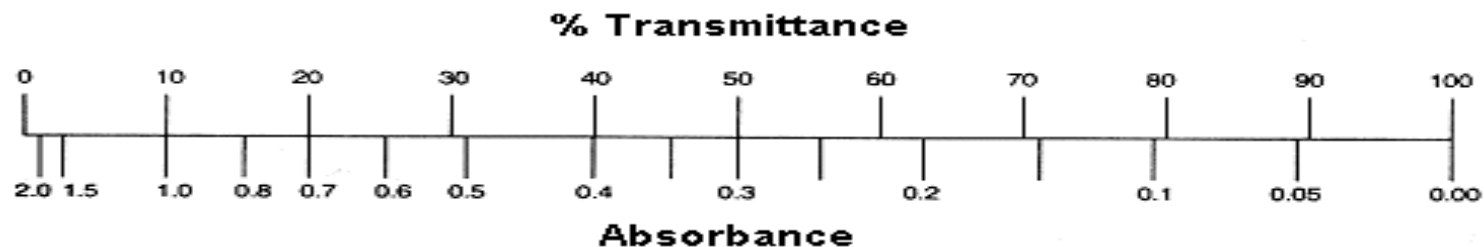
We don't measure absorbance. We measure transmittance.



- Transmittance:

- $T = P/P_0$: Absorbance:

$$A = -\log T = \log P_0/P$$



Beer's Law

The Beer-Lambert Law (λ specific):

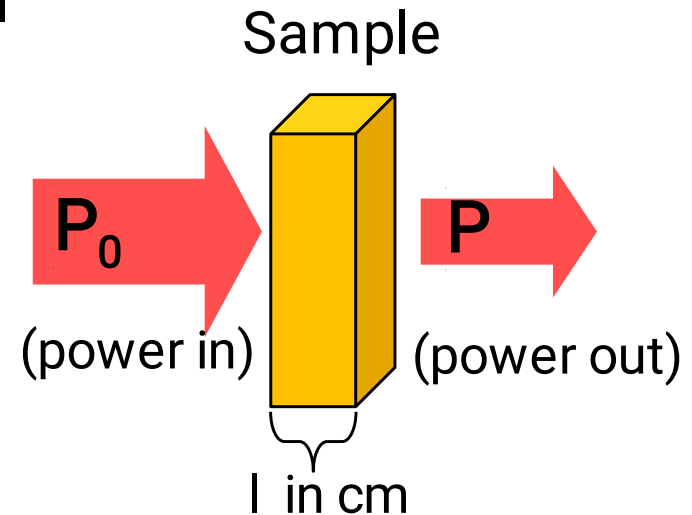
$$A = \epsilon c l$$

A = absorbance (unitless, $A = \log_{10} P_0/P$)

ϵ = molar absorptivity ($L \text{ mol}^{-1} \text{ cm}^{-1}$)

l = path length of the sample (cm)

c = concentration (mol/L or M)



Concentration \uparrow

Path length \uparrow

Molar Abs. \uparrow

Absorbanc \uparrow

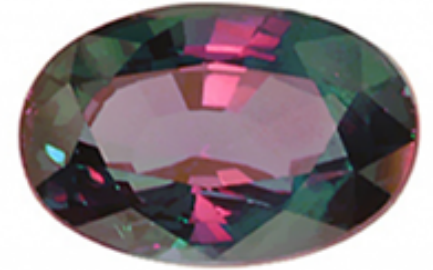
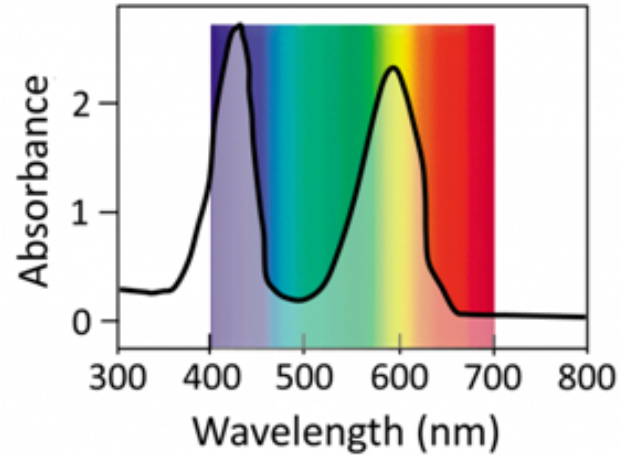
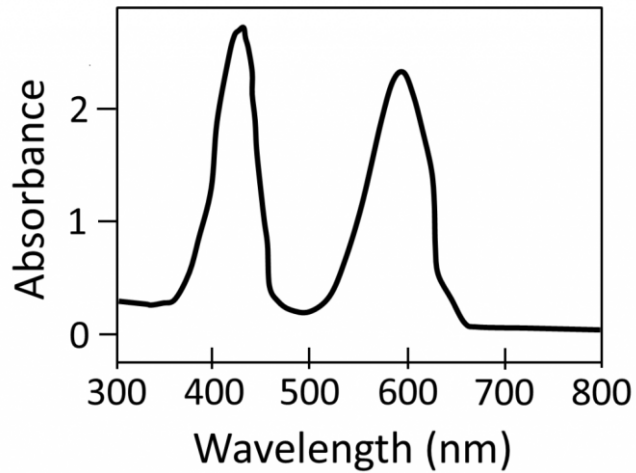
ϵ Absorbanc \uparrow

ϵ Absorbanc \uparrow

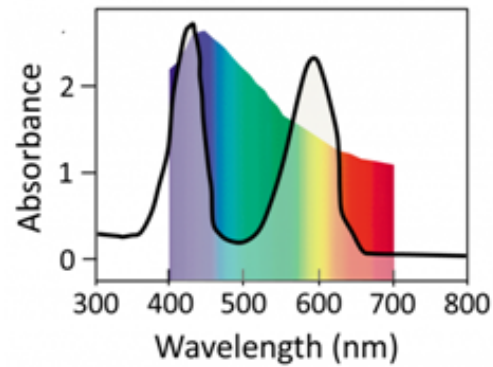
ϵ

Absorption Spectrum

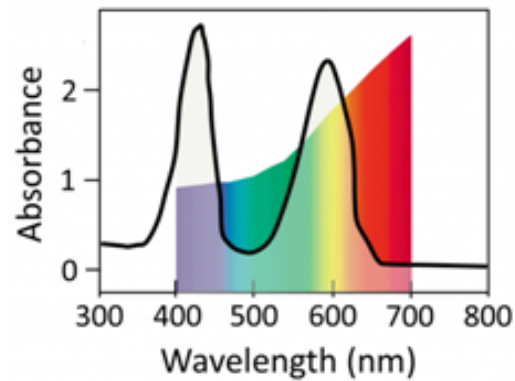
Alexandrite Gemstone
 BeAl_2O_4 (+ Cr^{3+} doping)



Sunlight



Candle/
Incandescent



Beer's Law

The Beer-Lambert Law:

A = absorbance (unitless, $A = \log_{10} P_0/P$)

ϵ = molar absorptivity ($\text{L mol}^{-1} \text{cm}^{-1}$)

l = path length of the sample (cm)

c = concentration (mol/L or M)

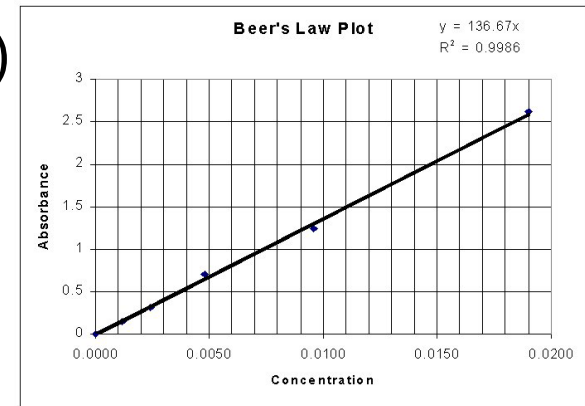
$$A = \epsilon c l$$

Find ϵ

- 1) Make a solution of known concentration (C)
- 2) Put in a cell of known length (l)
- 3) Measure A by UV-Vis
- 4) Calculate ϵ

$$A = \epsilon c l$$

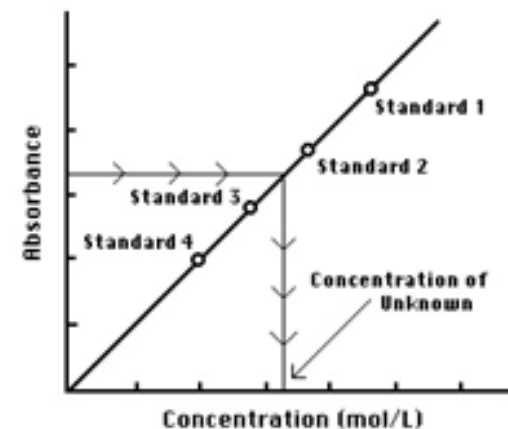
$$y = m x + b$$



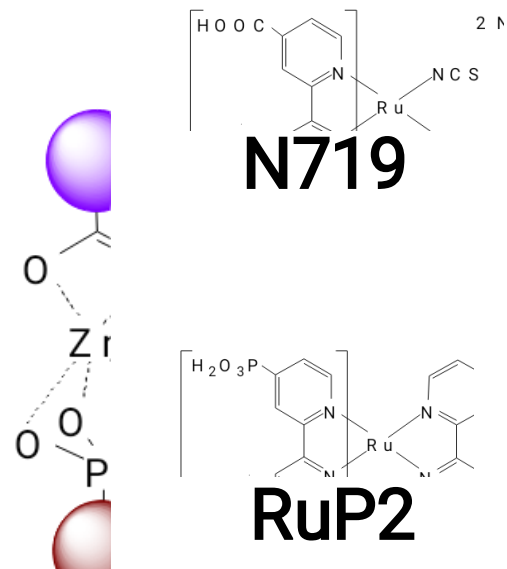
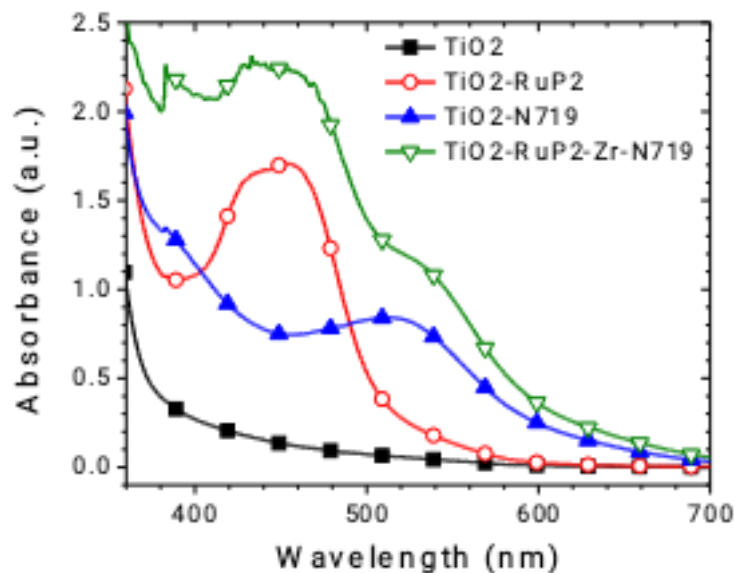
Find Concentrations

- 1) Know ϵ
- 2) Put sample in a cell of known length (l)
- 3) Measure A by UV-Vis
- 4) Calculate C

$$A = \epsilon c l$$



Beer's Law Applied to Mixtures



$$A_1 = \epsilon_1 c_1 l$$

$$A_{\text{total}} = A_1 + A_2 + A_3 \dots$$

$$A_{\text{total}} = \epsilon_1 c_1 l + \epsilon_2 c_2 l + \epsilon_3 c_3 l$$

$$A_{\text{total}} = l(\epsilon_1 c_1 + \epsilon_2 c_2 + \epsilon_3 c_3)$$

Limitations to Beer's Law

The Beer-Lambert Law:

$$A = \epsilon c l$$

Reflection/Scattering Loss

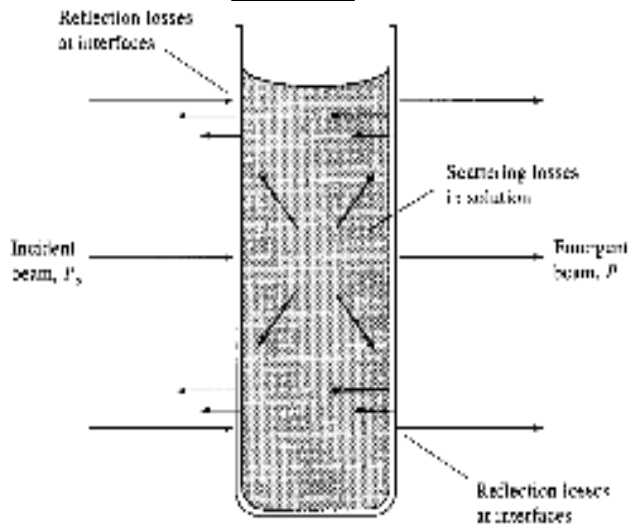


Figure 13-1 Reflection and scattering losses.

Reflection/Scattering

- Air bubbles
- Aggregates

Lamp effects

- Temperature (line broadening)
- Light source changes
- Solvent lensing

Absorbance too high (above 2)

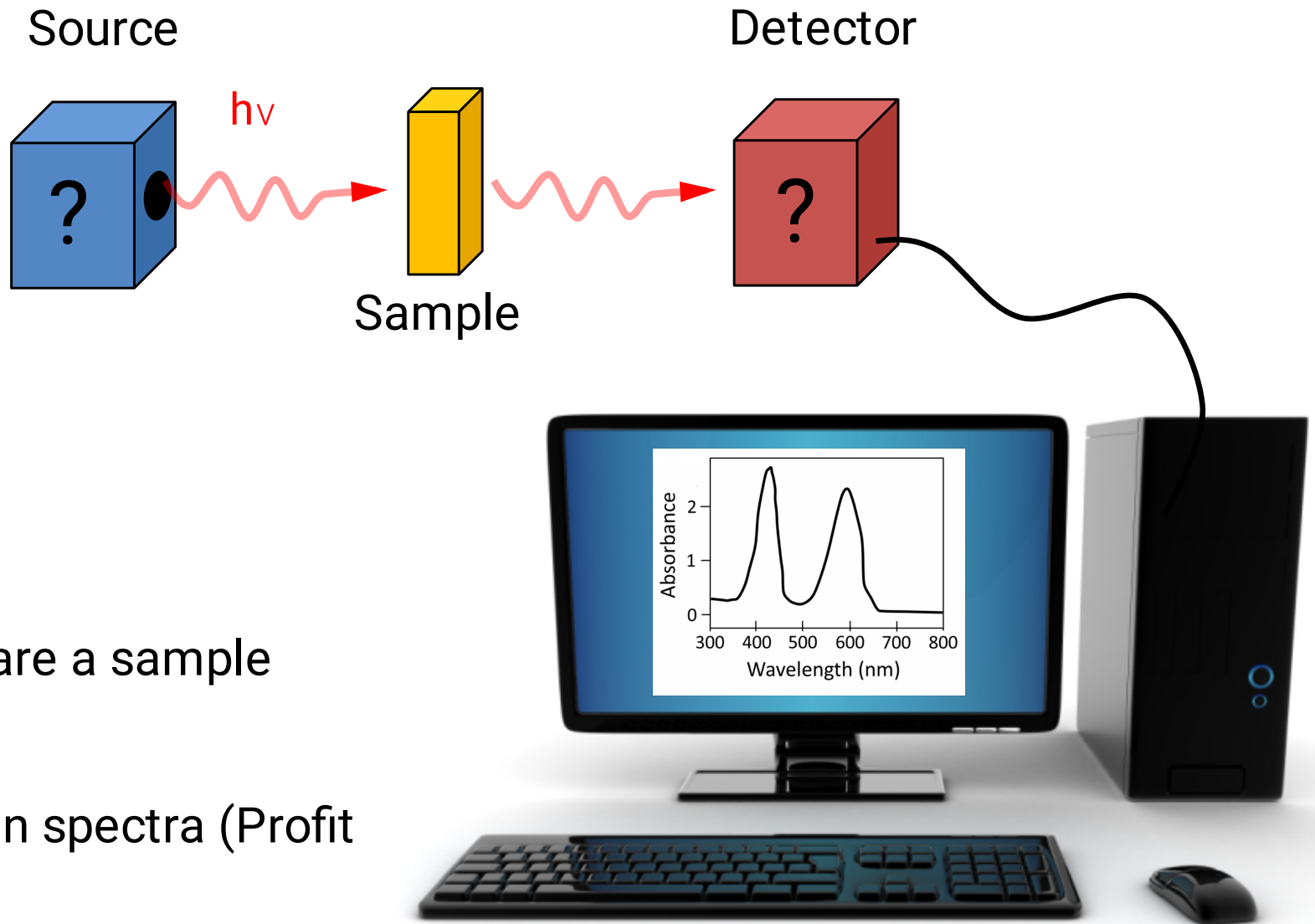
- Local environment effects
- Dimerization
- Refractive index change (ionic strength)

Sample changes

- Photoreaction/decomposition
- Side of the cuvette
- Hydrogen bonding
- Non-uniform through length

$$A = -\log T = \log \frac{P_0}{P}$$

Absorption Spectroscopy



Procedure

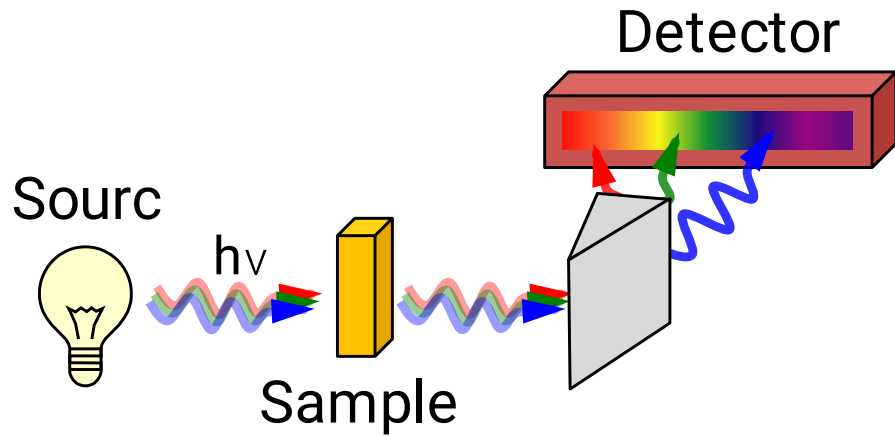
Step 1: Prepare a sample

Step 2: ???

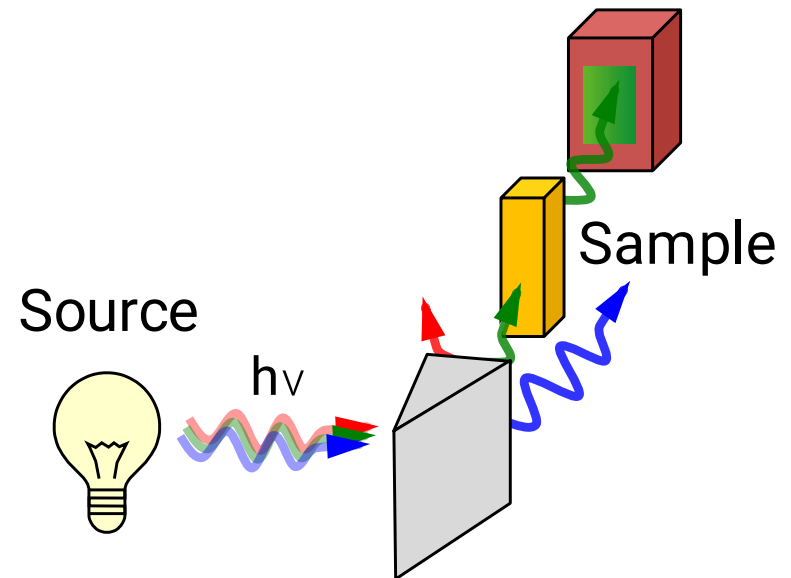
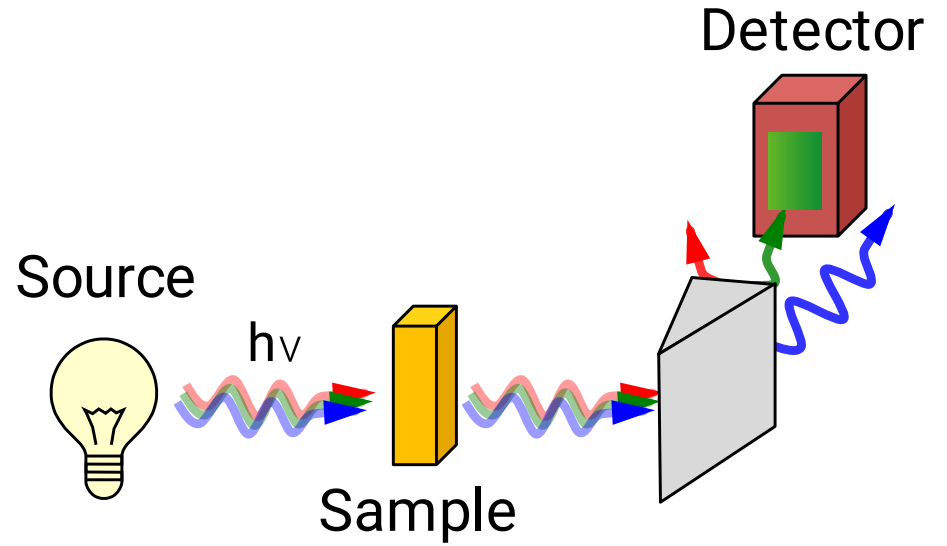
Step 3: Obtain spectra (Profit !)

Instrumentation

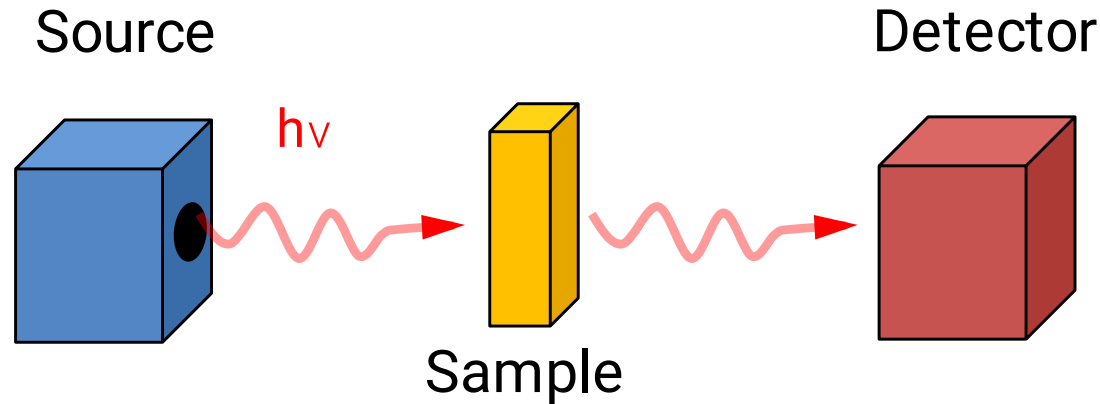
Full spectra detection



Single λ detection



Instrumentation



Full spectra detection

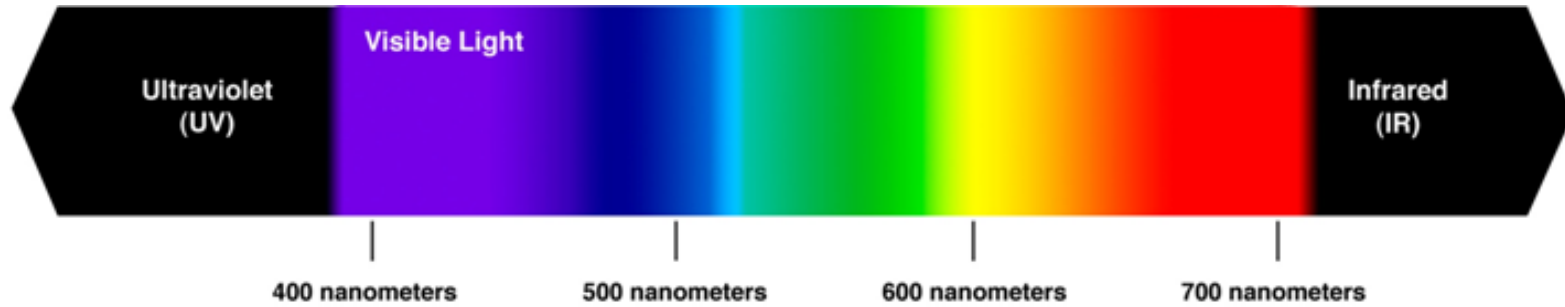
- Source
- Sample
- Monochrometer
- Area detector

Single λ detection

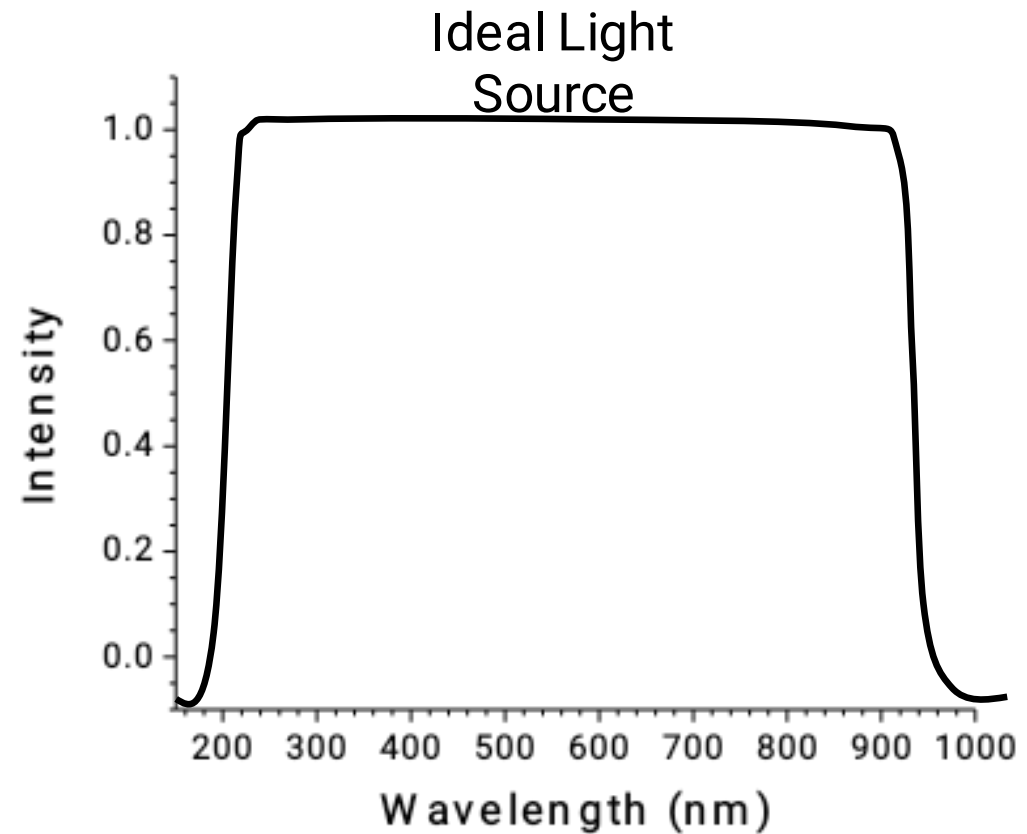
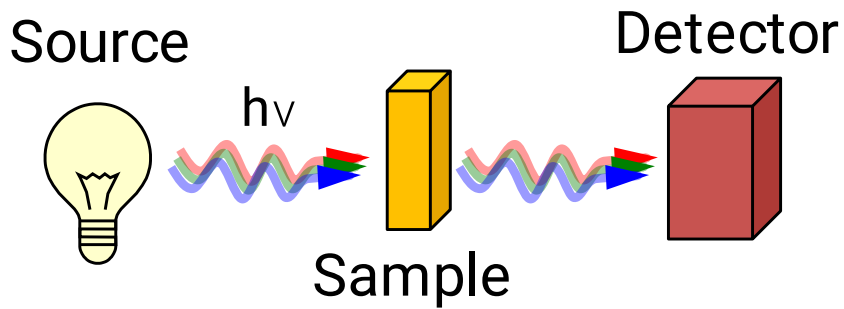
- Source
- Monochrometer
- Sample
- “Point” detector

1. Light sources
2. Monochrometer
3. Detectors
4. Samples

Light Sources, Ideal

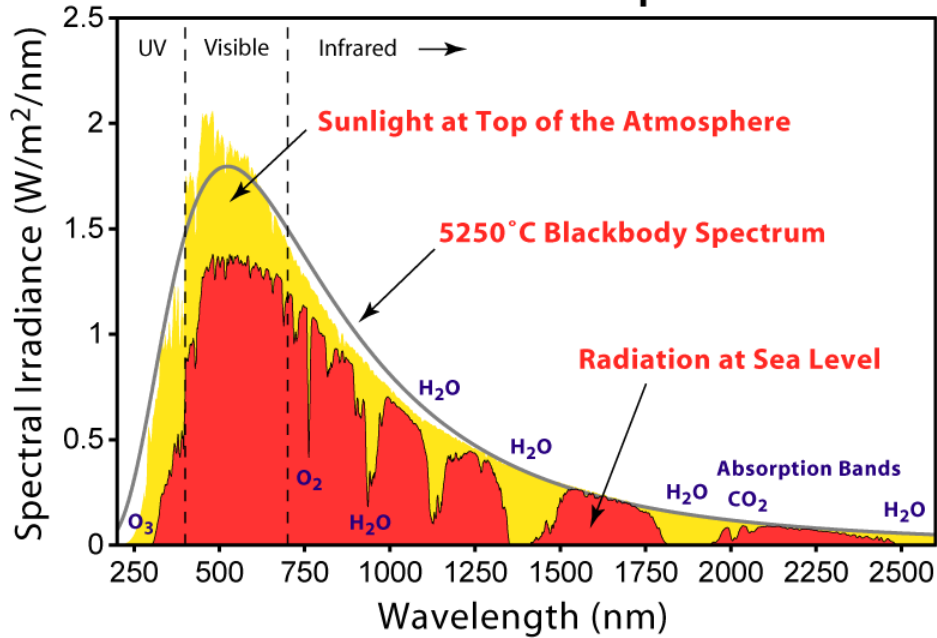


Experimentally we would like ~200 – 900 nm



Light Sources: The Sun

Solar Radiation Spectrum



Pros:

It's free!

Does not die

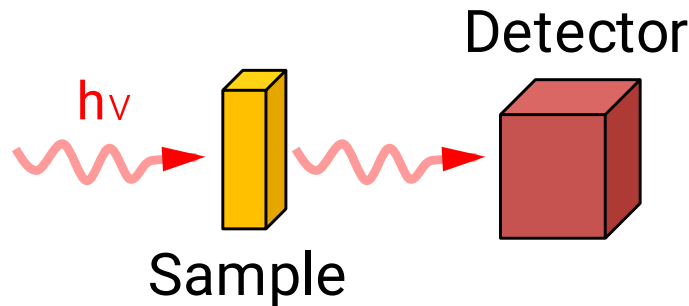
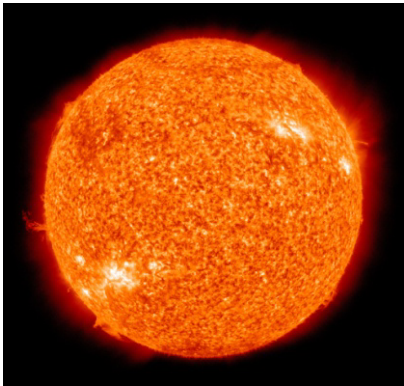
Relatively uniform from 400-800 nm

Cons:

Inconsistent

Minimal UV-light

Intense absorption lines



Light Sources: Xe Lamp

Electricity through Xe gas



Pros:

Mimics the sun (solar simulator)

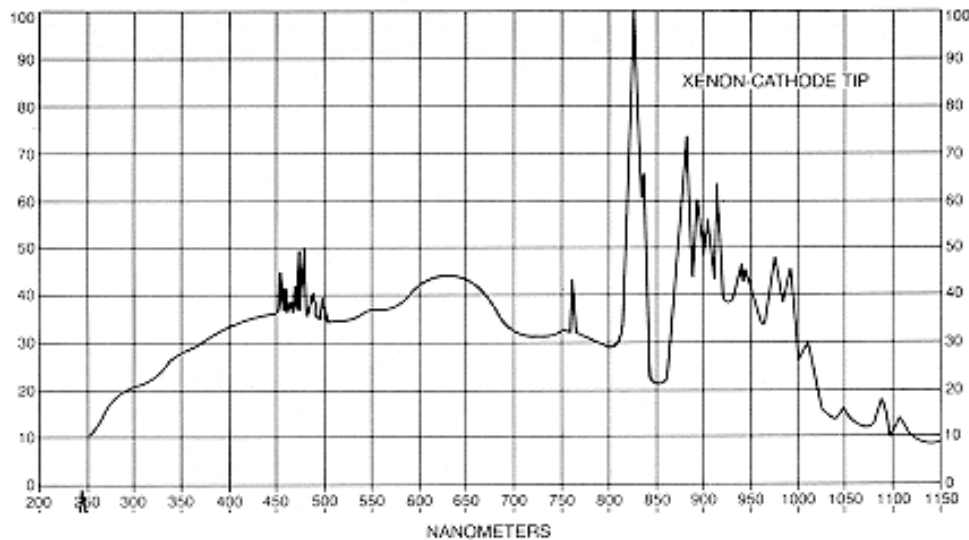
It's simple

Cons:

Relatively Expensive

Minimal UV-light (<300 nm)

Potential Instability



Sources of Instability in Xenon Arc Discharge Lamps

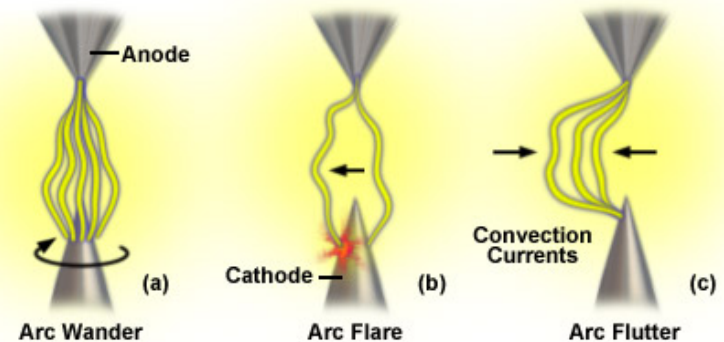
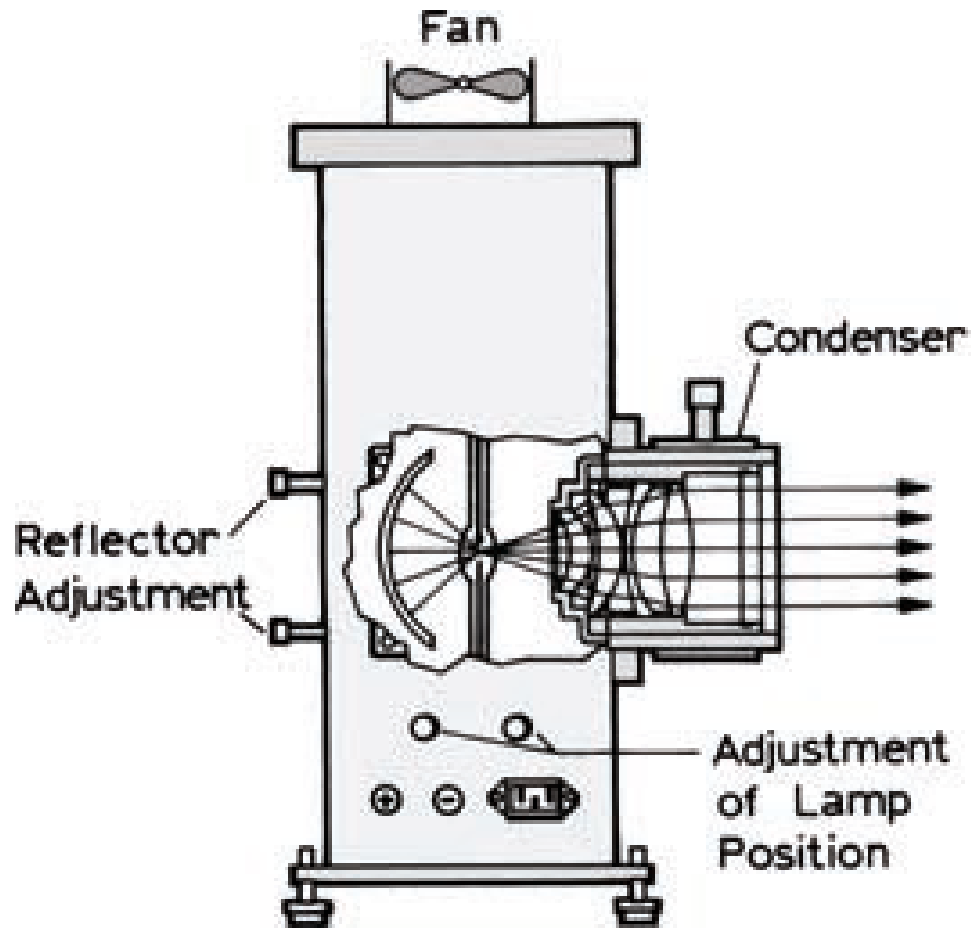
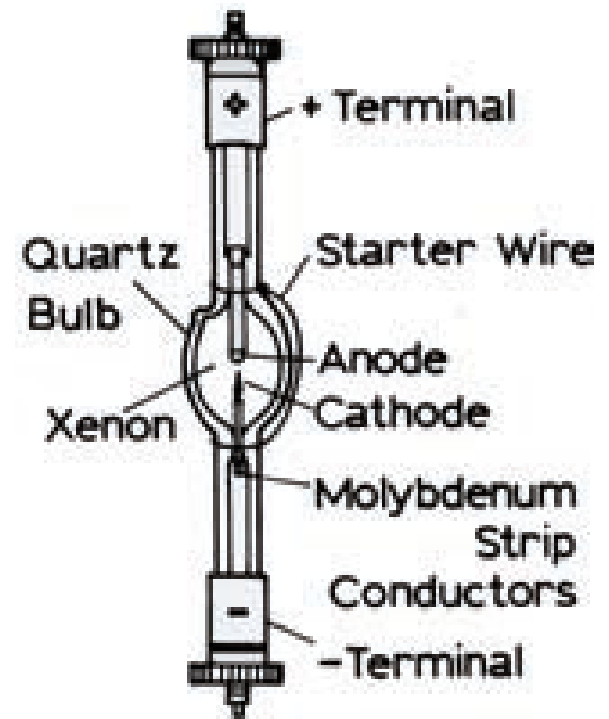


Figure 3

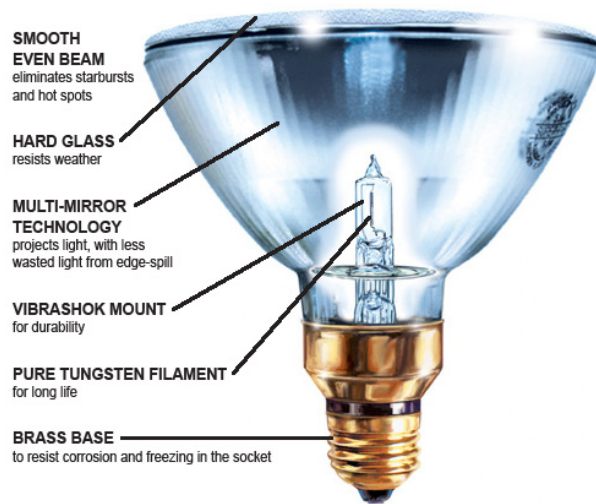
Light Sources: Xe Lamp



Xenon Arc Lamp



Light Sources: Tungsten Halogen Lamp



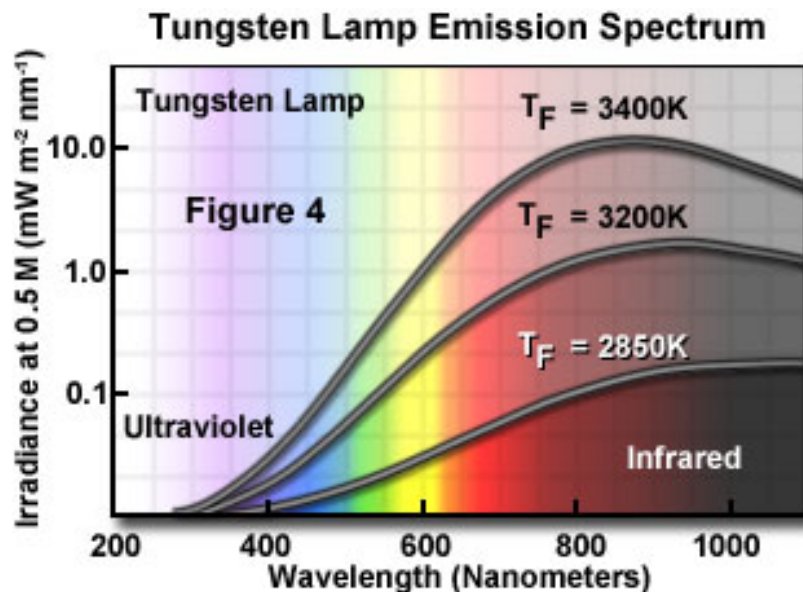
Pros:

- Compact size
- High intensity
- Low cost
- Long lifetime
- Fast turn on
- Stable

Halogen gas and the tungsten filament
Higher pressure (7-8 ATM)

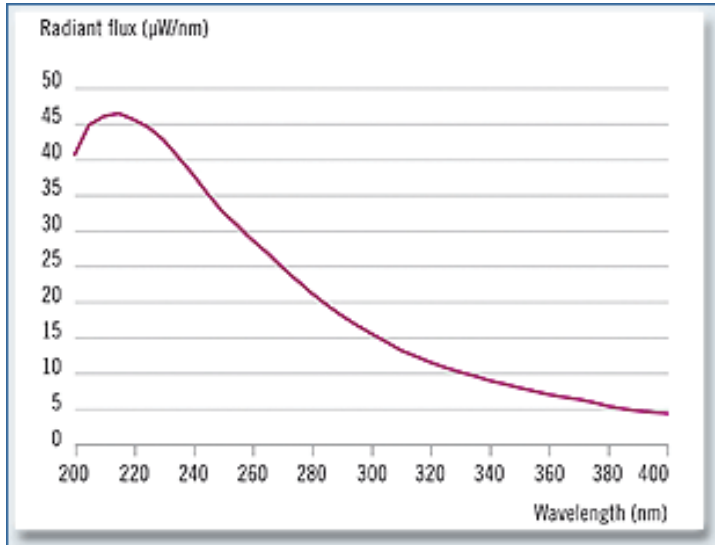
Cons:

- Very hot
- Bulb can explode
- Minimal UV-light (<300 nm)



“White” Light

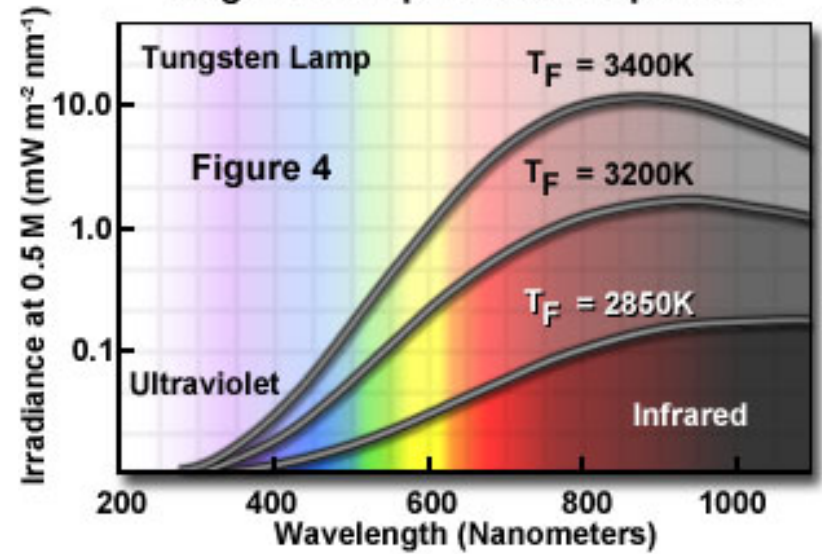
Deuterium lamp



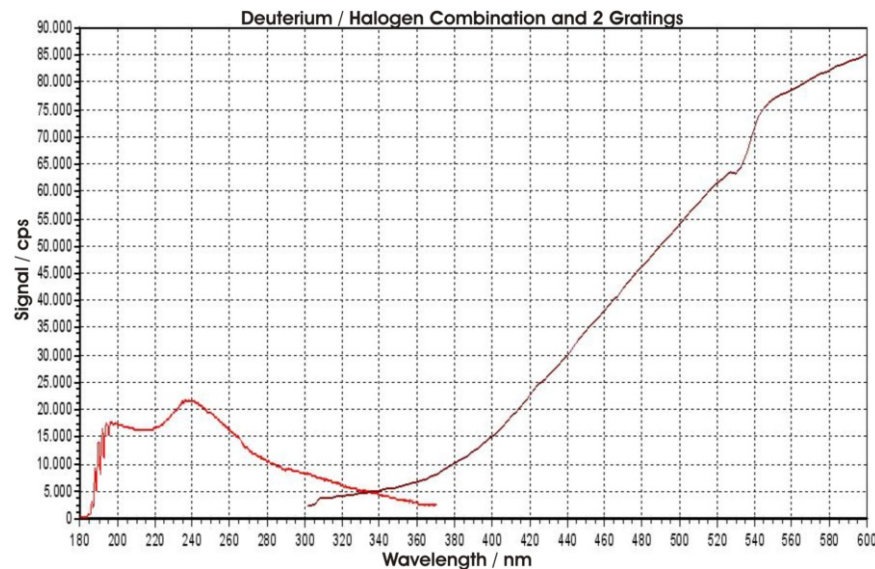
Deuterium lamp – 200-330

+

Tungsten Lamp Emission Spectrum

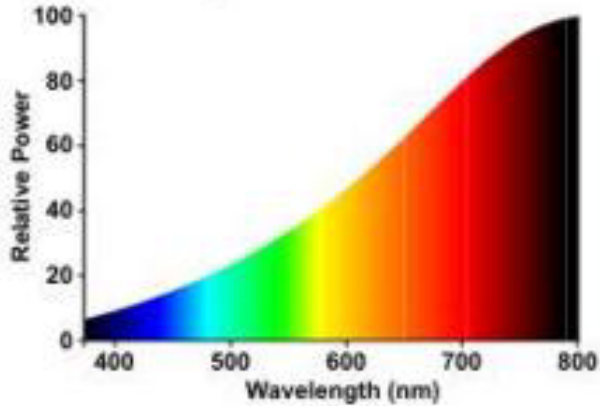


Tungsten lamp – >300 nm

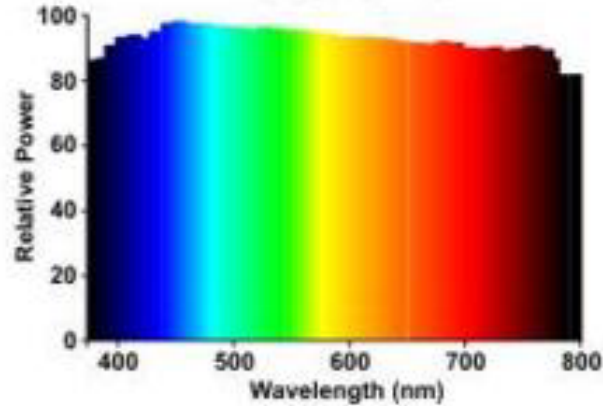


Other Light Sources

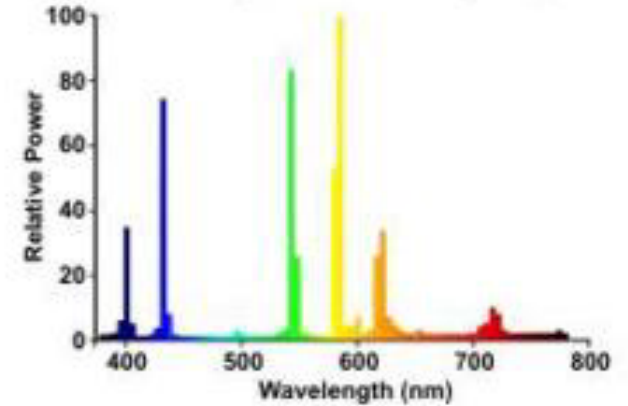
Tungsten Incandescent



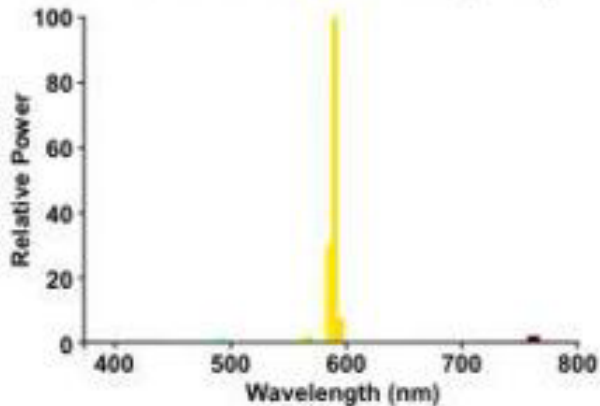
Daylight (D65)



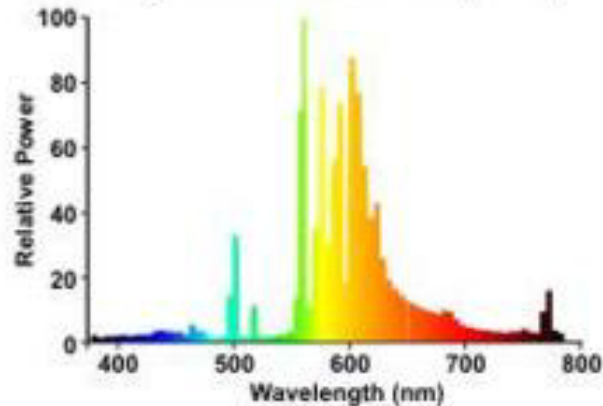
Mercury Fluorescent (MBF)



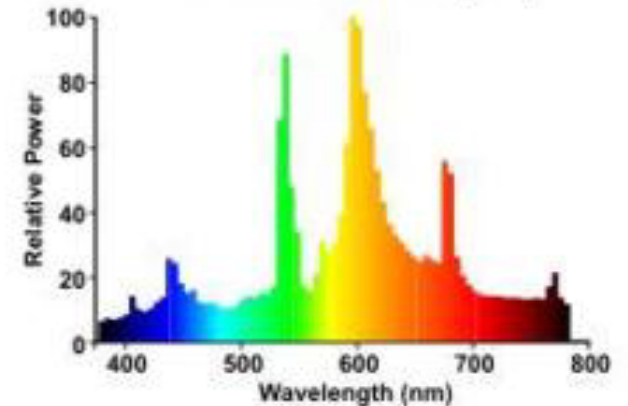
Low Pressure Sodium (SOX)



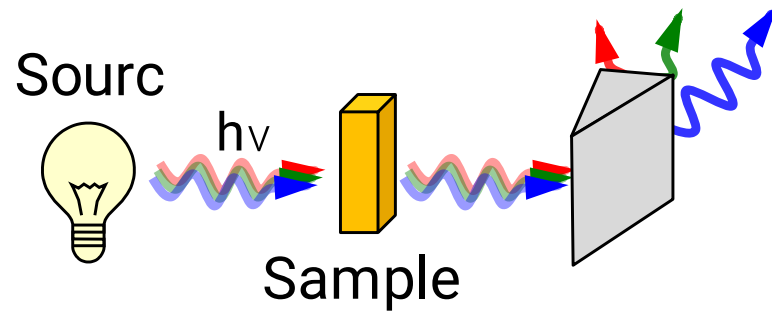
High Pressure Sodium (SON)



Metal Halide 3000K (MBI)



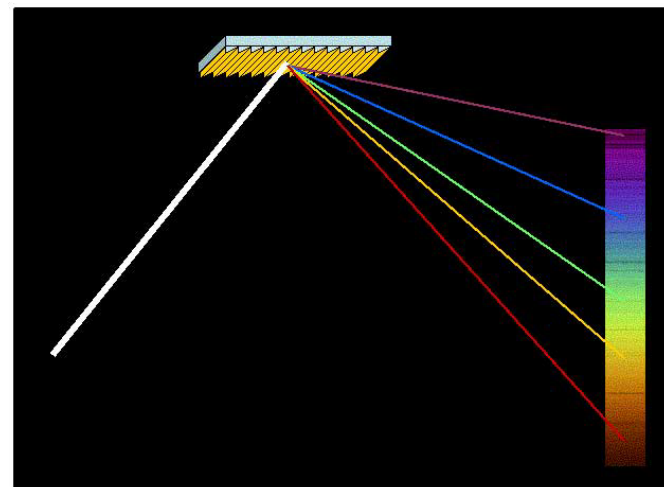
Separating the Light



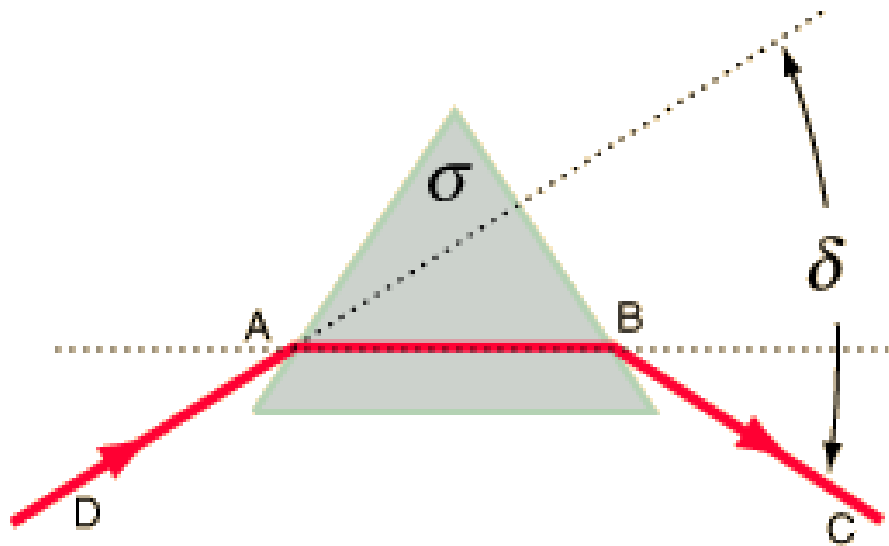
Prism



Grating



Monochromator: Prism



$$\frac{n_{\text{prism}}}{n_0} = \frac{\sin \frac{1}{2} (\sigma + \delta)}{\sin \frac{1}{2} \sigma}$$

n_0 = refractive index of air

n_{prism} = refractive index of prism

σ = prism apex angle

δ = deviation angle

n_0 is constant
 σ is constant
 n_{prism} is λ dependent

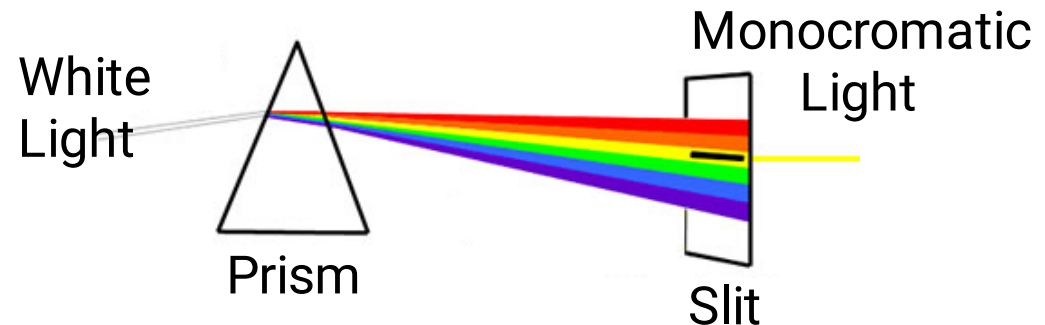
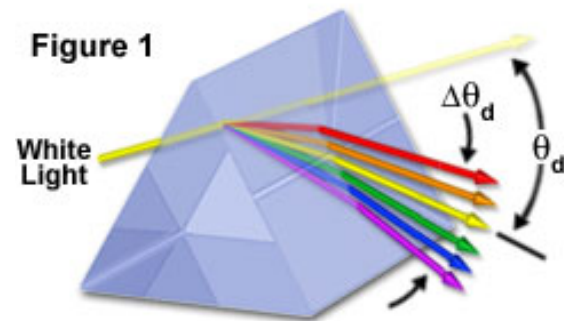
Wavelength
 h



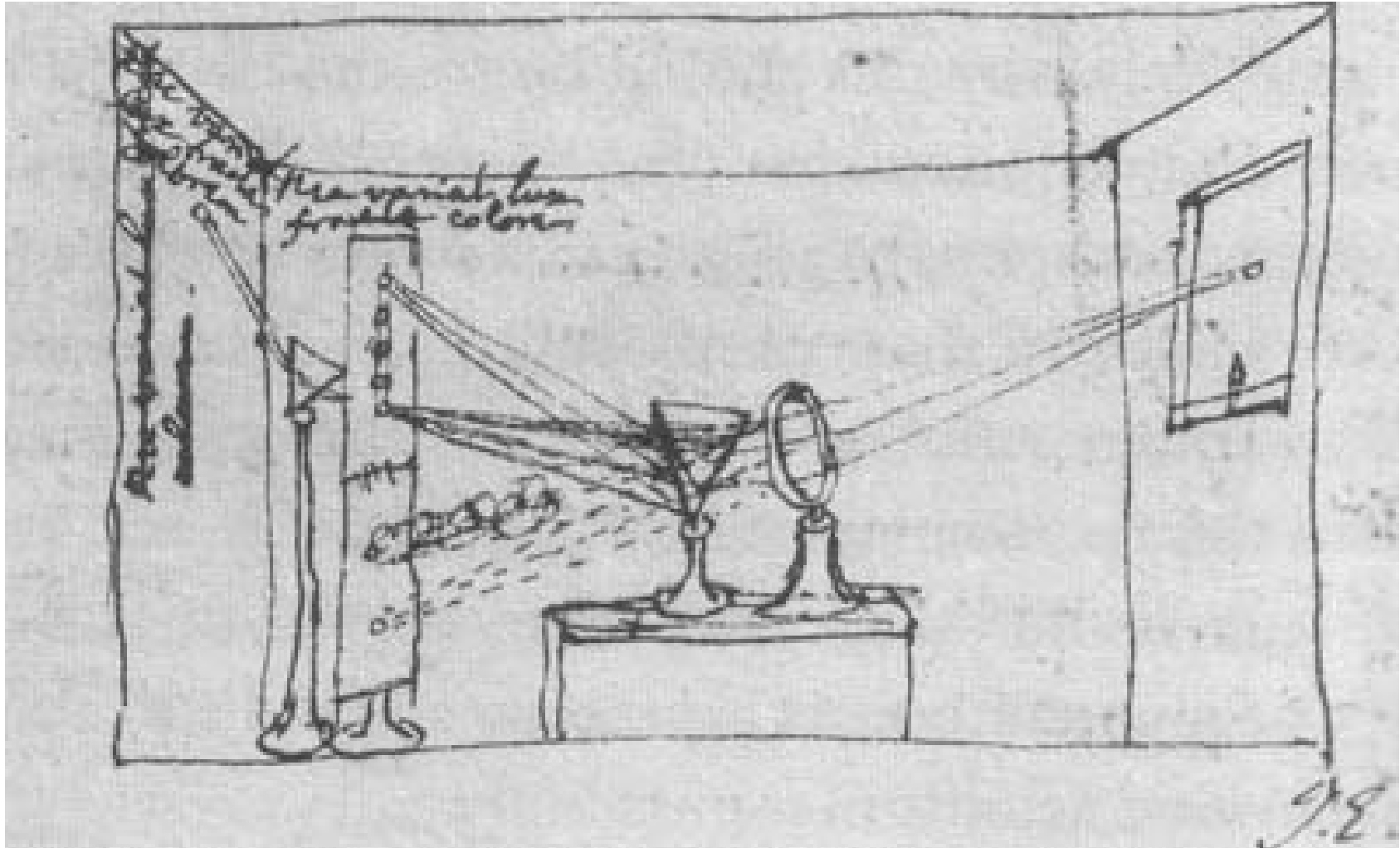
Deviation



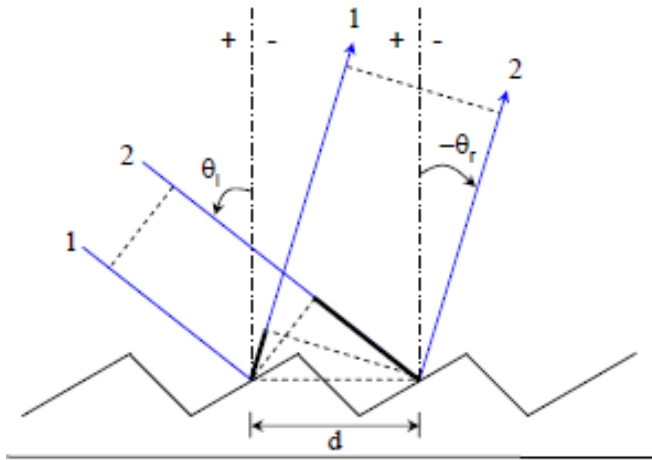
Equilateral Dispersing Prism



Monochromator: Prism



Monochromator: Grating



d is constant
 θ_i is constant
 θ_r is λ dependent

Wavelength
 h



Diffraction



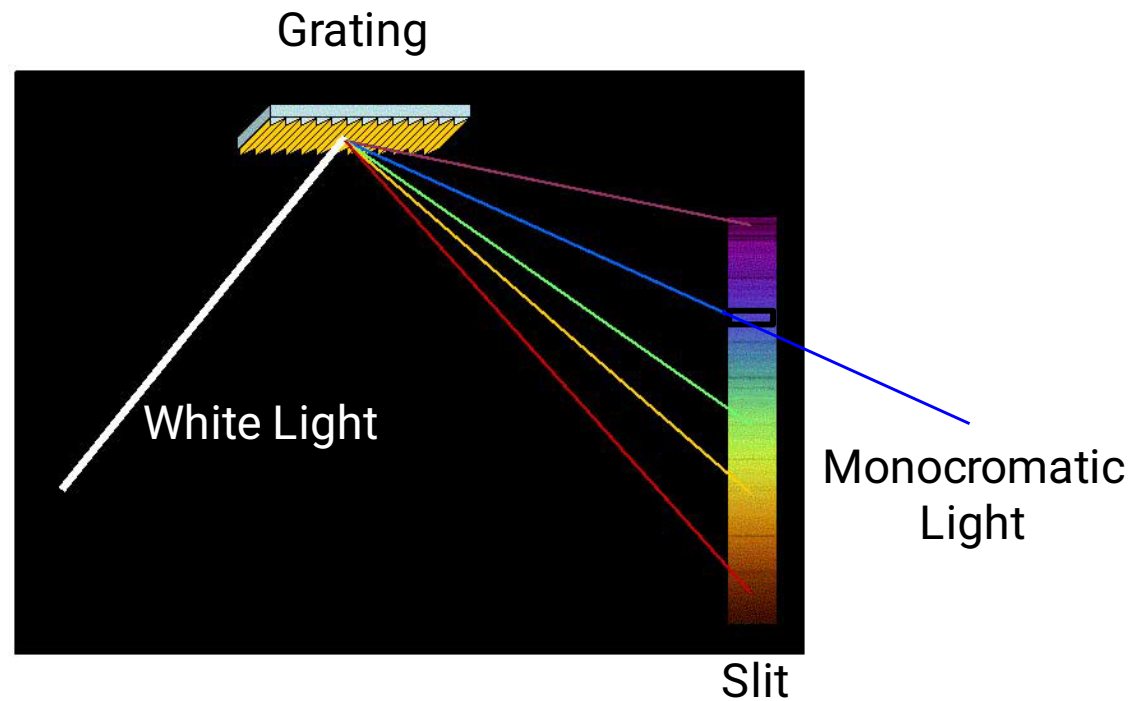
$$\lambda = 2d(\sin \theta_i + \sin \theta_r)$$

λ = wavelength

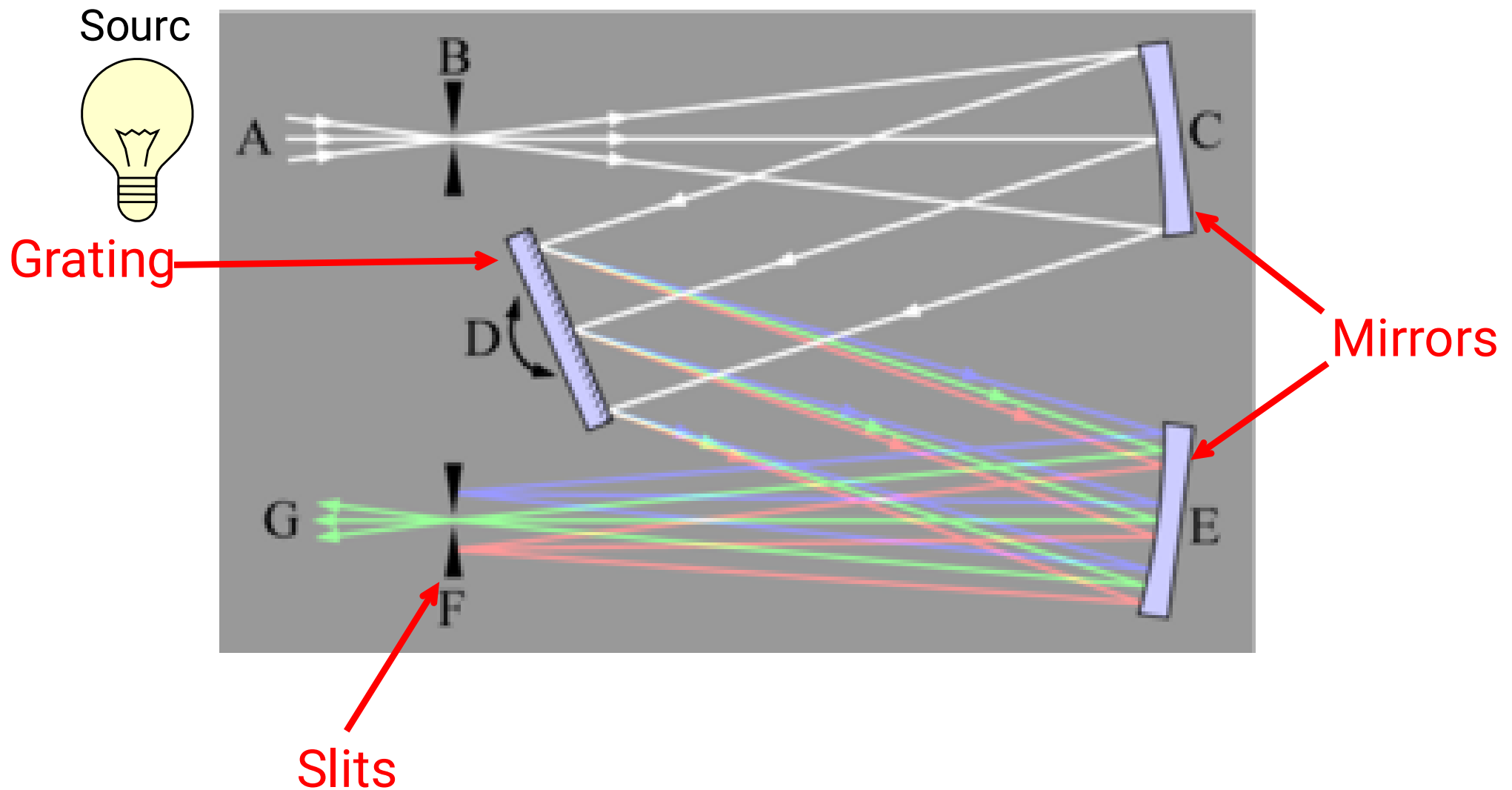
d = grating spacing

θ_i = incident angle

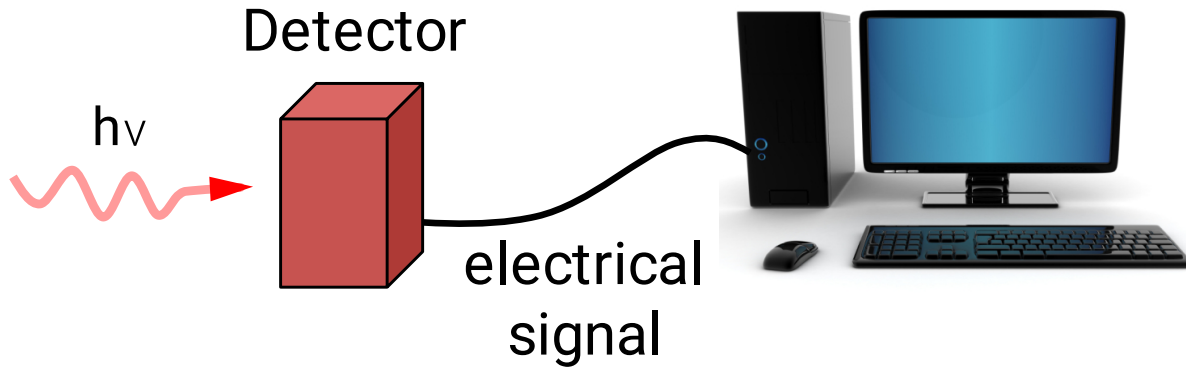
θ_r = diffracted angle



Monochromator: Grating



Detectors



- high sensitivity
- high signal/noise
- constant response for λ s
- fast response time

Single λ detection

Diode

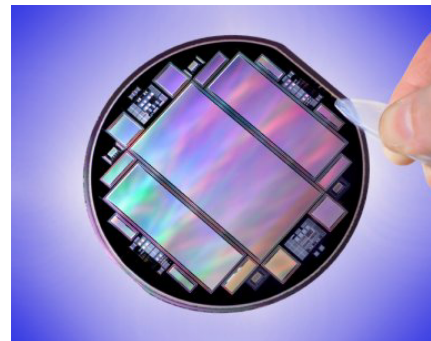


PMT

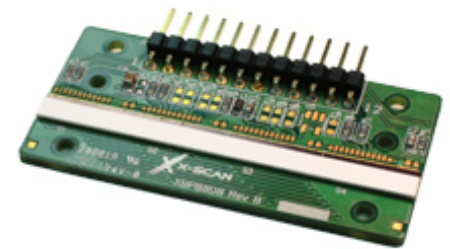


Full spectra detection

CCD



Diode Array



Detectors: Diode

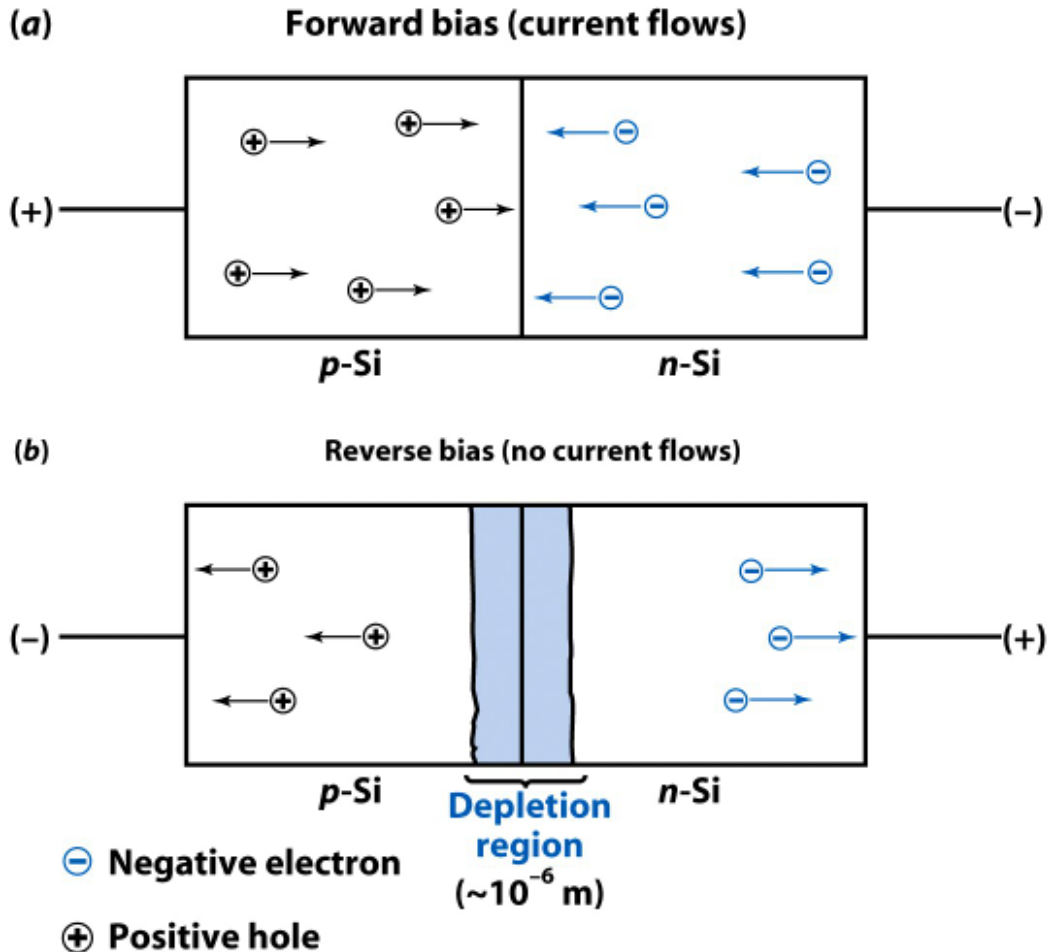


Figure 15-27
Quantitative Chemical Analysis, Seventh Edition
© 2007 W. H. Freeman and Company

Forward Bias:

Apply a positive potential
holes + e^- = exciton = light
Light Emitting Diode

Zero Bias:

Apply 0 potential
exciton = holes + e^- = current
silicon solar cell

Negative Bias:

Apply a negative potential
exciton = holes + e^- = more
current
photodetector

n-type (extra electrons)- P or As
doped

p-type (extra holes)- Al or B doped

Detectors: Diode

Pros:

Long Lifetime

Small/Compact

Inexpensive

Linear response

190-1000 nm

Cons:

No wavelength discrimination

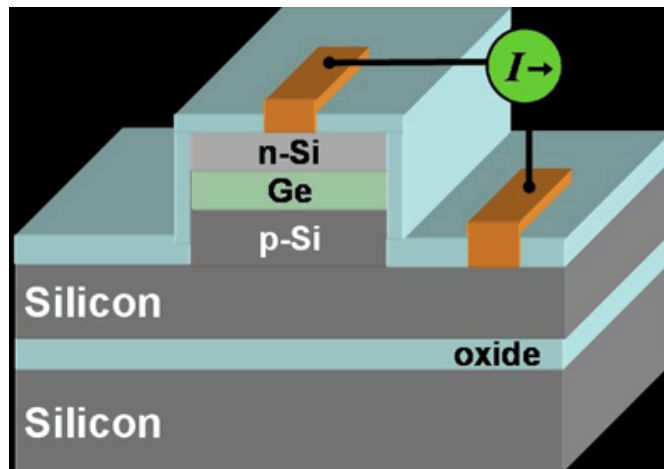
Minimal internal gain

Much lower sensitivity

Small active area

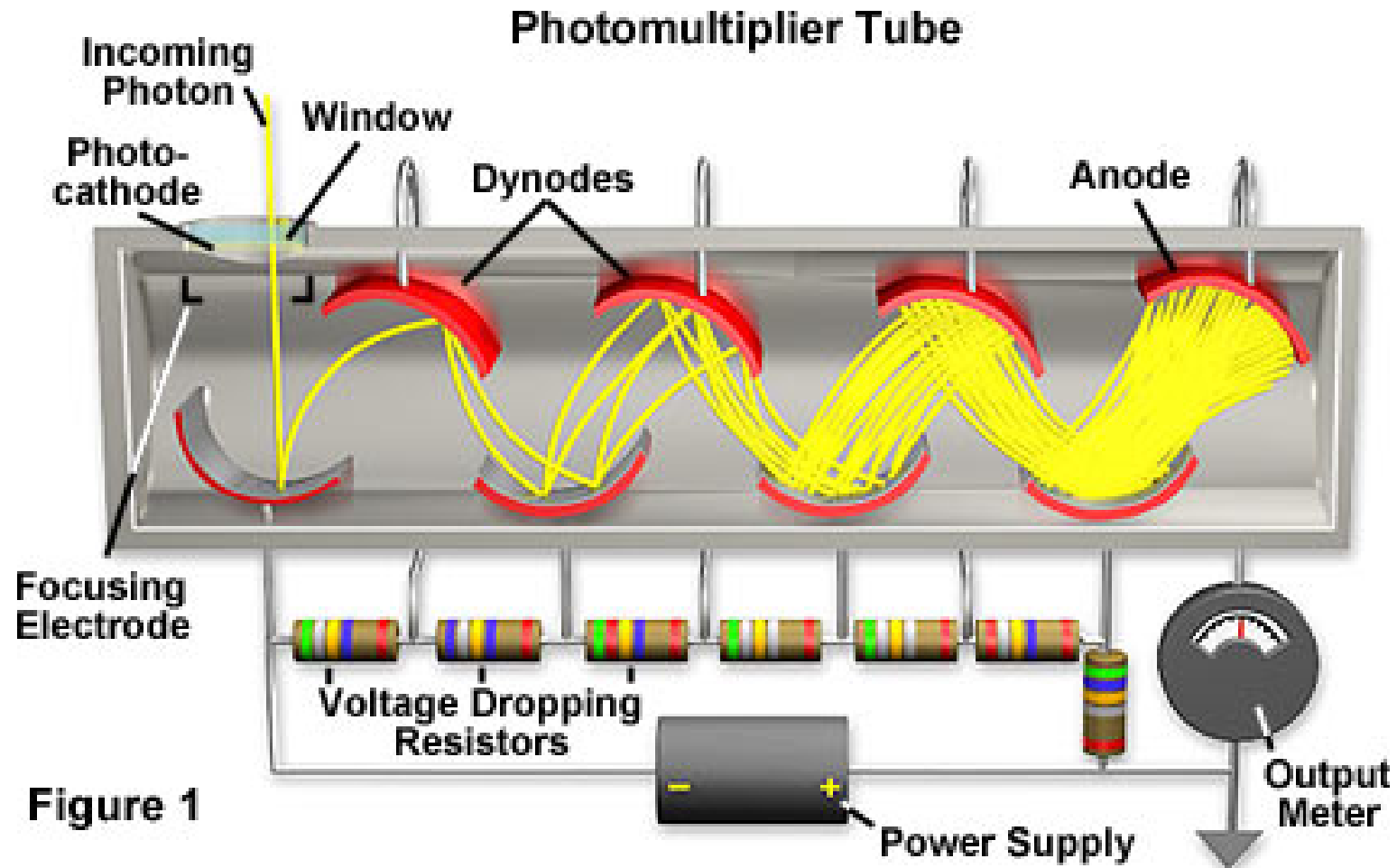
Slow (>50 ns)

Low dynamic range



0.025 mm wide

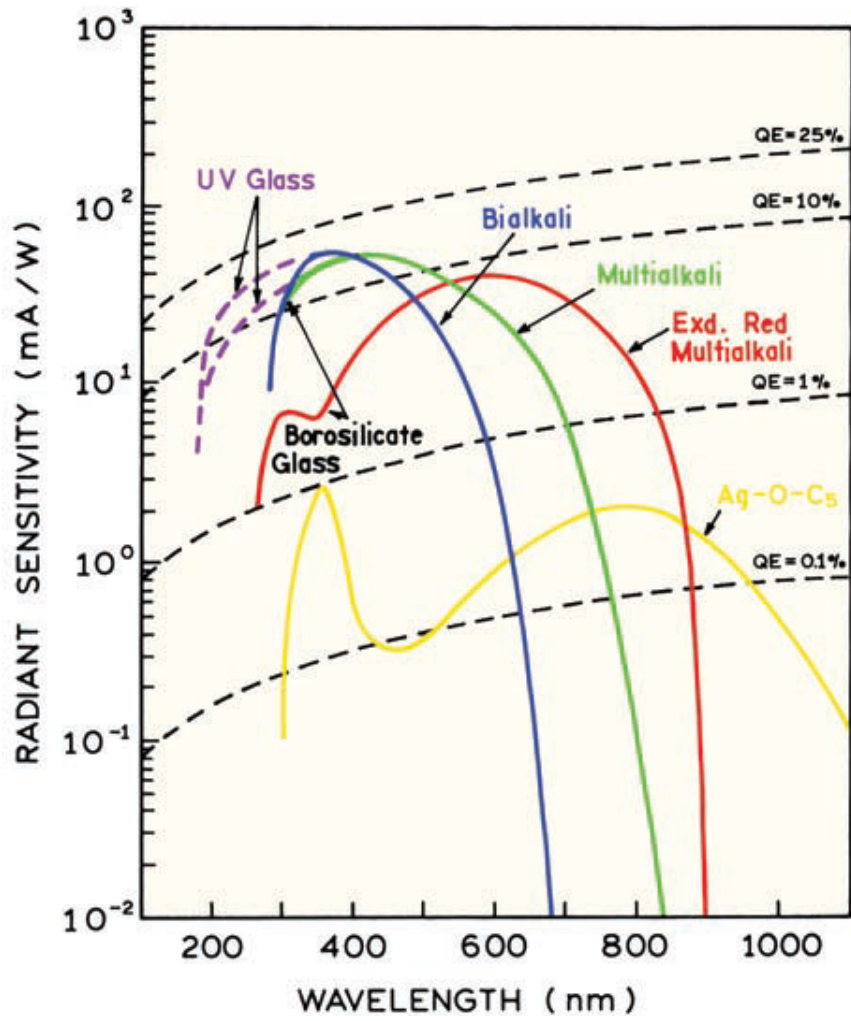
Detectors: PMT



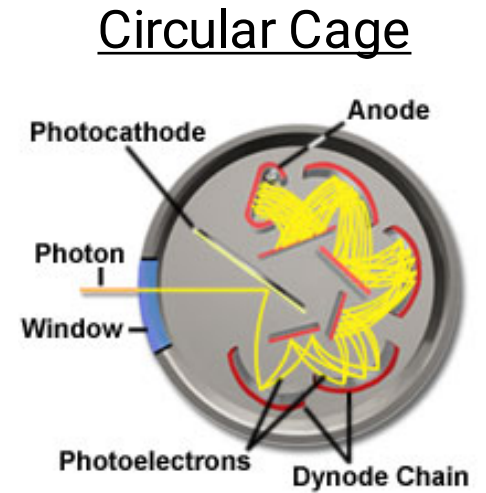
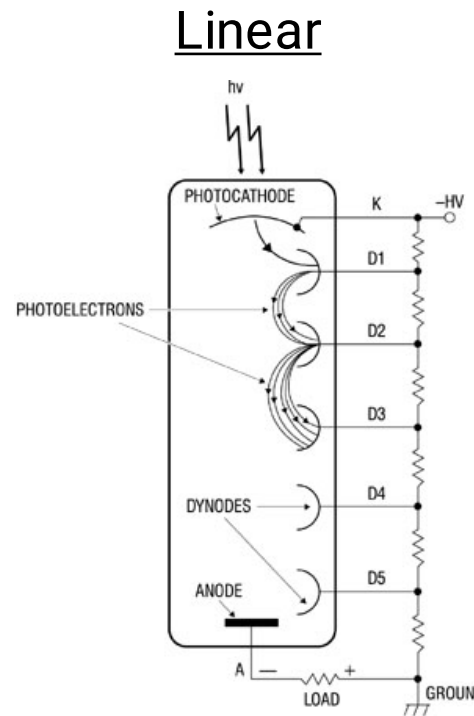
- Cathode: 1 photon = 5-20 electrons
- More positive potential with each dynode
- Operated at -1000 to -2000 V

Detectors: PMT

Photocathodes



Architectures



Detectors: PMT

Pros:

Extremely sensitive

UV-Vis-nIR

100,000,000x current amplifier
(single photons)

Low Noise

Compact

Inexpensive (\$175-500)

Cons:

No wavelength discrimination

Wavelength dependent τ

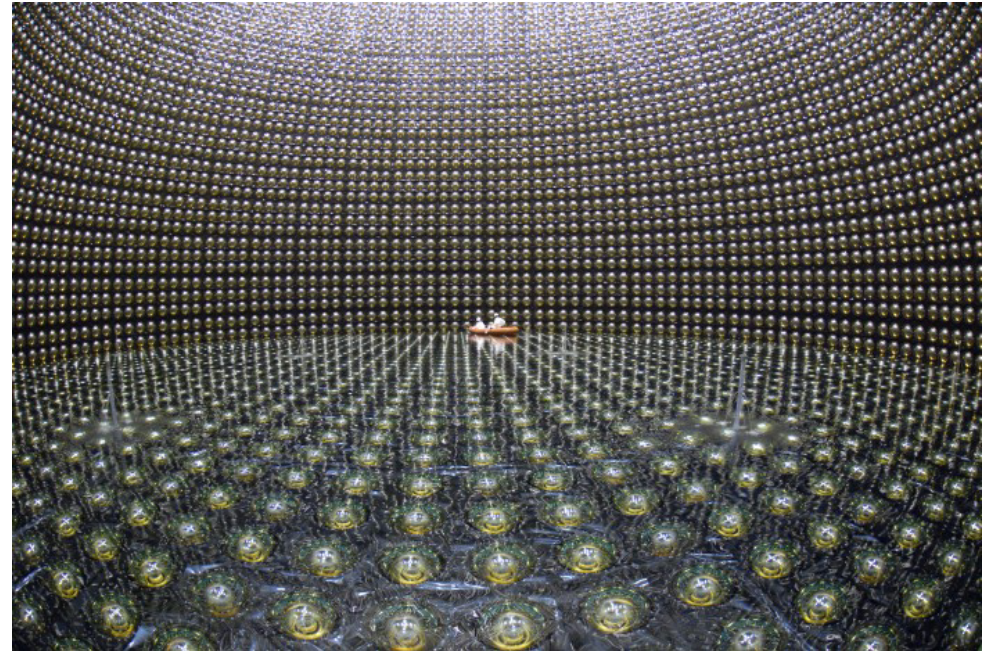
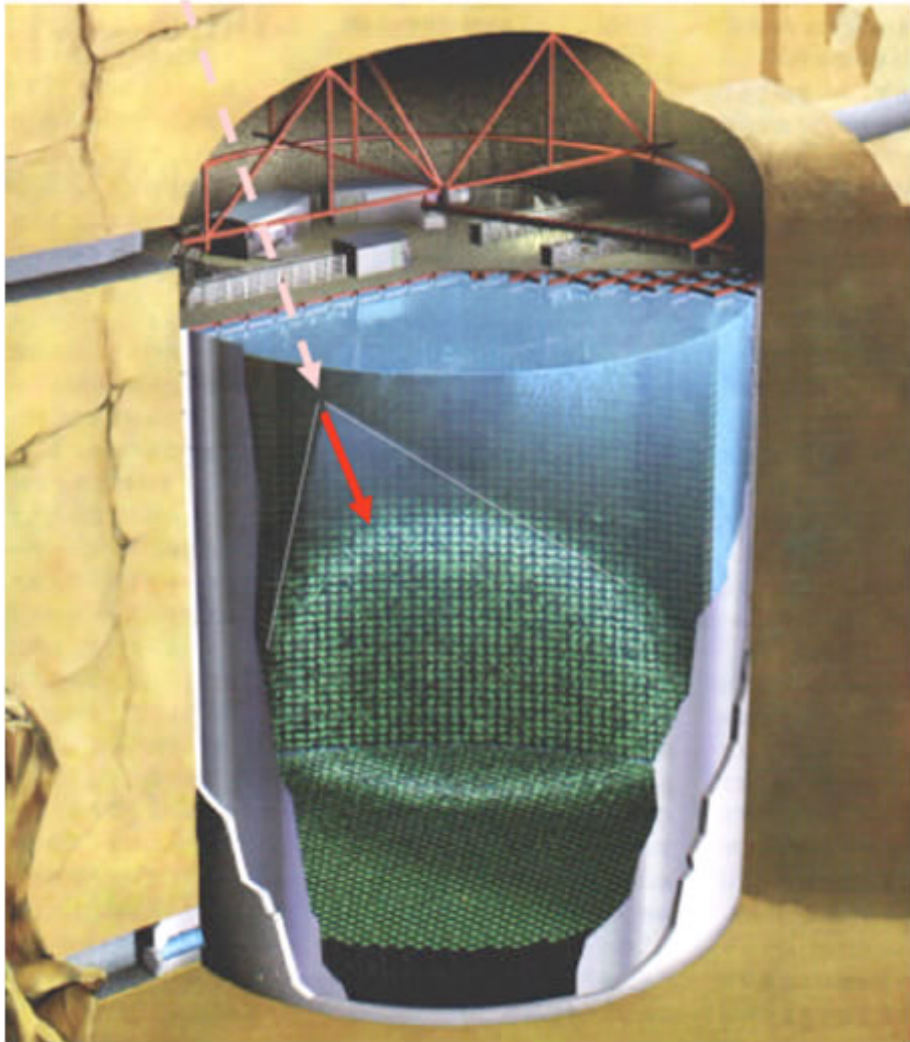
Saturation

Magnetic Field Effects



Detectors: PMT

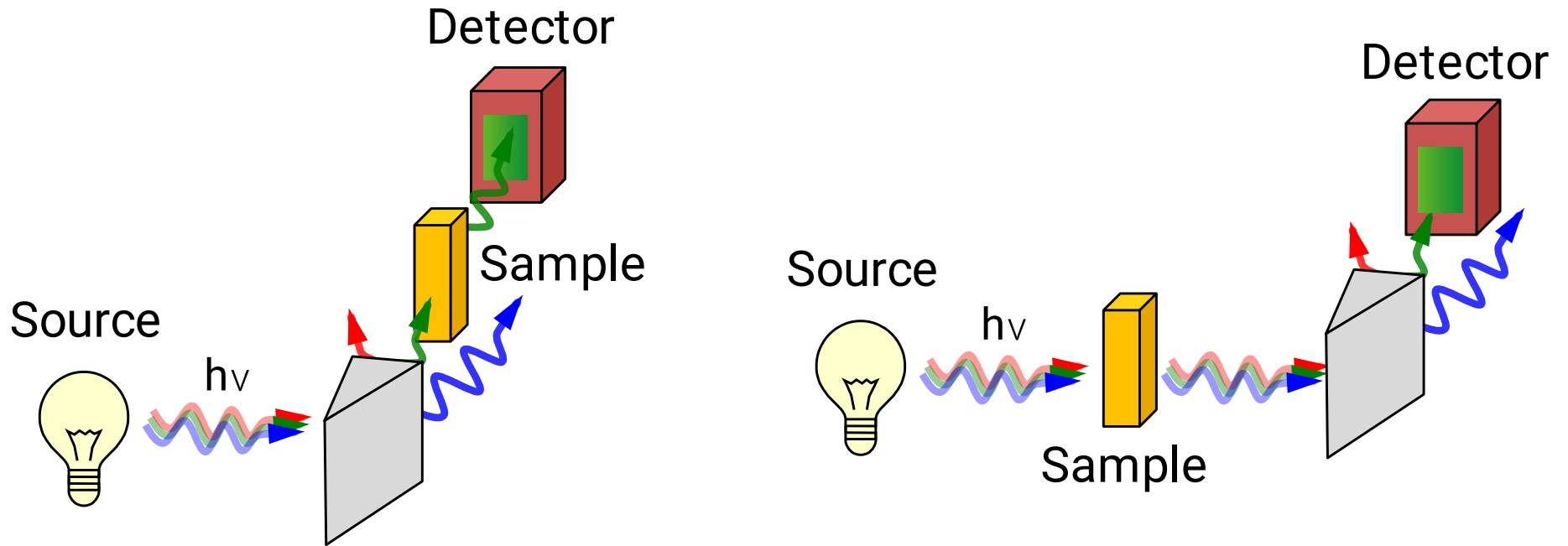
Super-Kamiokande Experiment



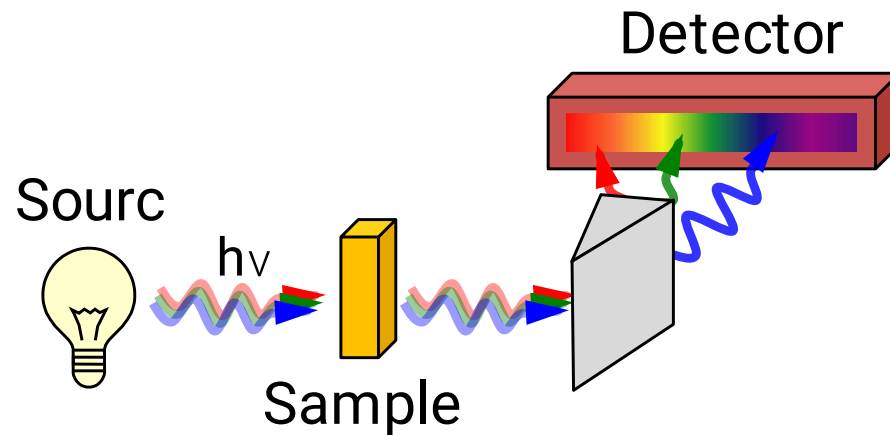
- 1 km underground
- $h = 40 \text{ m}$, $d = 40 \text{ m}$
- 50,000 tons of water
- 11,000 PMTs
- neutrino + water = Cherenkov Radiation

Instrumentation

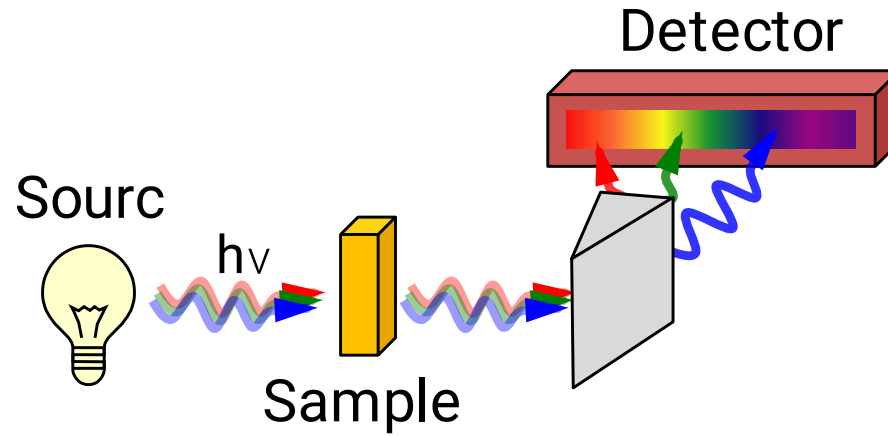
Single λ detection



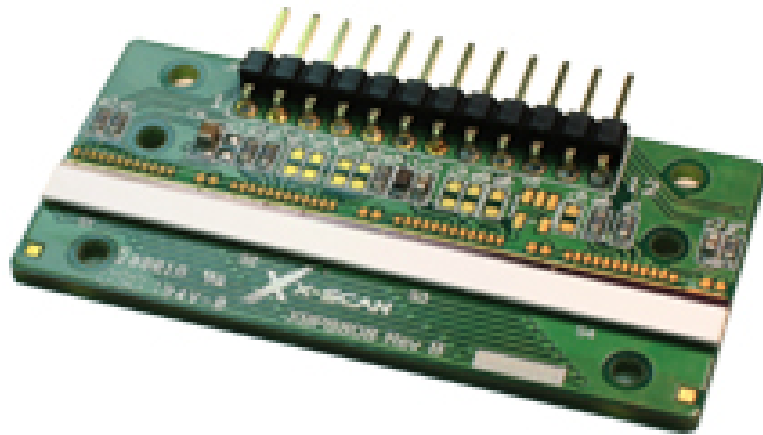
Full spectra detection



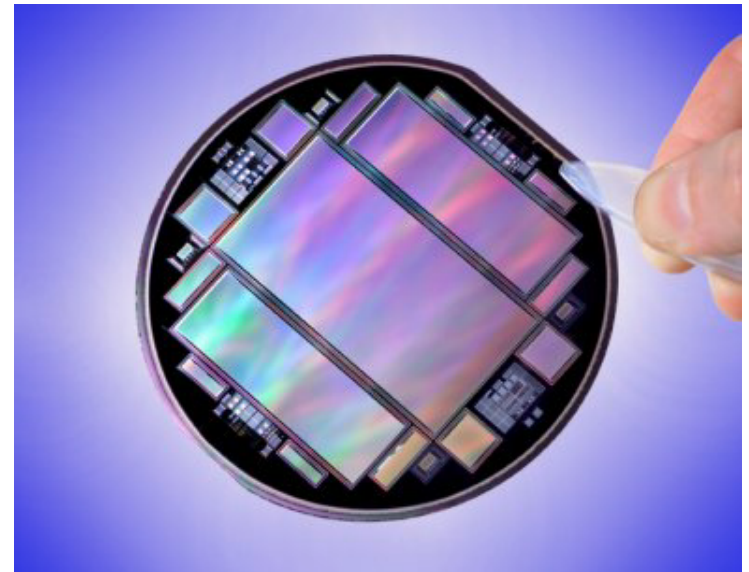
Full Spectrum Detection



Diode Array

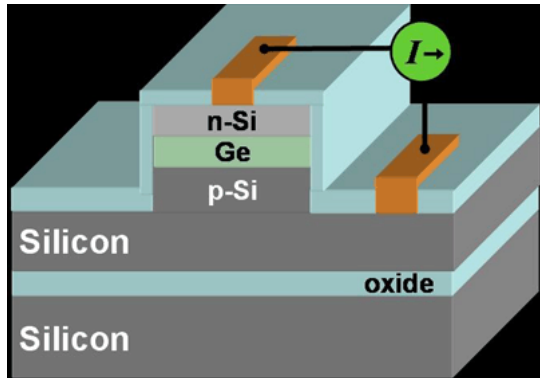


CCD

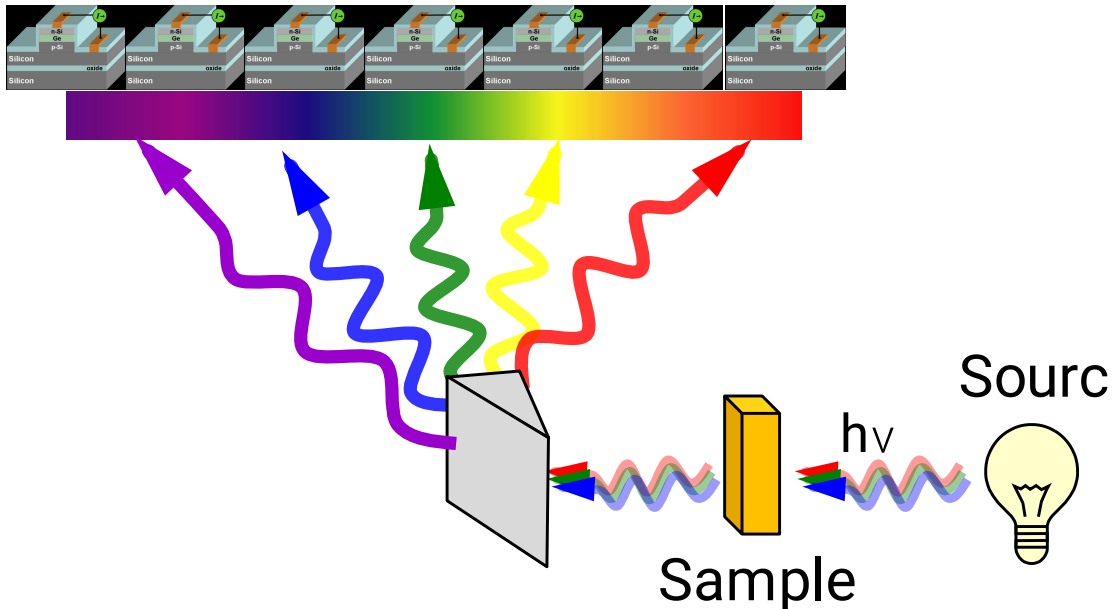


Detectors: Diode Array

Diode



Diode Array



Pros:

Quick measurement

Full spectra in “real time”

Inexpensive

Less moving parts

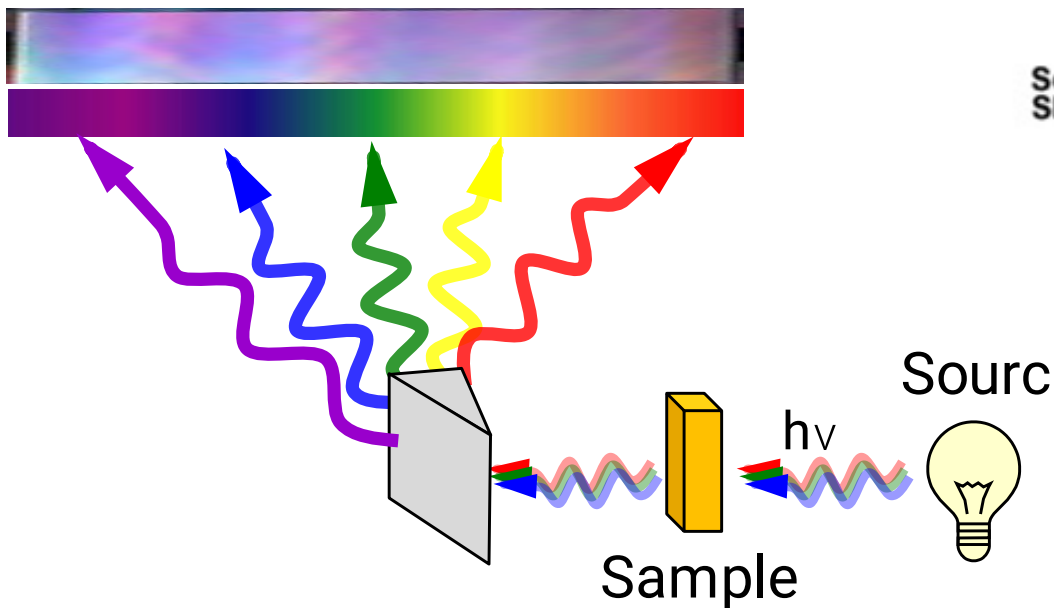
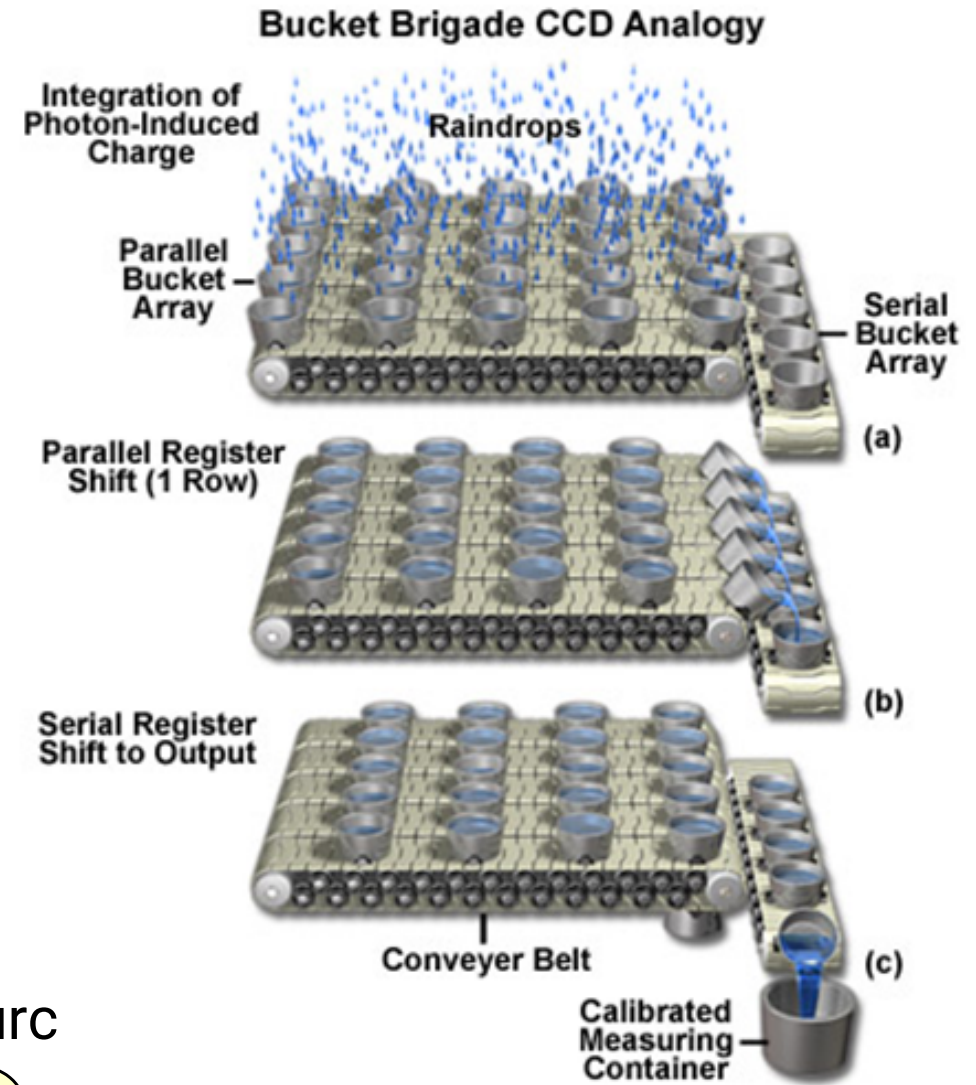
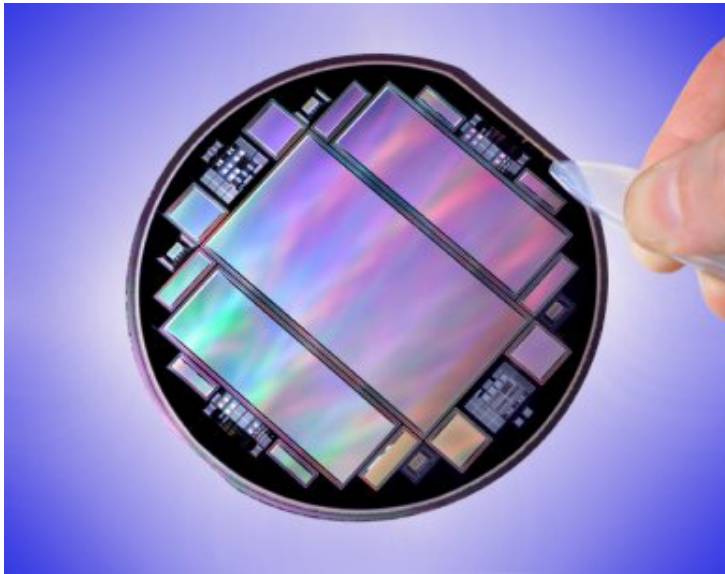
Cons:

Lower resolution (~ 1 nm)

Slow (> 50 ns)

More expensive than a single λ

Detectors: Charge-Coupled Device



Detectors: CCD

Anatomy of a Charge Coupled Device (CCD)

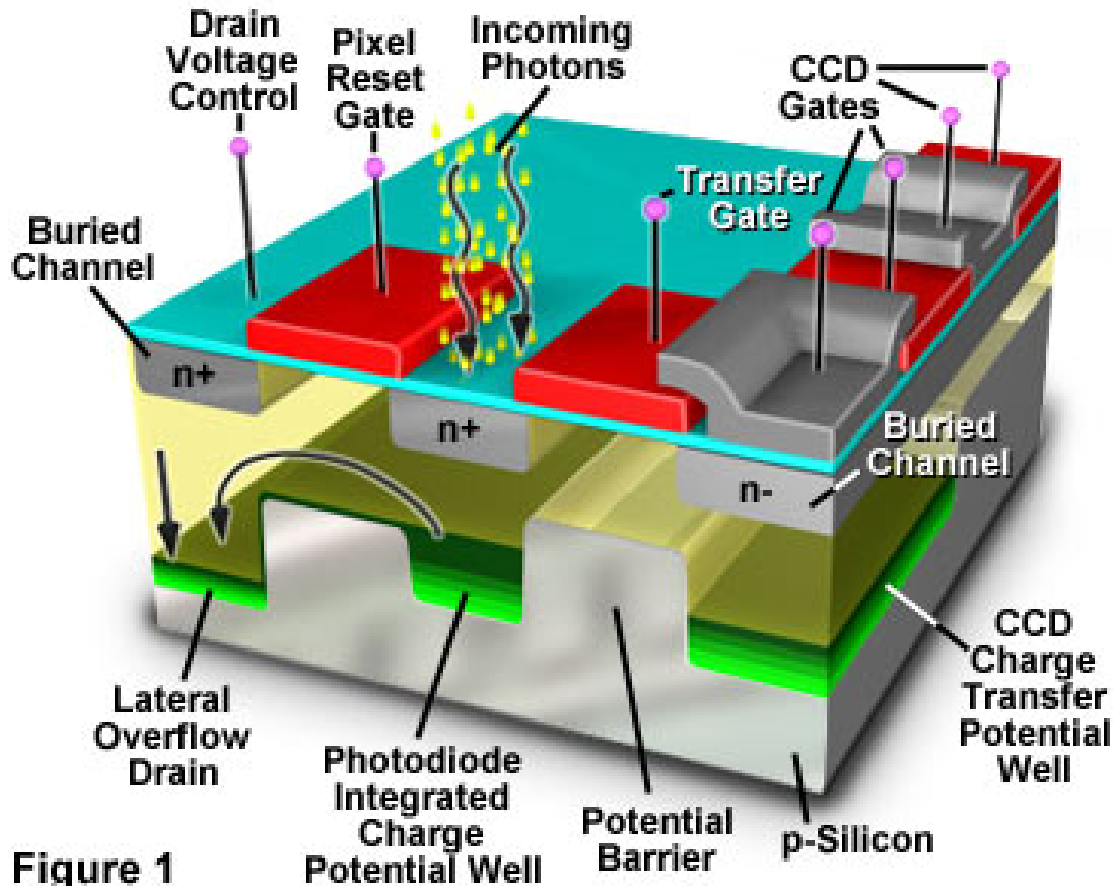


Figure 1

Pros:

Fast

Efficient (~80 % quantum yield)

Full visible spectrum

Wins you the 2006 Nobel Prize (Smith and Boyle)

Cons:

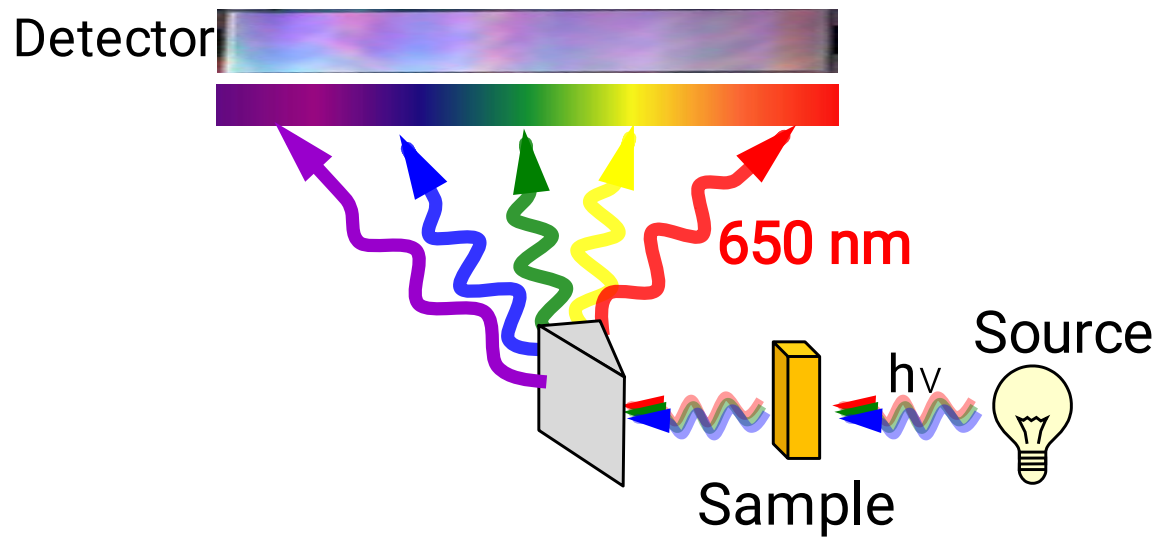
Lower dynamic range

Fast (<50 ns)

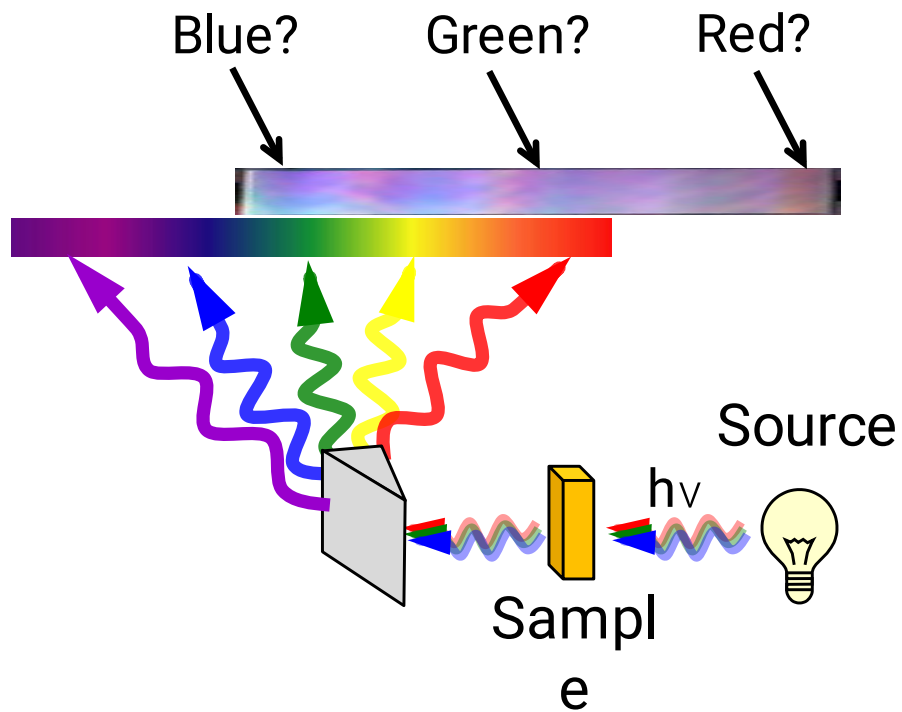
Gaps between pixels

Expensive (~\$10,000-20,000)

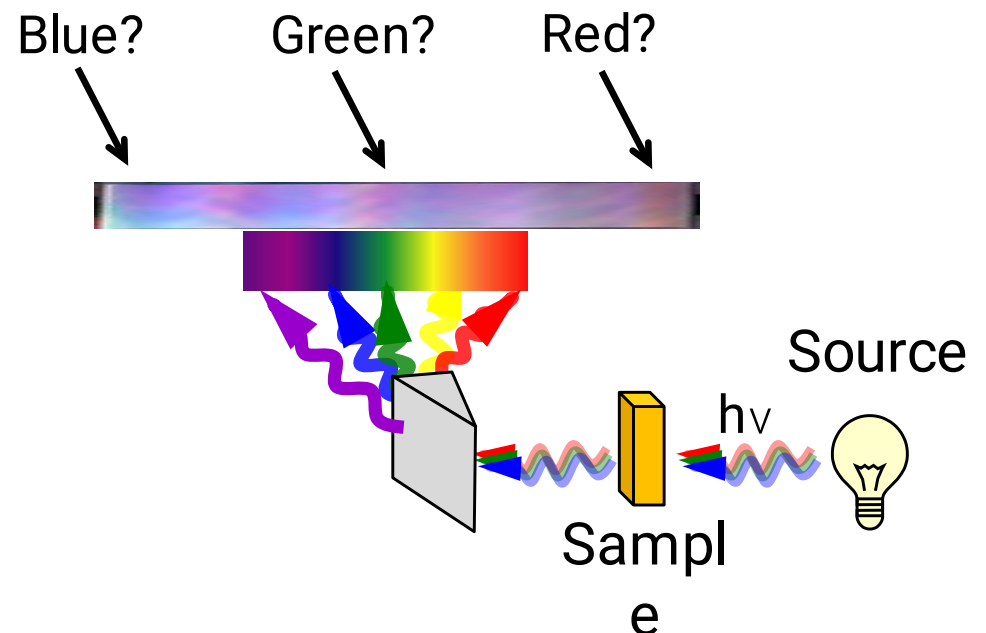
Area Detector Calibration



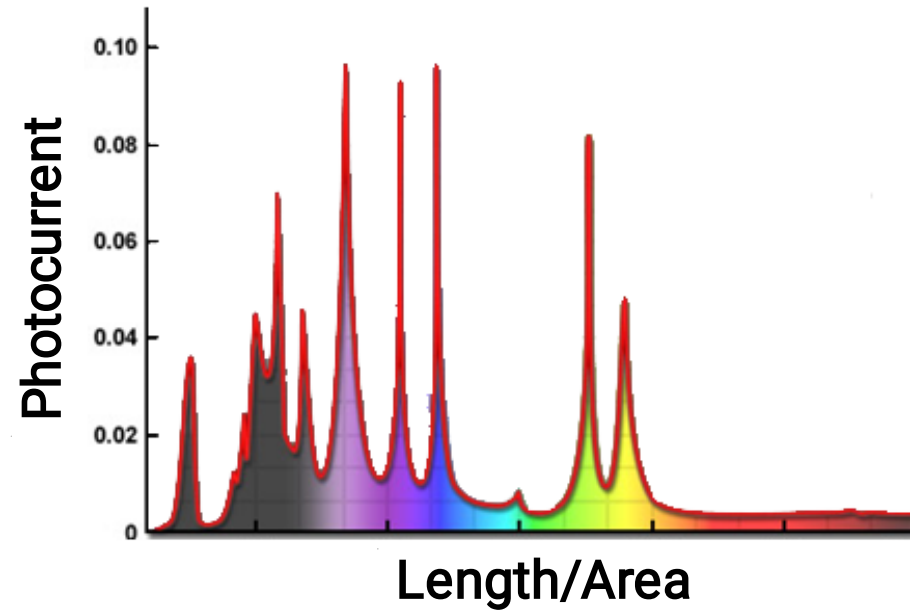
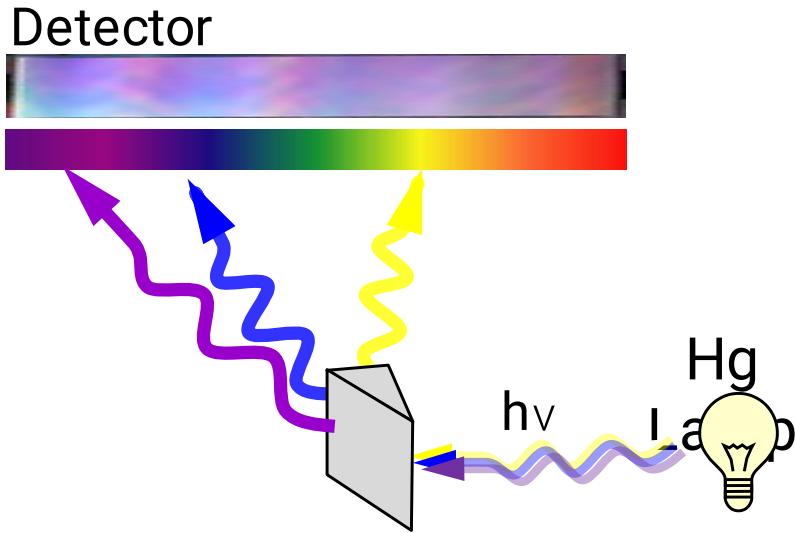
Detector Offset



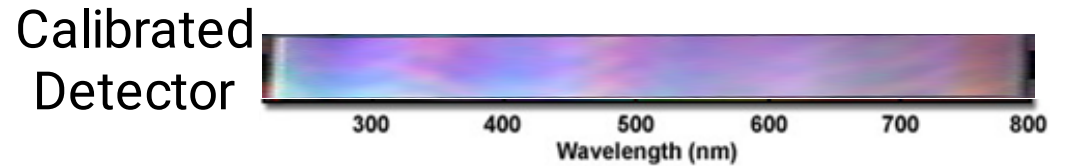
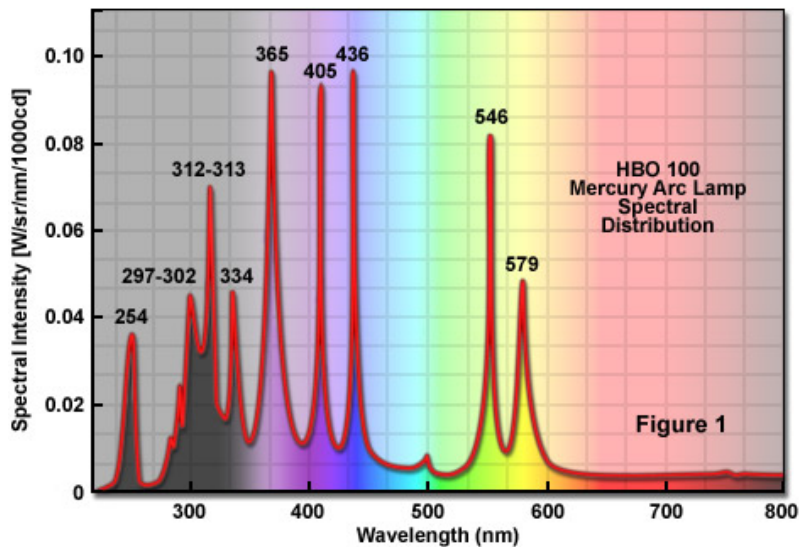
Detector To Close



Area Detector Calibration

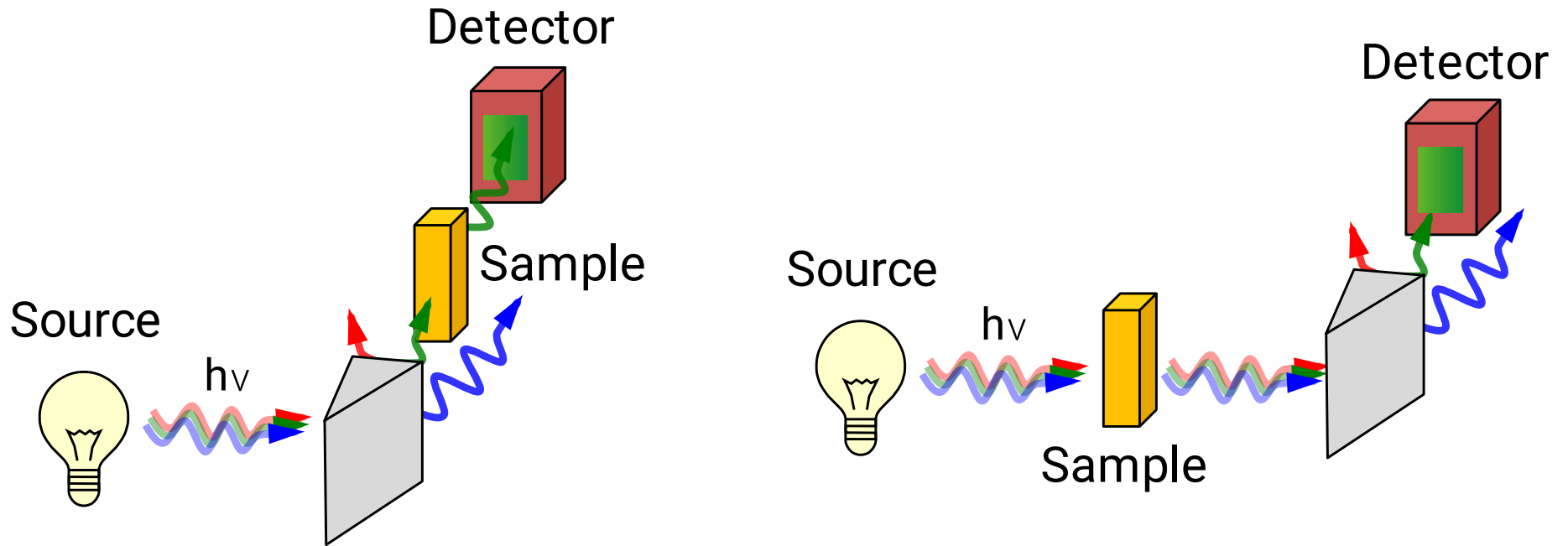


Hg Lamp Spectrum

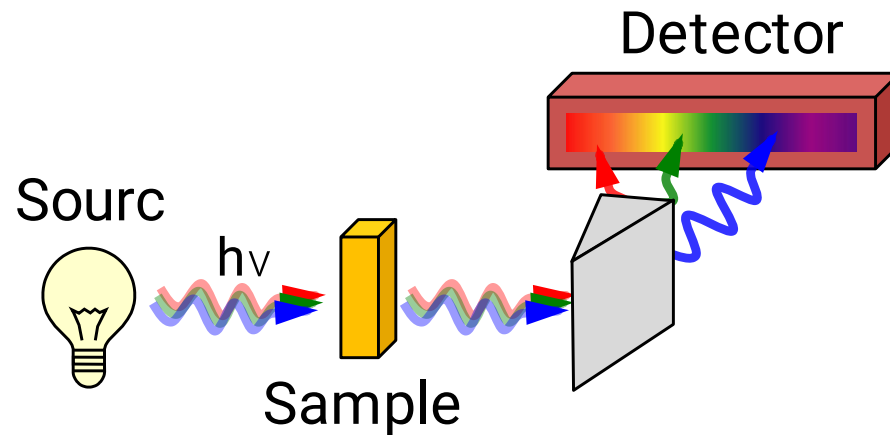


Instrumentation

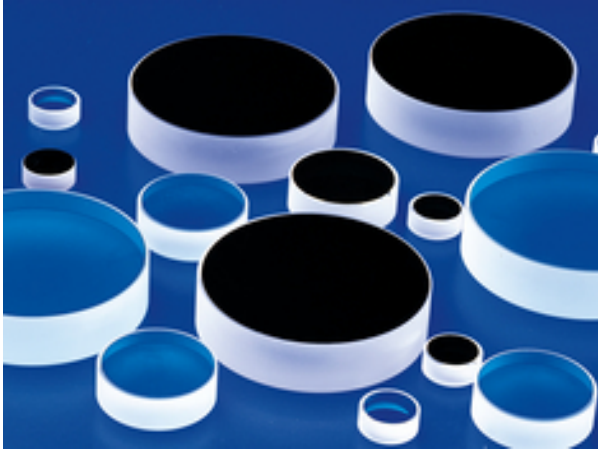
Single λ detection



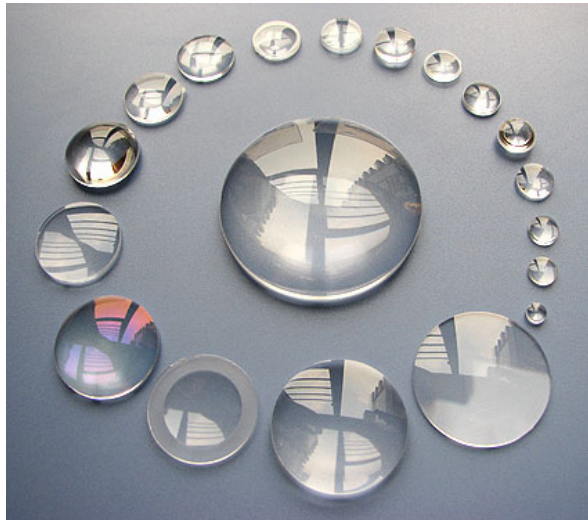
Full spectra detection



Other Components

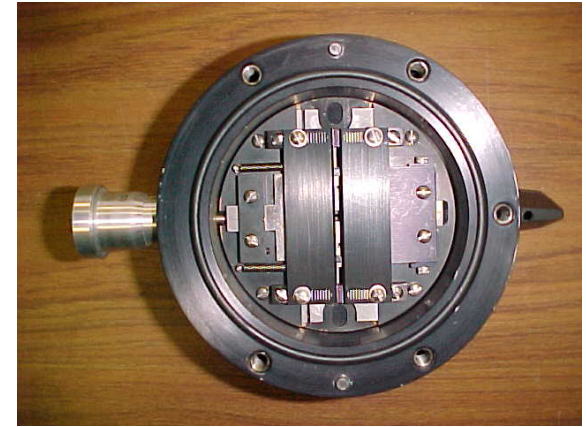
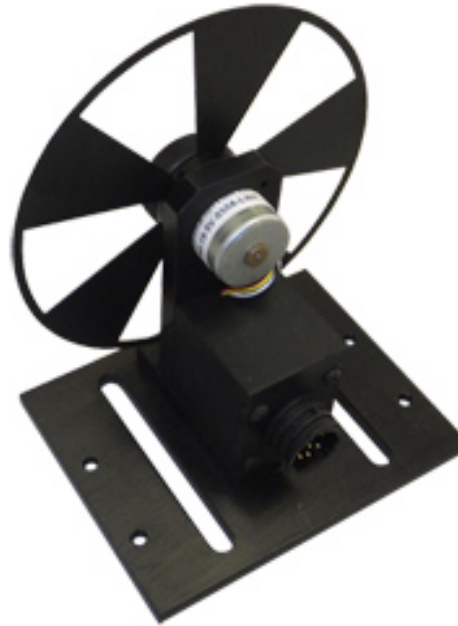


Mirrors



Lenses

Chopper



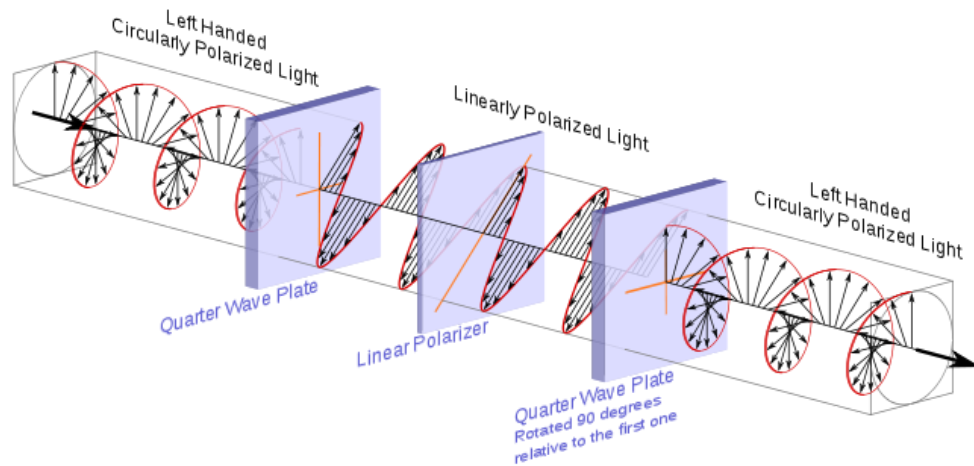
Entrance/Exit Slits



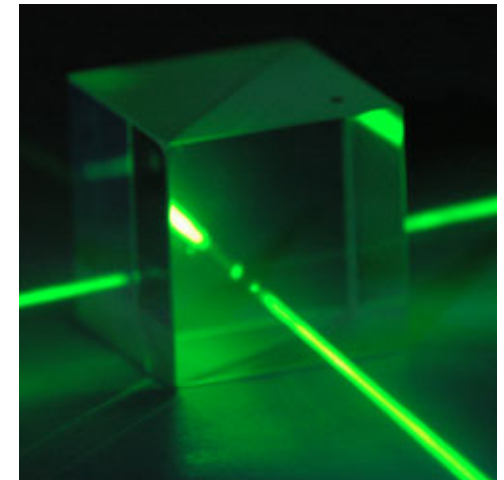
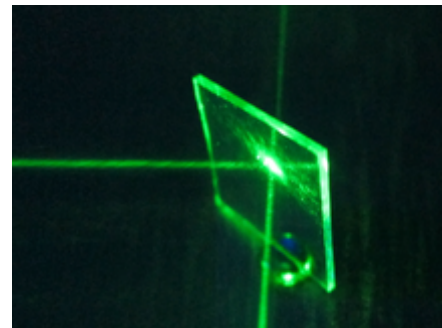
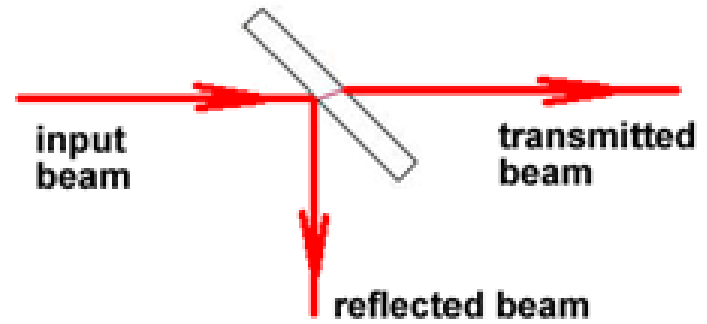
Shutter

Other Components

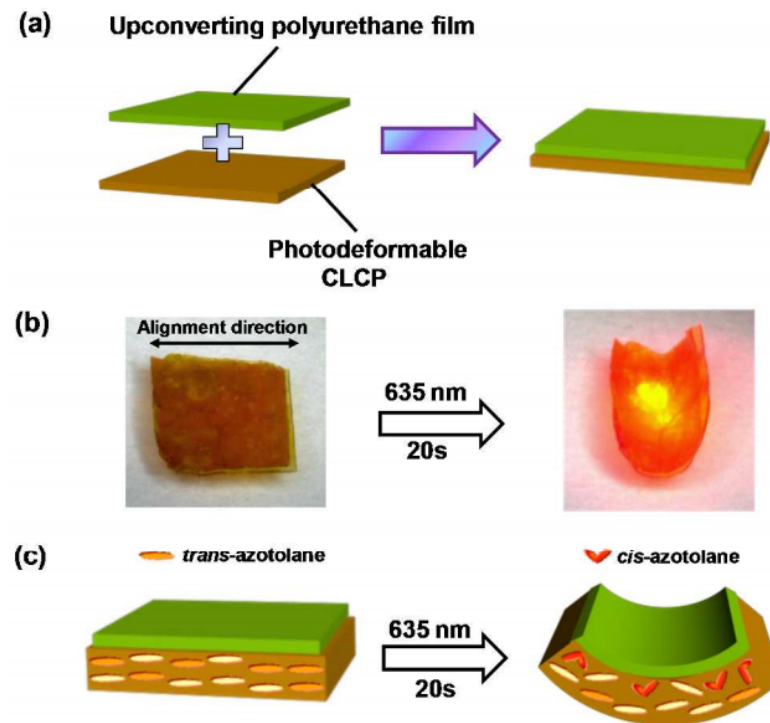
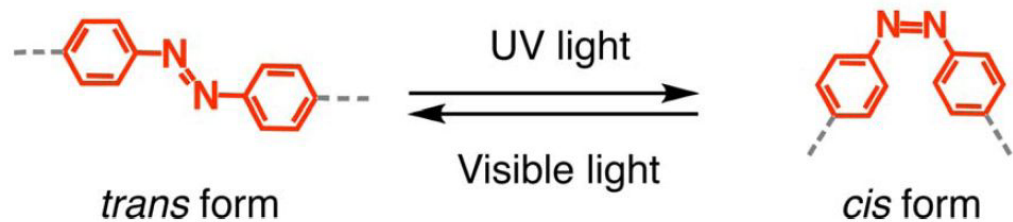
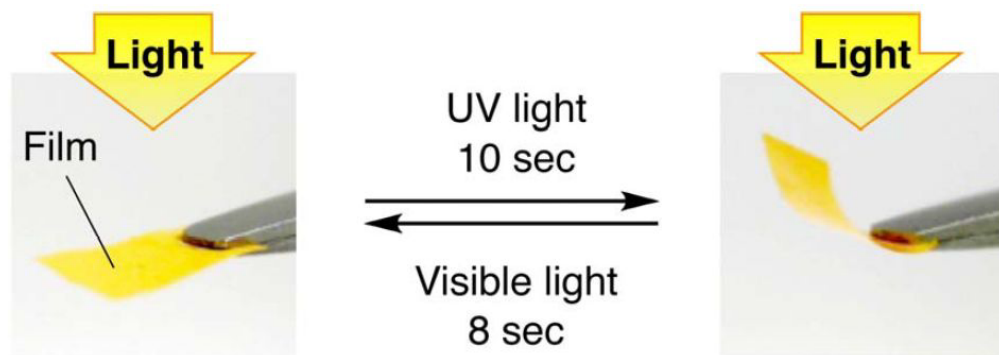
Polarizer



Beam Splitter



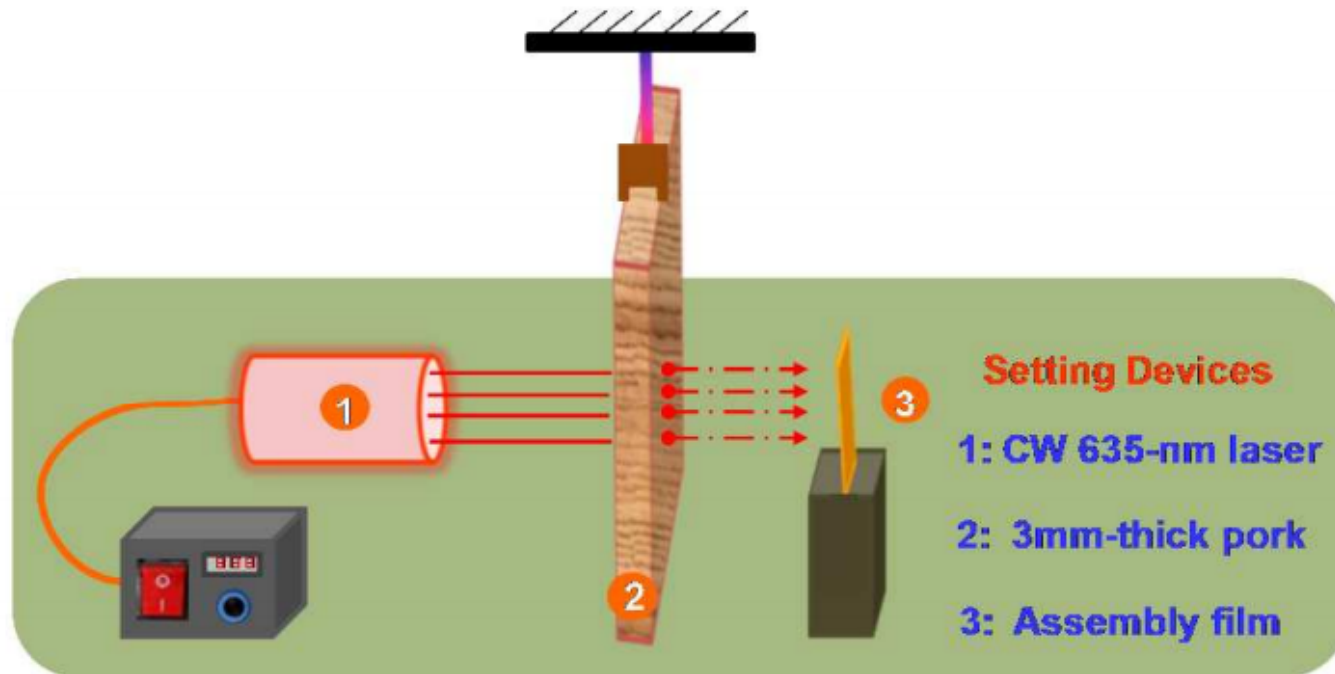
Side Note: Pub Highlight



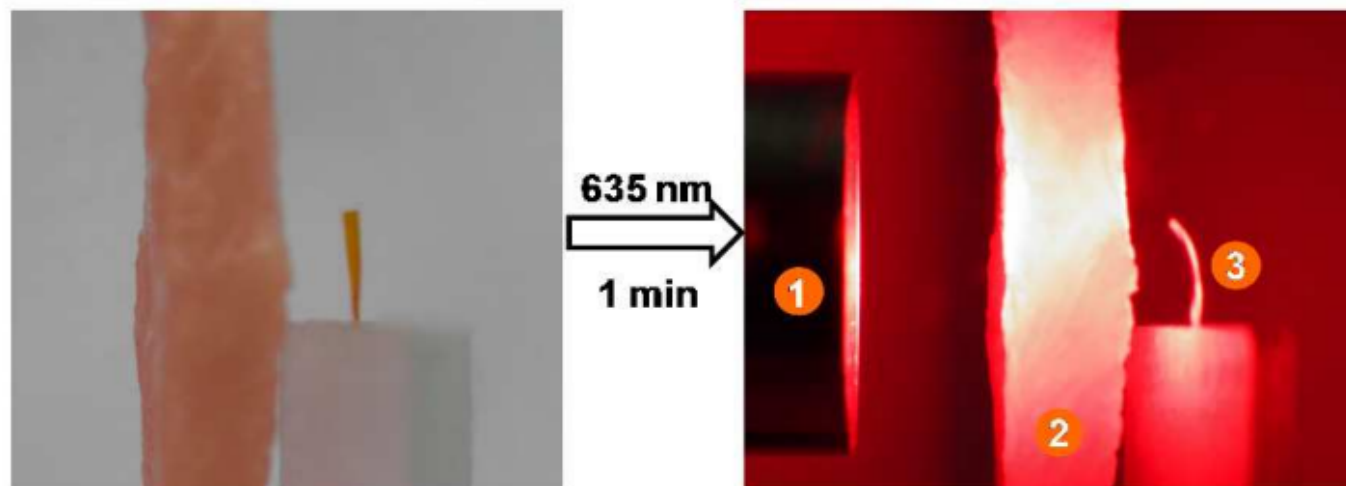
DOI: 10.1021/ja406020r

Side Note: Pub Highlight

(a)

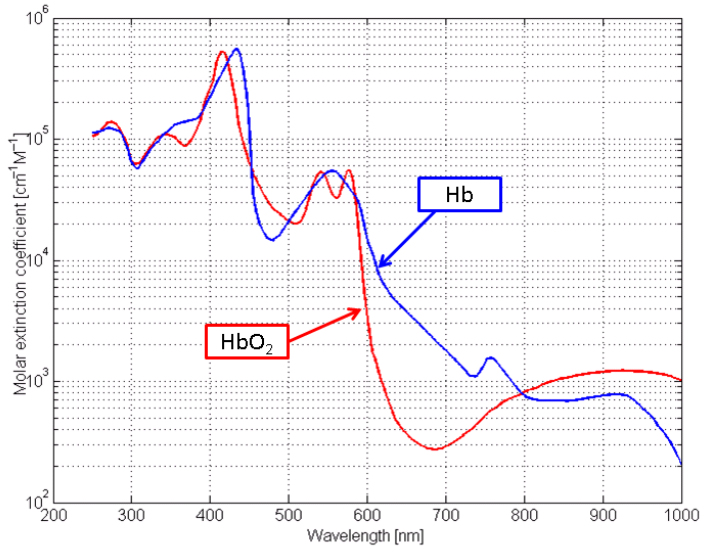


(b)

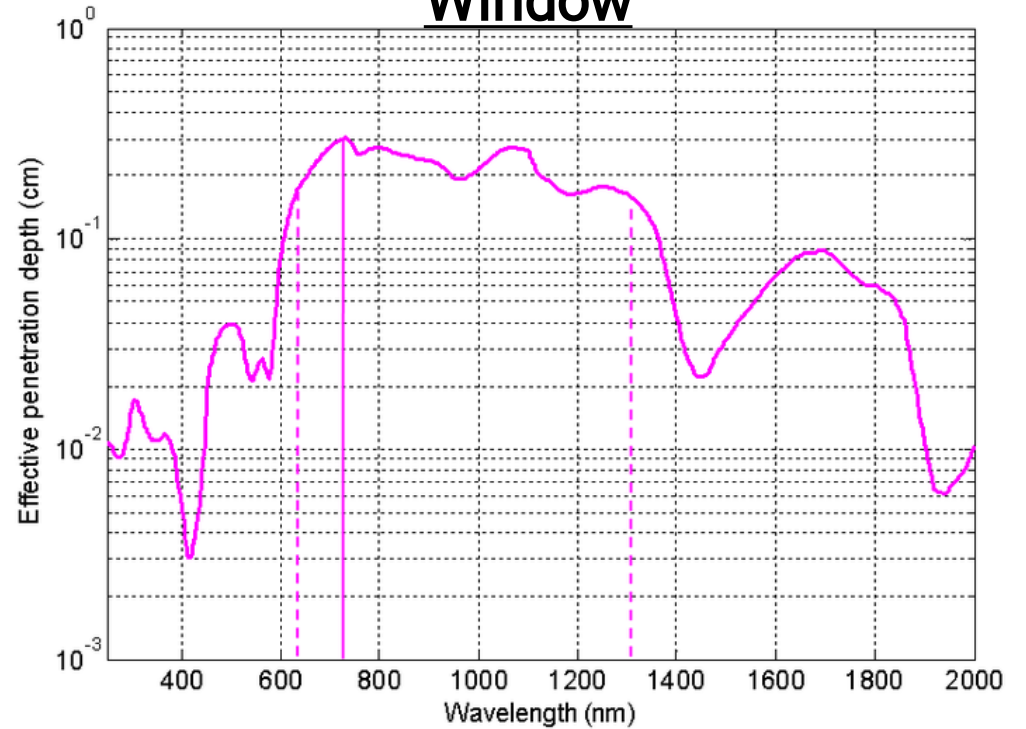


Side Note: Pub Highlight

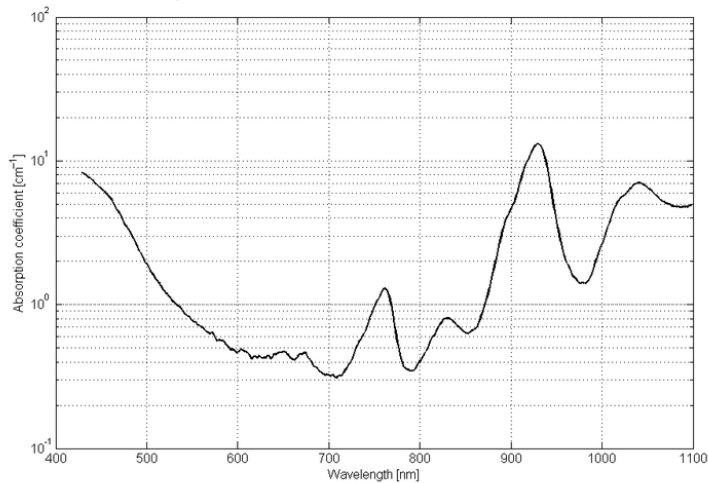
Hemoglobin Absorption



Biological Tissue Window



Pig Lard Absorption



Side Note: Pub Highlight

Plasmonic Heating

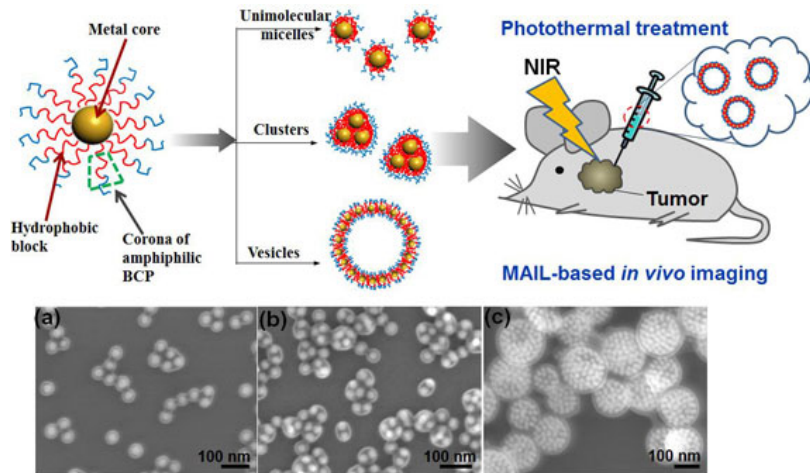
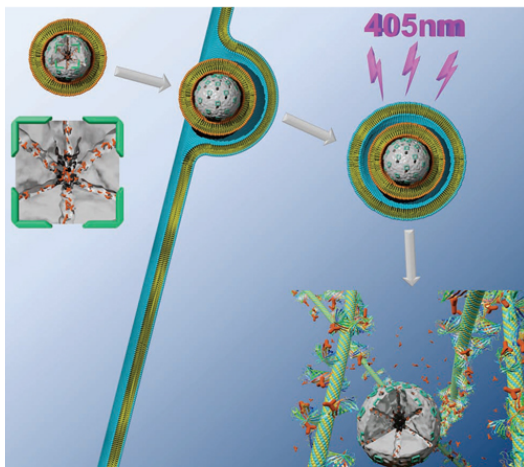
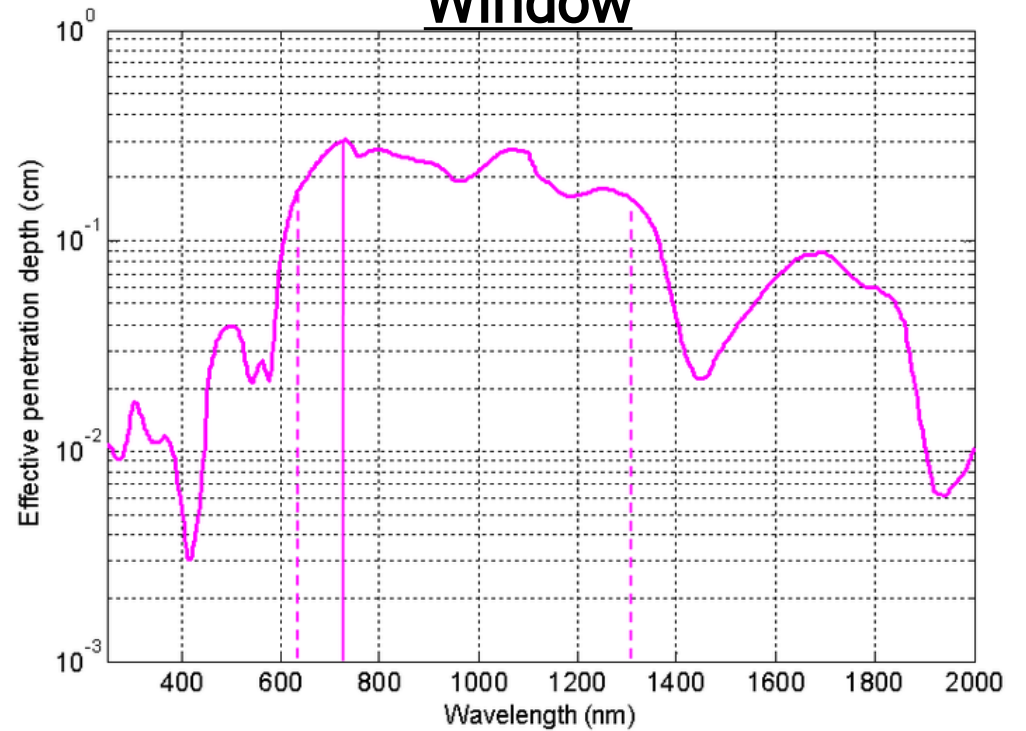


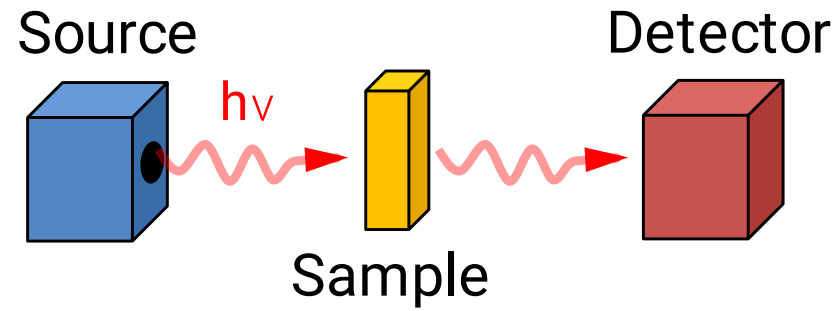
Photo Drug Delivery



Biological Tissue Window



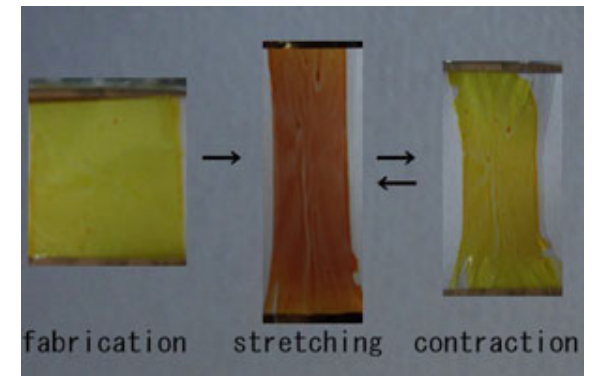
THE SAMPLE



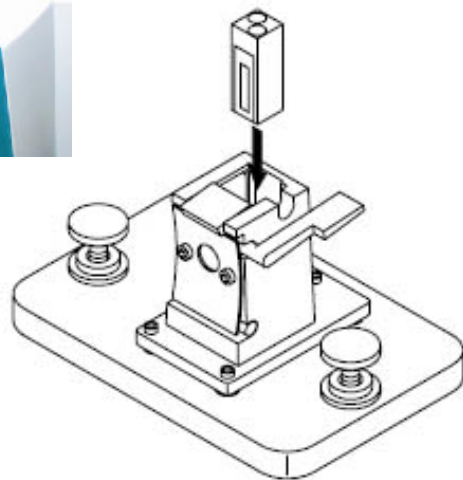
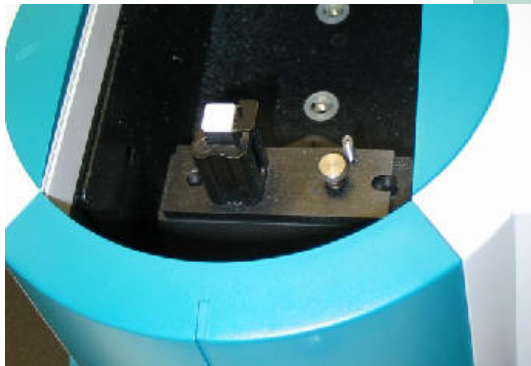
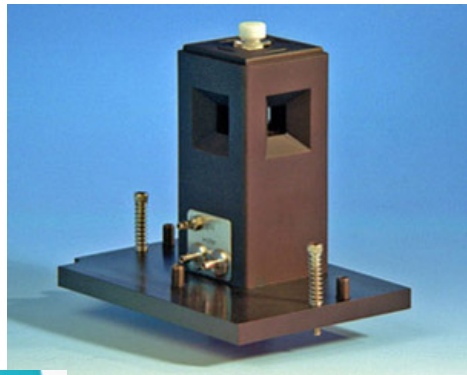
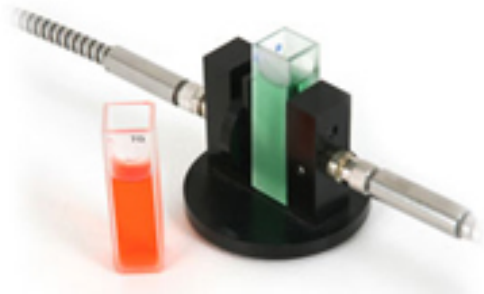
Solutions



Solids



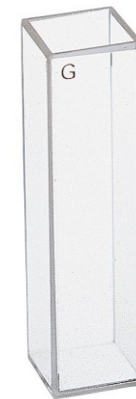
The Sample: Cuvette for Solutions



Plastic



Glass



Quartz

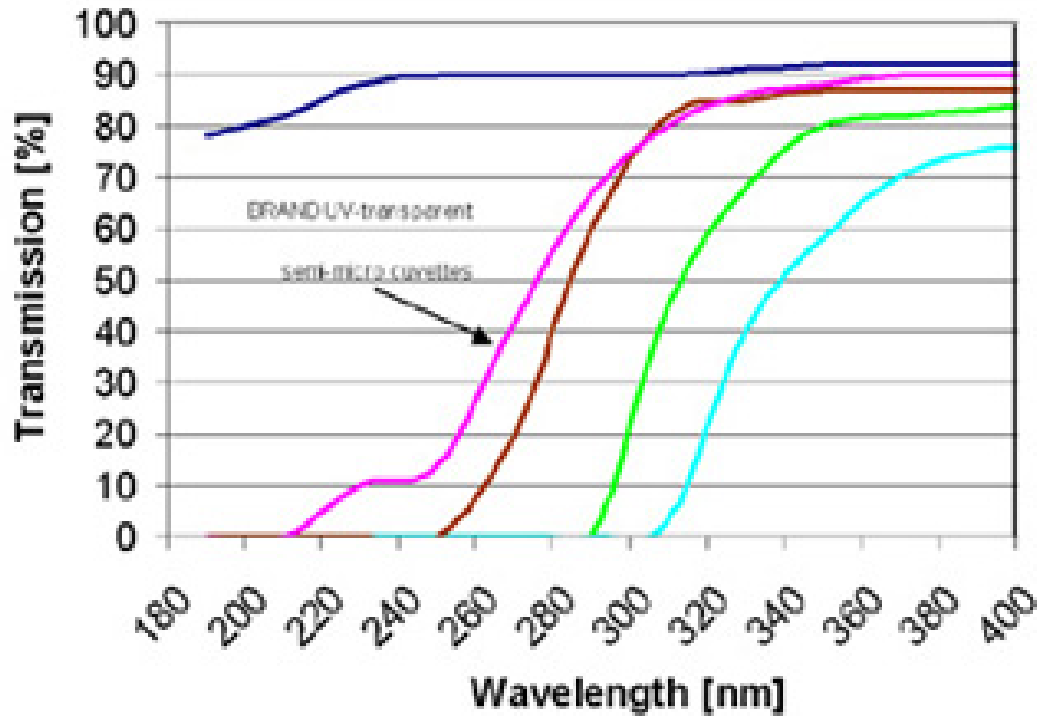


Typically 1 x 1 cm

$$A = \epsilon c l$$

The Sample: Cuvette

Transmission Window



(—) PC	> 340 nm	
(—) Polystyrene	> 320 nm	\$0.25
(—) PMMA	> 300 nm	\$0.29
(—) Glass	> 270 nm	\$100
(—) Quartz	> 170 nm	\$200



Polystyrene

Aceton
e →



The Sample: Specialty Cuvettes

$$A = \frac{\text{Path length}}{\epsilon C l}$$

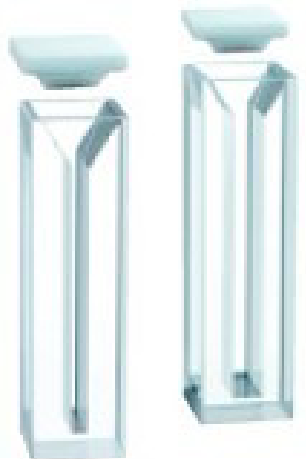
Dilute Samples



10 x 1
cm

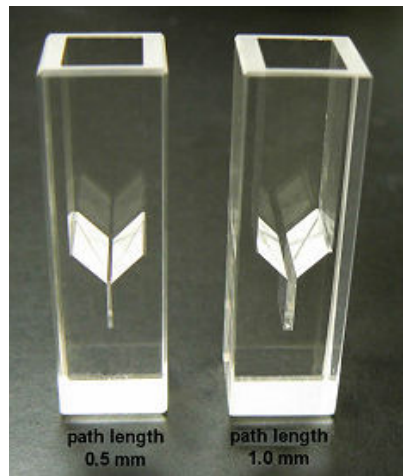
1 x 1
cm

Concentrated Samples



0.2 cm

0.5 cm



path length
0.5 mm

path length
1.0 mm

Flow Cell



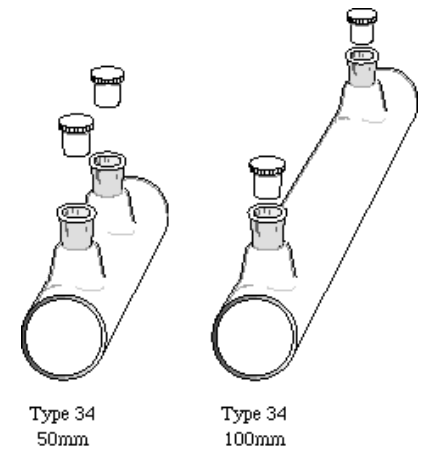
Spec-
chem



Air-free



Gas Cell

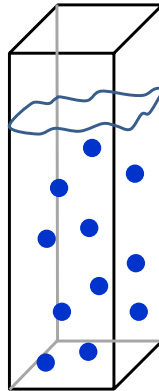


Type 34
50mm

Type 34
100mm

The Sample: Solvent

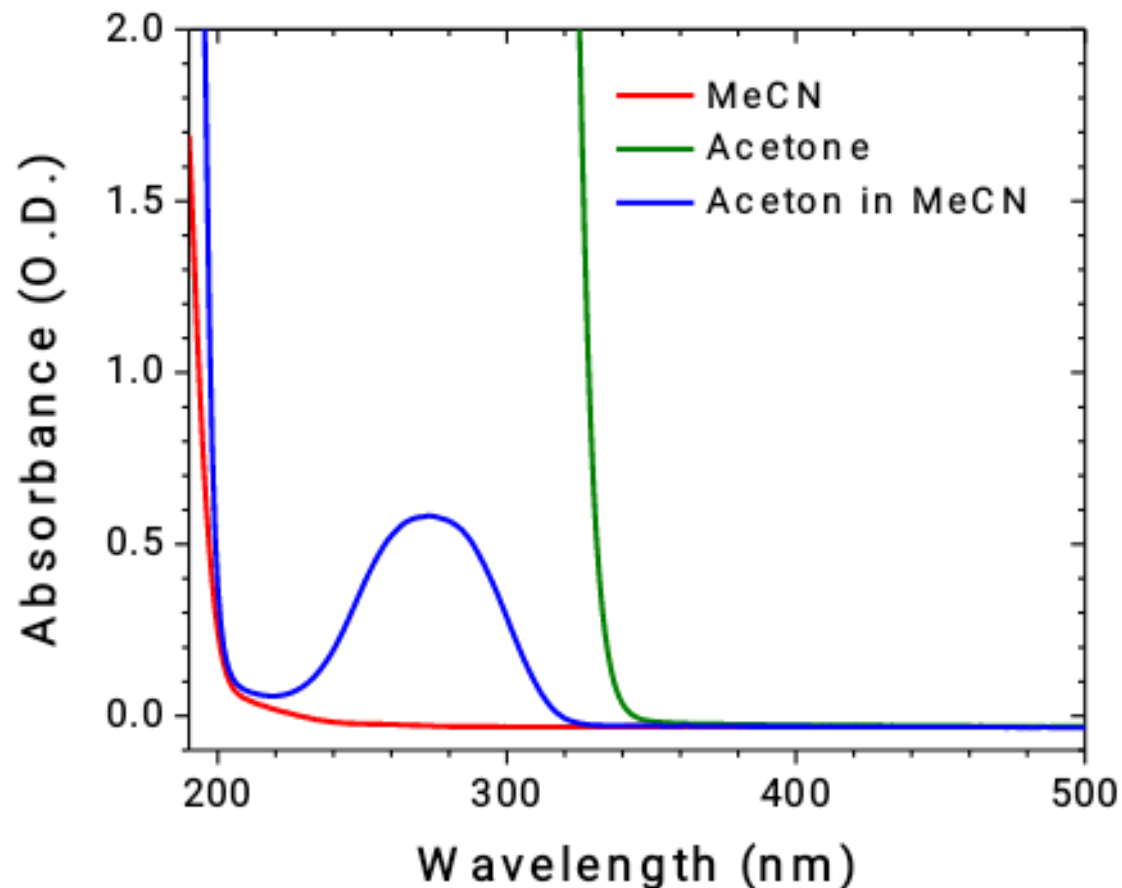
- Concentration (typically <math><50\ \mu\text{M}</math>)
- Solubility
- Ionic strength
- Hydrogen bonding
- Aggregation
- π -stacking
- Solvent absorption



Common solvent cutoffs in nm:

water	190
acetonitrile	190
isooctane	195
cyclohexane	200
n-hexane	200
ethanol	205
methanol	210
ether	210
1,4-dioxane	215
THF	220
CH_2Cl_2	235
Chloroform	240
CCl_4	265
benzene	280
toluene	285
acetone	340

The Sample: Solvent

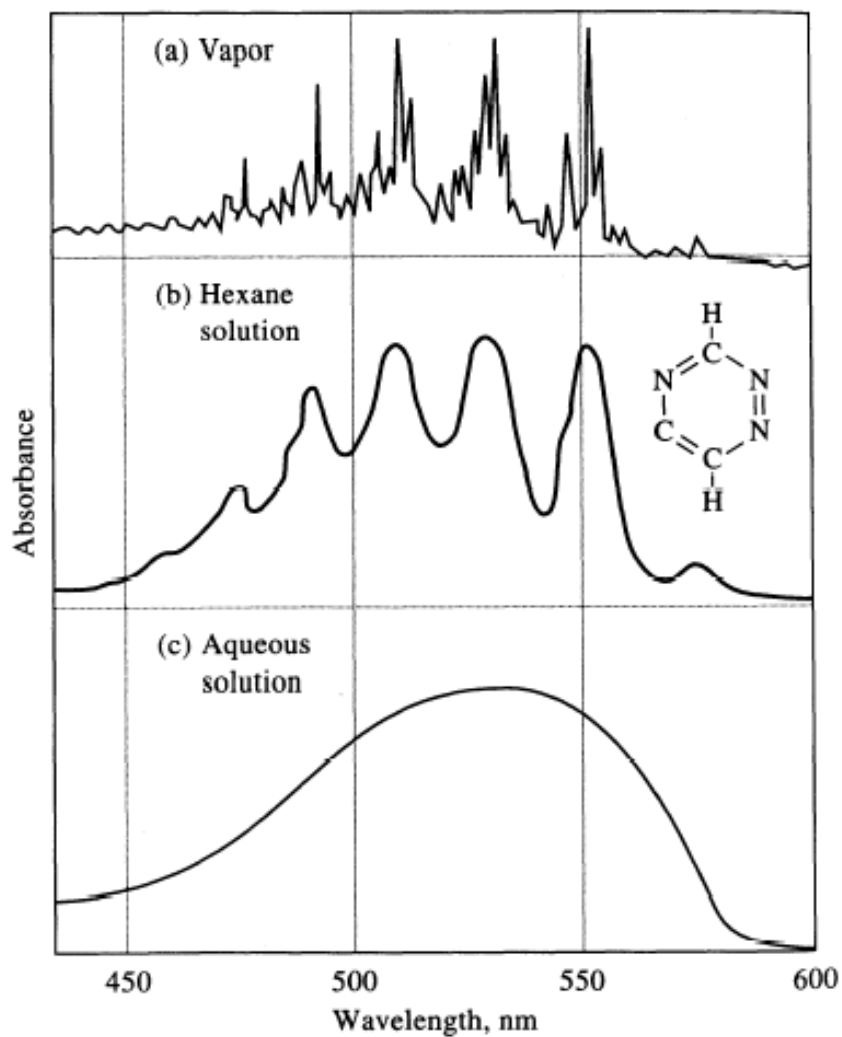


Common solvent cutoffs in nm:

water	190
acetonitrile	190
isooctane	195
cyclohexane	200
n-hexane	200
ethanol	205
methanol	210
ether	210
1,4-dioxane	215
THF	220
CH ₂ Cl ₂	235
Chloroform	240
CCl ₄	265
benzene	280
toluene	285
acetone	340

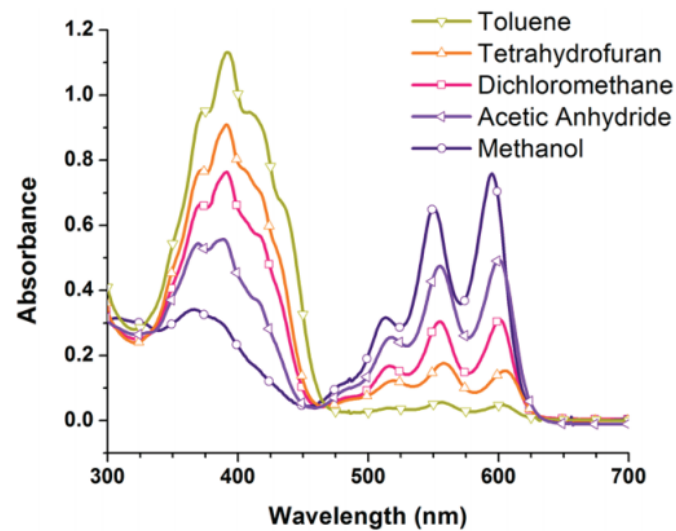
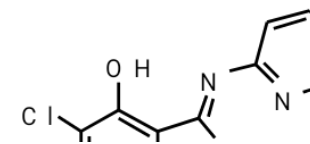
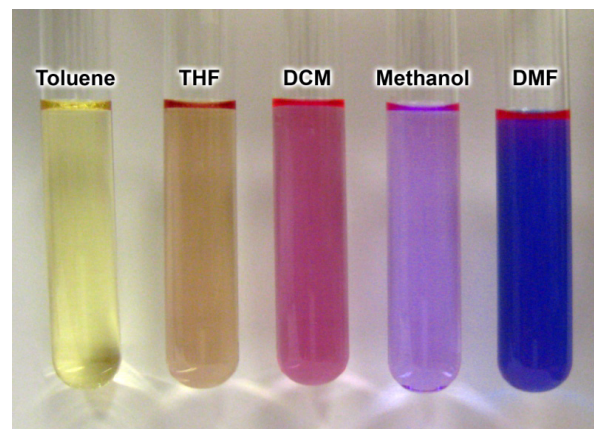
The Sample: Solvent

Vibrational Structure



1,2,4,5-Tetrazine

Solvatochromism



Correcting for background

$$A = -\log T = \log P_0/P$$

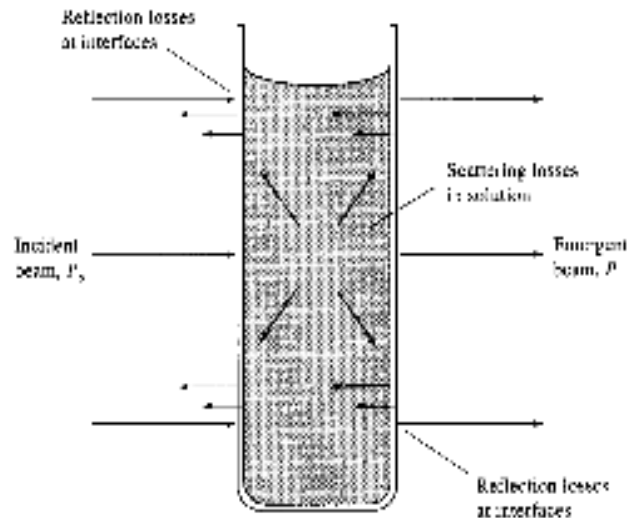
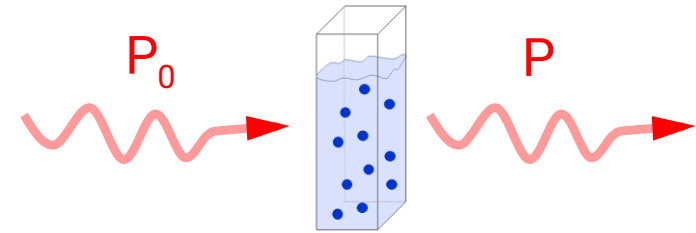
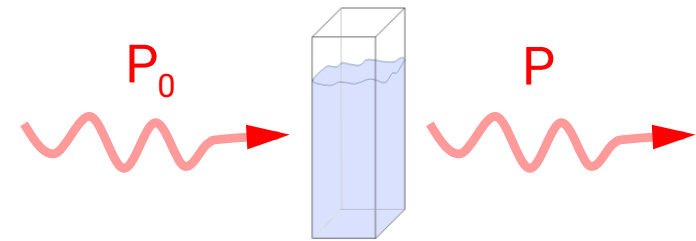


Figure 13-1 Reflection and scattering losses.



cuvette + solvent + sample

$$A_{\text{all}} = A_{\text{cuvette}} + A_{\text{solvent}} + A_{\text{sample}}$$



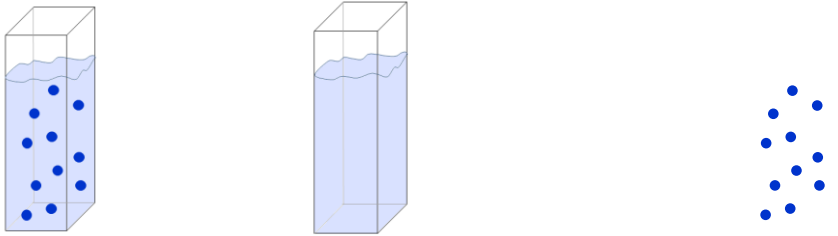
cuvette + solvent

$$A_{\text{background}} = A_{\text{cuvette}} + A_{\text{solvent}}$$

We want to know A ($\log P_0/P$) for only our sample!

$$A_{\text{all}} - A_{\text{background}} = A_{\text{sample}}$$

Instrument Architectures



$$A_{\text{all}} - A_{\text{background}} = A_{\text{sample}}$$

How do we measure background (reference) and sample?

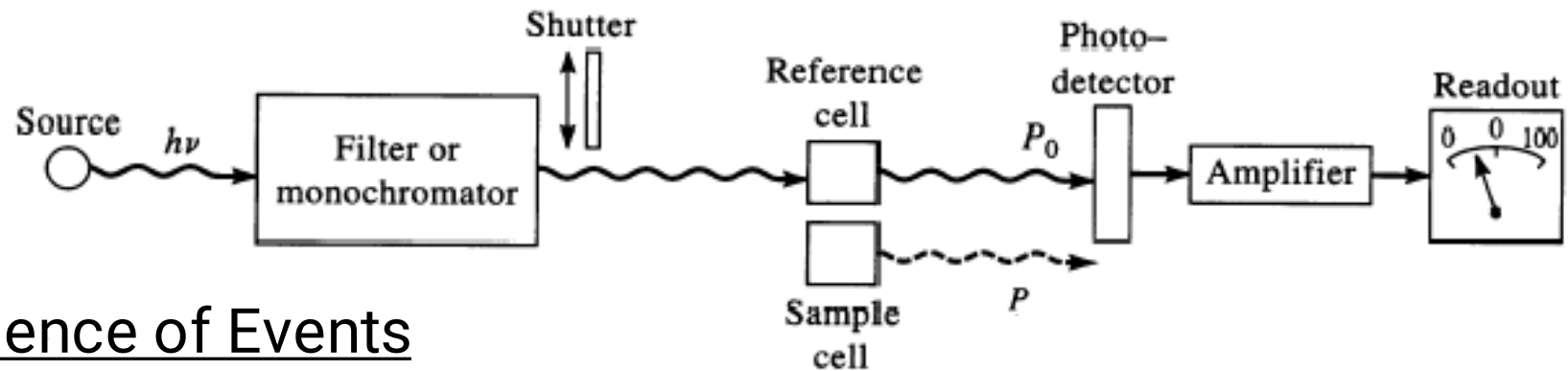
Architectures

1) Single Beam

2) Double Beam

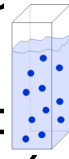
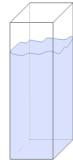
- Spatially Separated
- Temporally Separated

Single Beam Instrument



Sequence of Events

- 1) Light Source On
- 2) Reference in holder
- 3) Open Shutter
- 4) Measure light (P_0)
- 5) Raster λ and repeat 4
- 6) Close Shutter
- 7) Sample cell in holder
- 8) Open Shutter
- 9) Measure intensity (P)
- 10) Raster λ and repeat 9
- 11) Close Shutter



Pros:

- Simple
- Less expensive
- Less optics
- Less moving parts
- Higher light intensity
- Can use the same cuvette

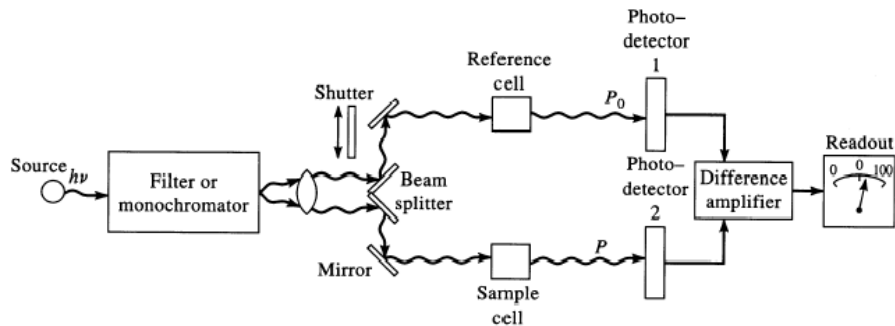
Cons:

- Changes over time
- Better for short term experiments
- Manually move samples

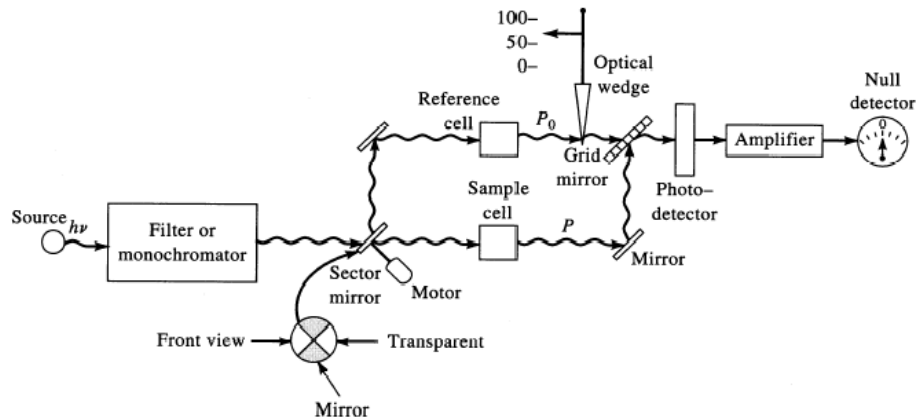
$$A = -\log T = \log P_0/P$$

Double Beam Instrument

Spatially Separated



Temporally Separated



Compensates for:

1) Lamp Fluctuations

Sources of Instability in Metal Halide Arc Discharge Lamps

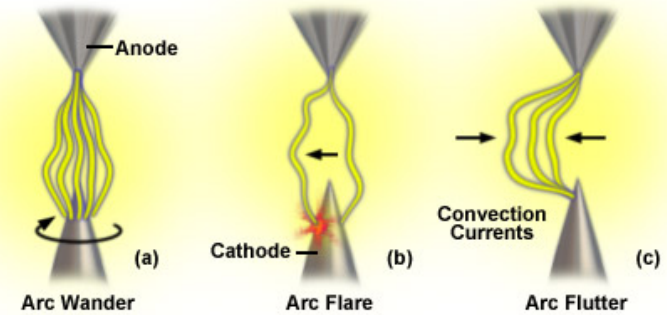
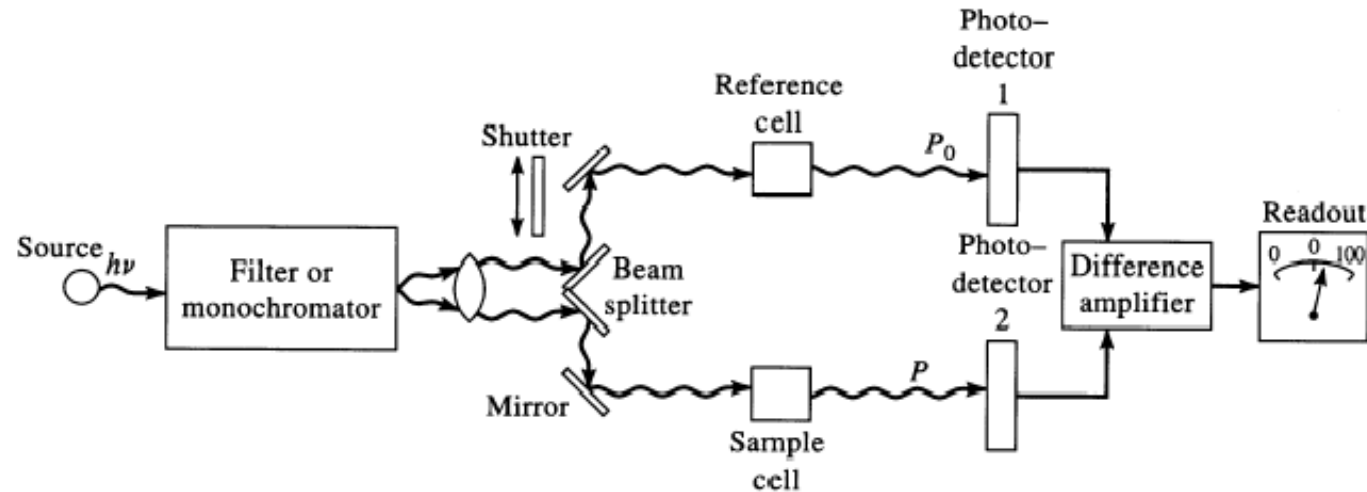


Figure 1

- 2) Temperature changes
- 3) Amplifier changes
- 4) Electromagnetic noise
- 5) Voltage spikes
- 6) Continuous recording

Double Beam Instrument: Spatial



Sequence of Events

- 1) Light Source On
- 2) Reference and sample in holder
- 3) Open Shutter
- 4) Measure detector 1 (P_0) and 2 (P)
- 5) Raster λ and repeat 4
- 6) Close Shutter

Pros:

- Both samples simultaneously
- Less moving parts (than temporal)

Cons:

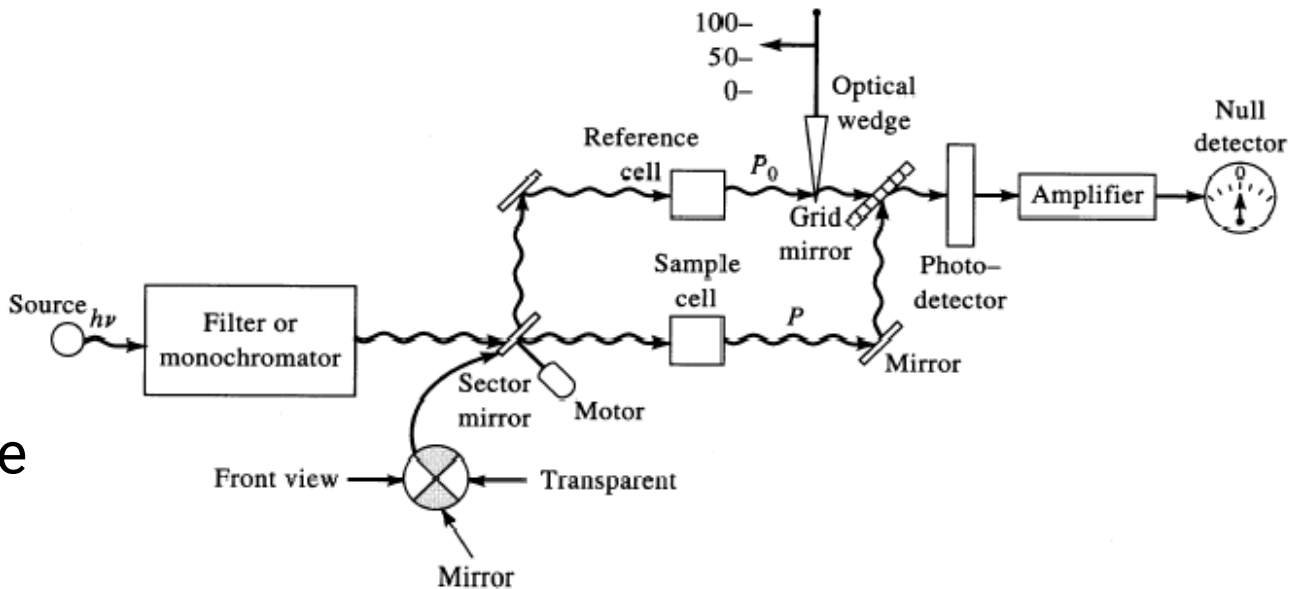
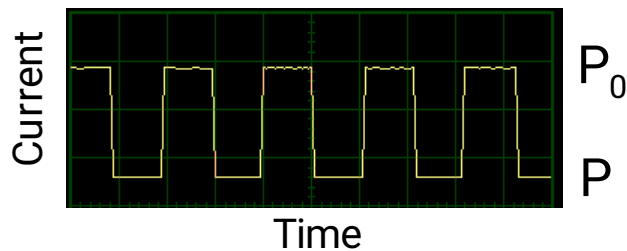
- Two different cuvettes
- Two different detectors
- $\frac{1}{2}$ the intensity
- More expensive

$$A = -\log T = \log P_0/P$$

Double Beam Instrument: Temporal

Sequence of Events

- 1) Light Source On
- 2) Reference and sample
- 3) Rotate Chopper
- 4) Open Shutter
- 5) Monitor detector



Pros:

Both samples
"simultaneously"
Same Detector

Cons:

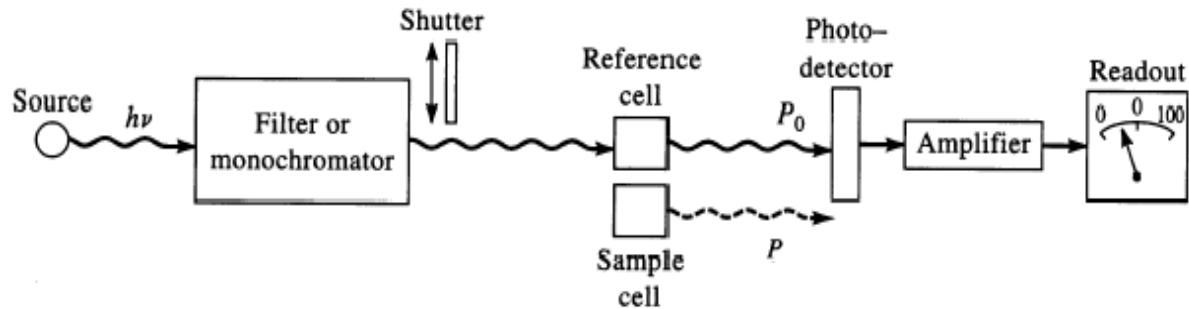
Two different cuvettes
 $\frac{1}{2}$ the intensity
rotating mirrors
not really simultaneous

- 6) Raster λ and repeat 4
- 7) Close Shutter

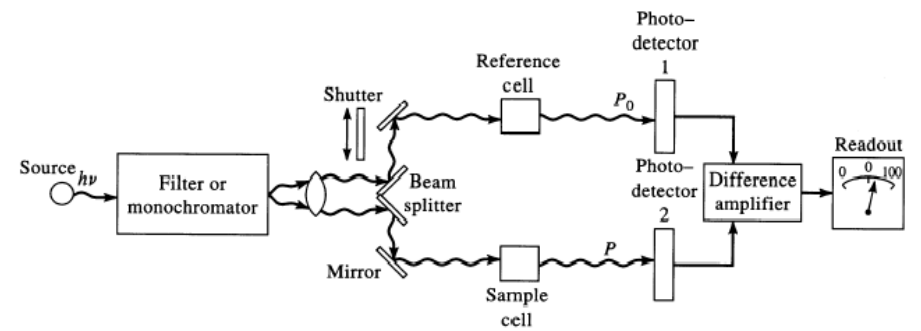
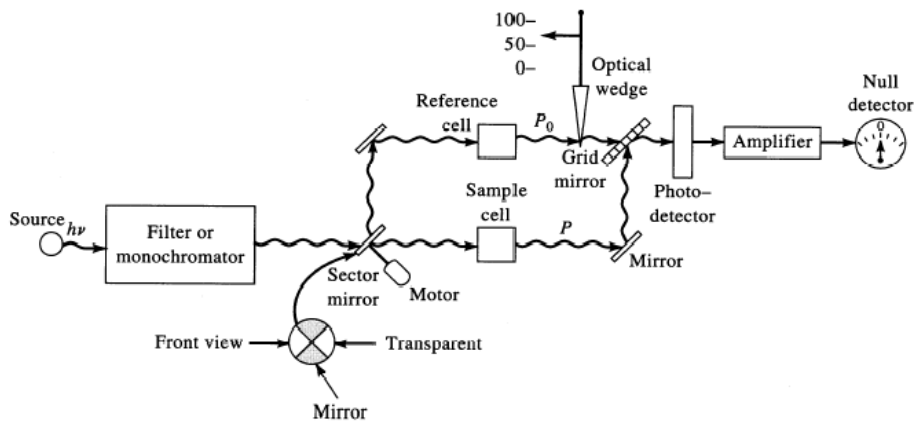
$$A = -\log T = \log P_0/P$$

Instrument Architectures

Single Beam

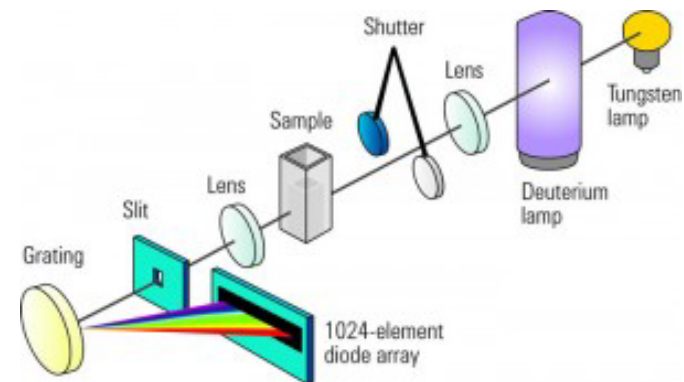
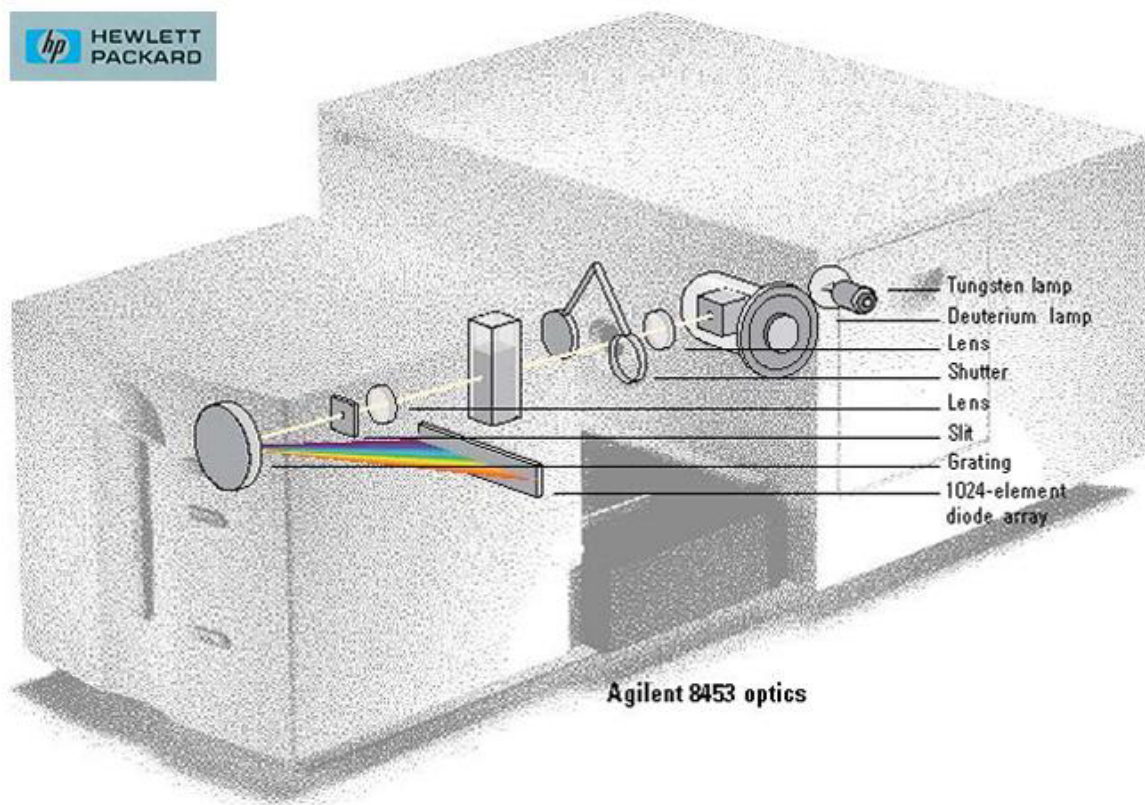


Double Beam



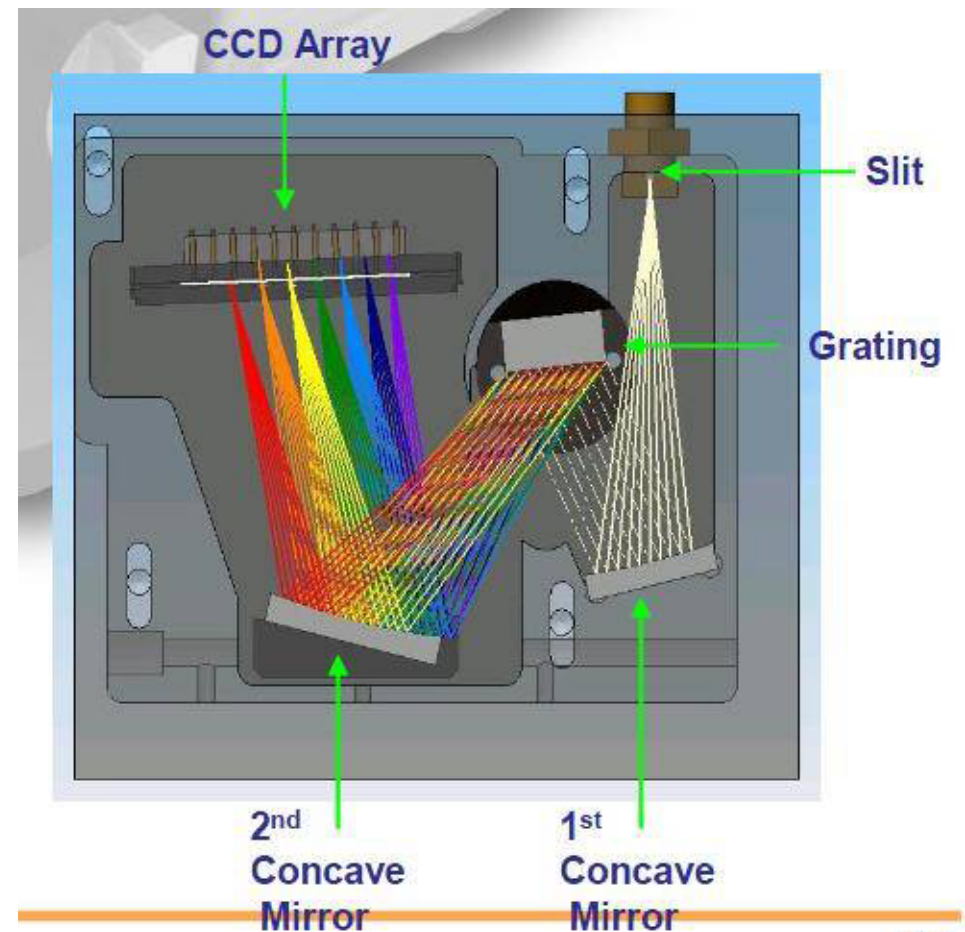
Instrument Architectures

Agilent 8453: Single Beam, Diode Array Detector



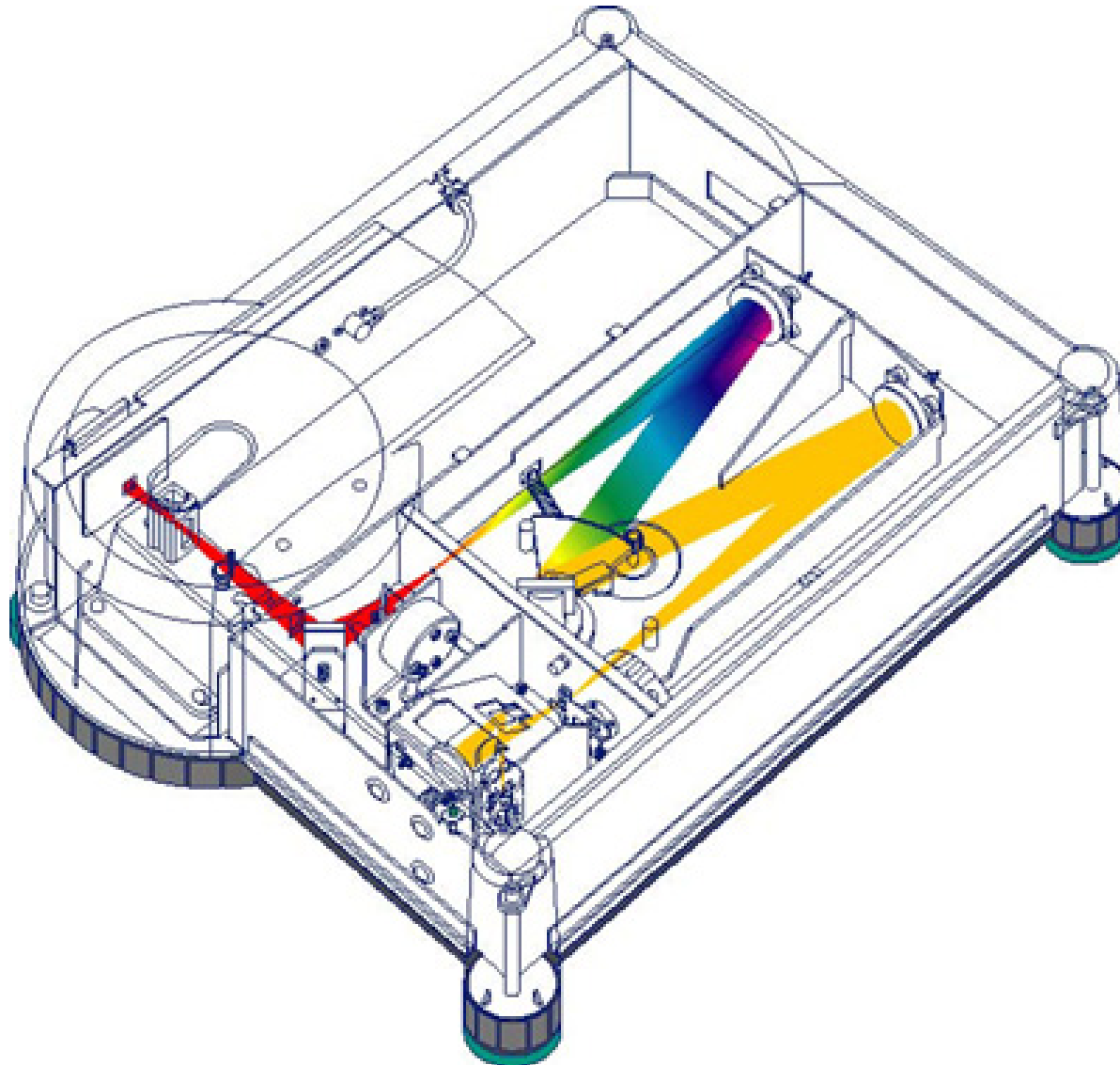
Instrument Architectures

Ocean Optics: Single Beam, CCD Detector



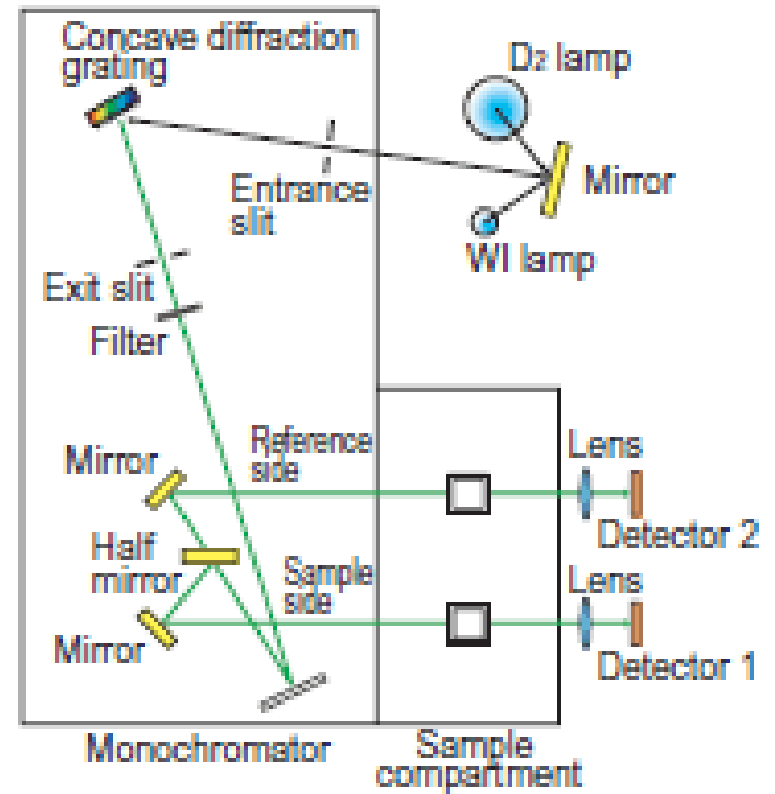
Instrument Architectures

Cary 50: Single Beam, PMT detector



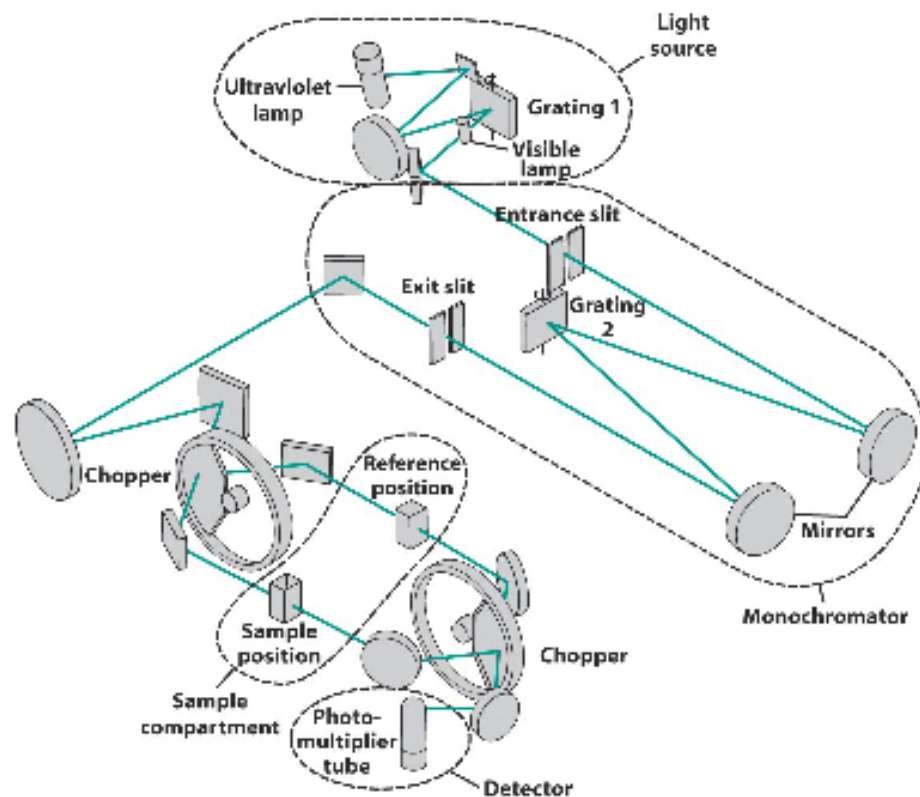
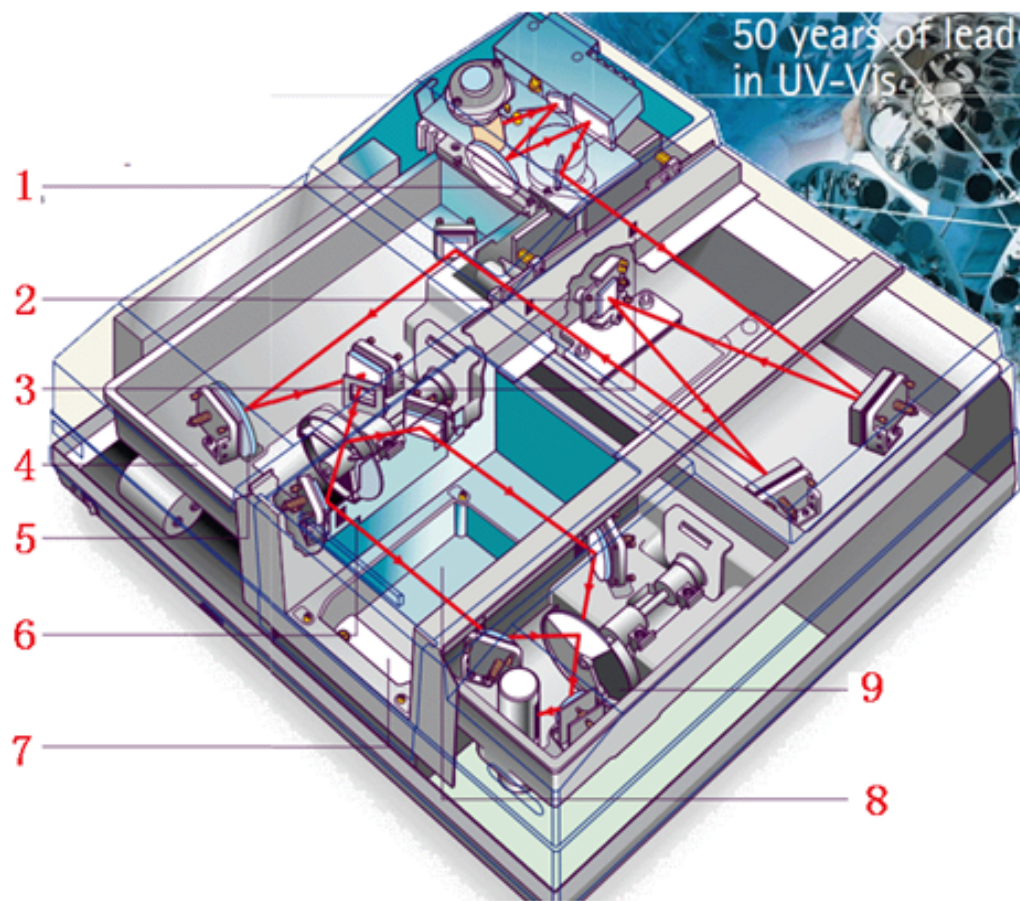
Instrument Architectures

Hitachi U-2900: Double Beam, 2 x PMT detector



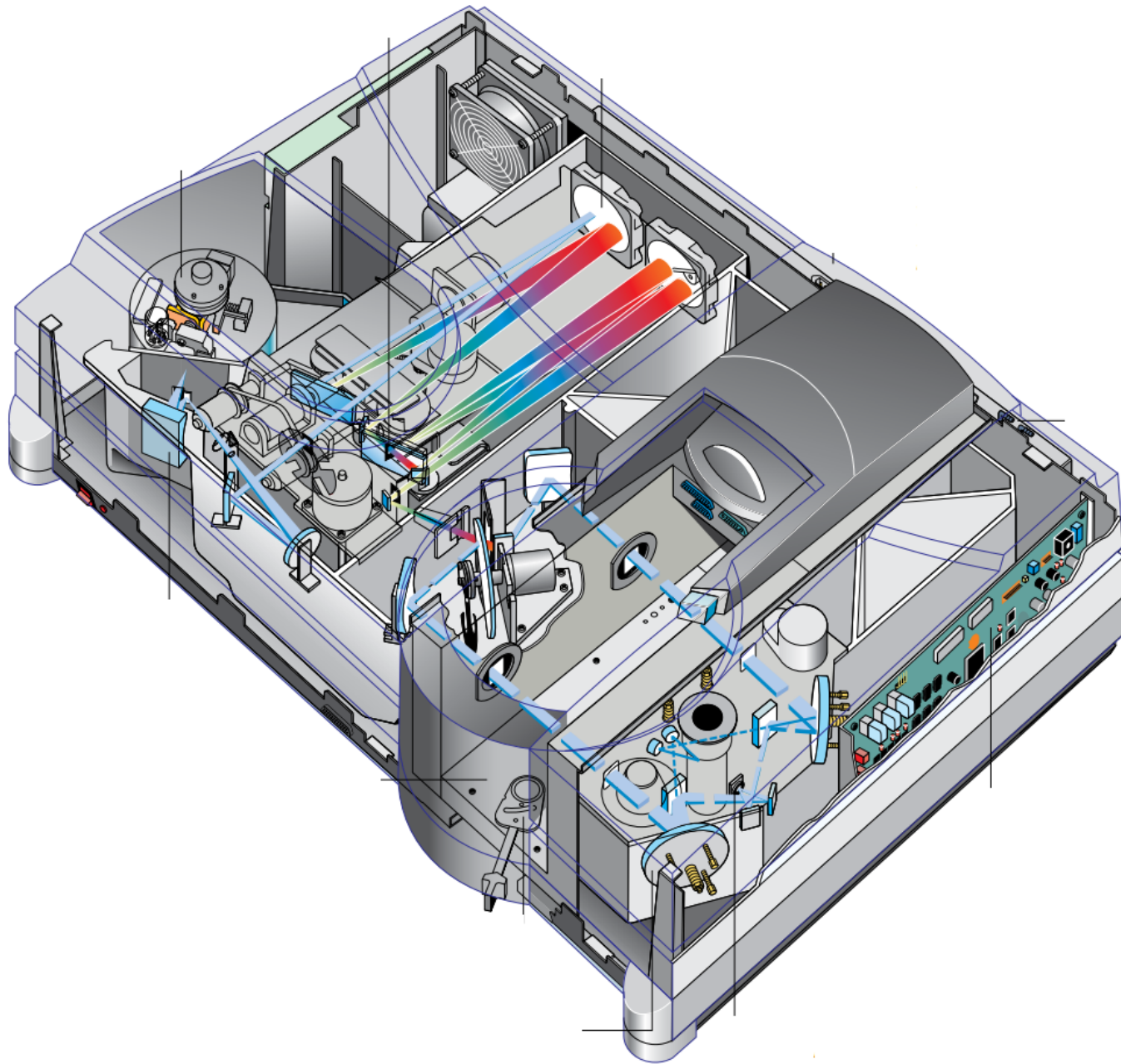
Instrument Architectures

Cary 300: Double Beam, PMT detector



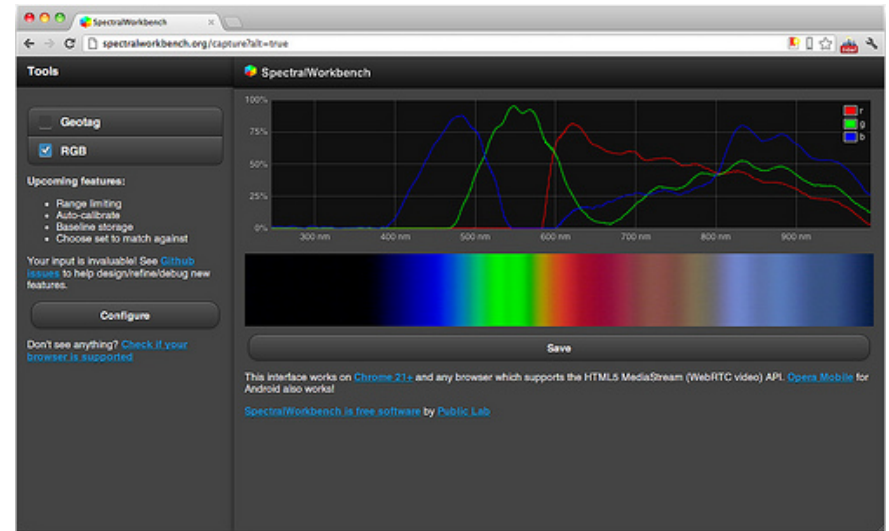
Instrument Architectures

Cary 5000: Double Beam, PMT detector



Single Beam Instrument

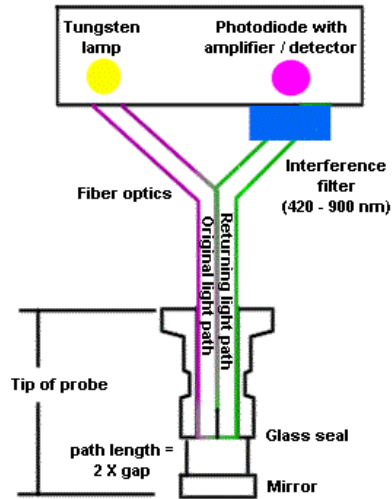
DIY Spectrometer



<http://publiclab.org/wiki/spectrometer>

Other Sampling Accessories

Probe-type



Cryostat



Fiber Optics



Microplate Spectrometer



The Sample: Solids

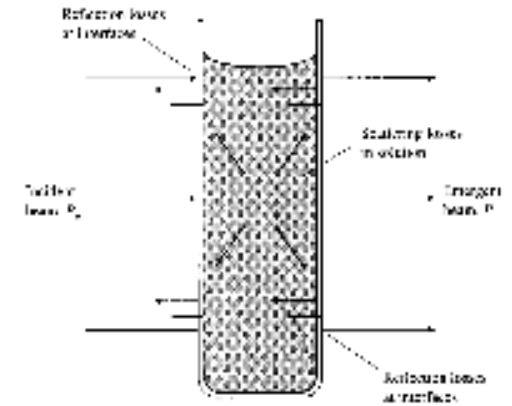
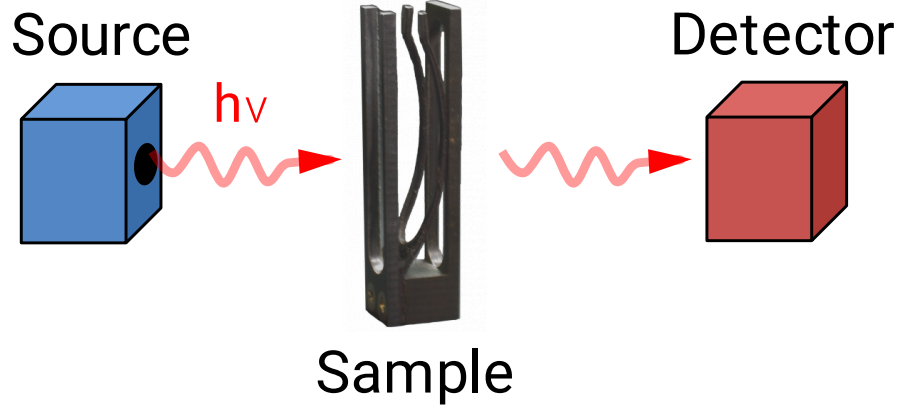
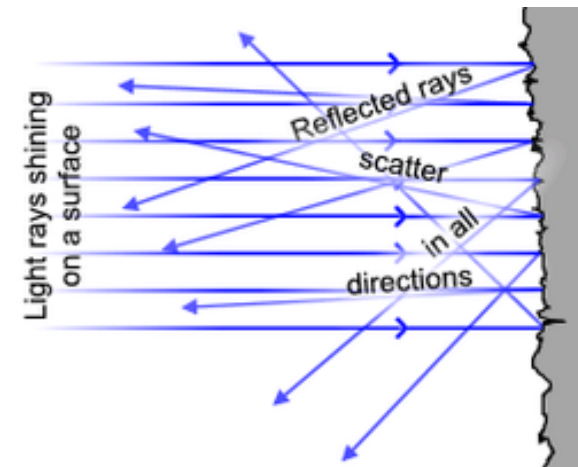


Figure 1.1-1 Reflection and scattering, loss.

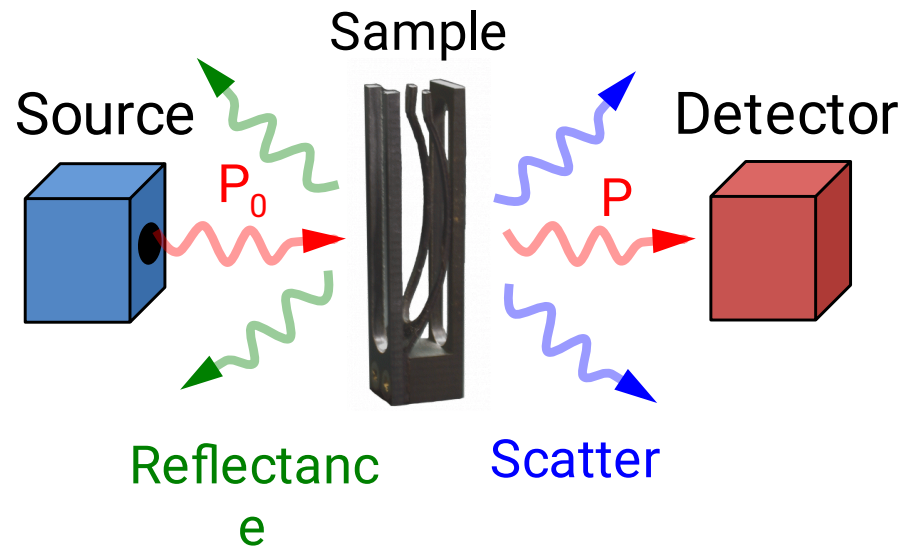
Solids/Films

- More scatter, more reflectance
- No reference



The Sample: Solids

$$A = -\log T = \log P_0/P$$



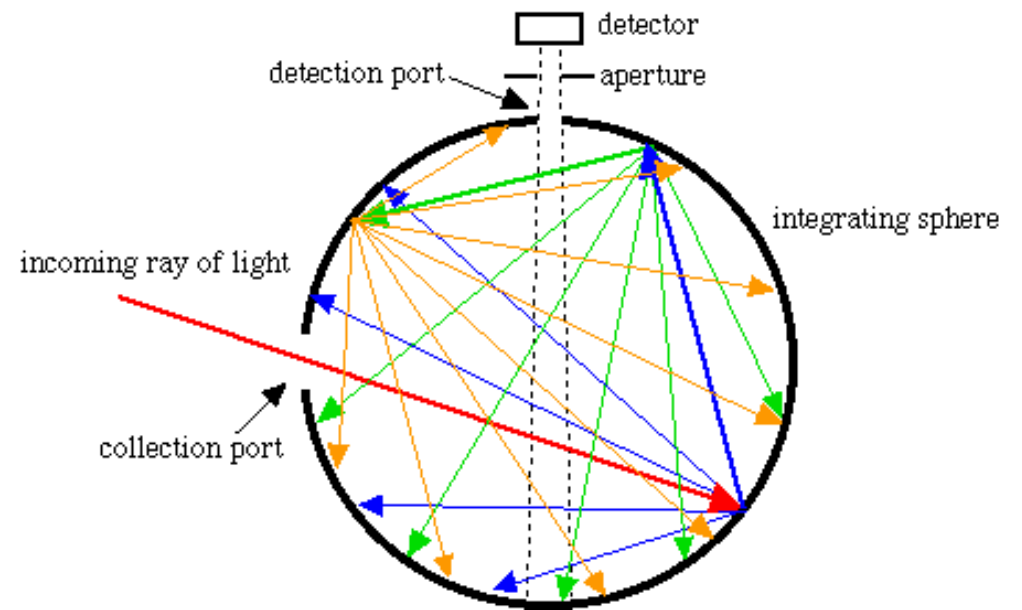
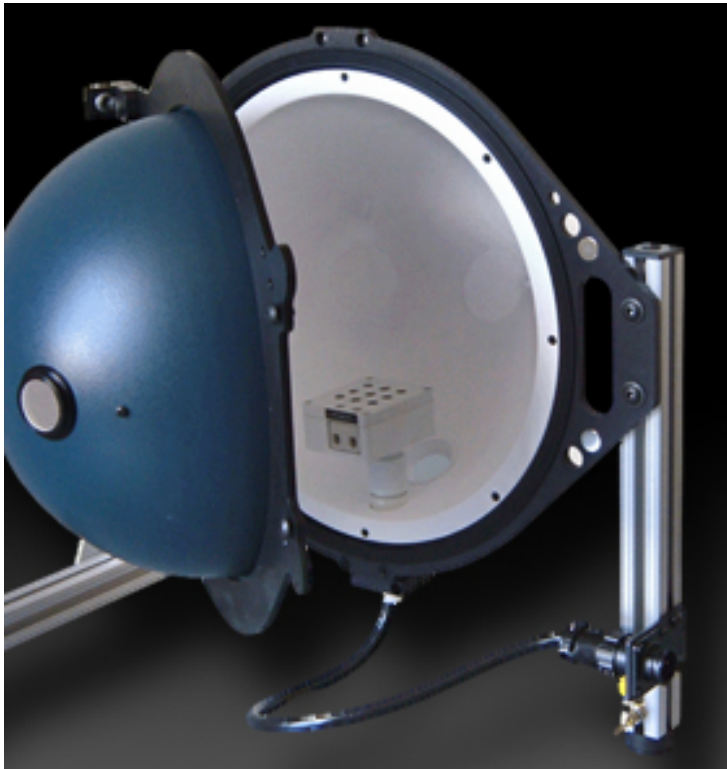
P does not take into account **reflectance** and **scatter**!

Measured $A >$ Actual A

More scatter/reflectance = More error

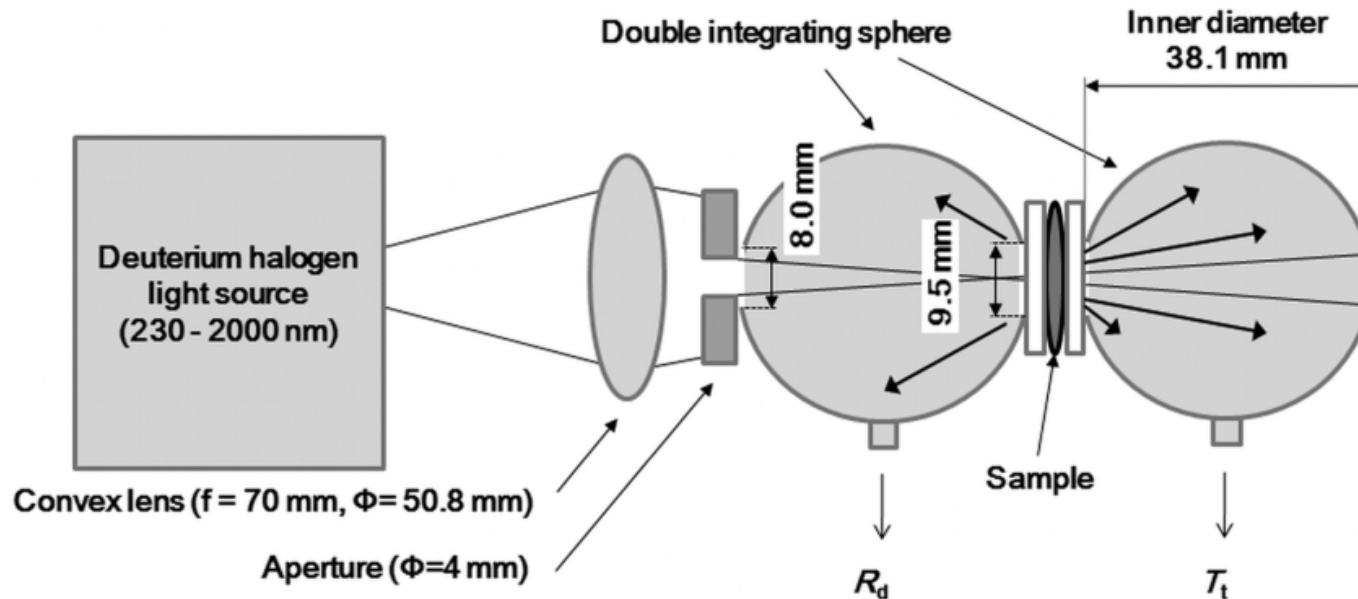
The Sample: Solids

Integrating Sphere



Solid Sample

$$A = -\log T = \log P_0/P$$



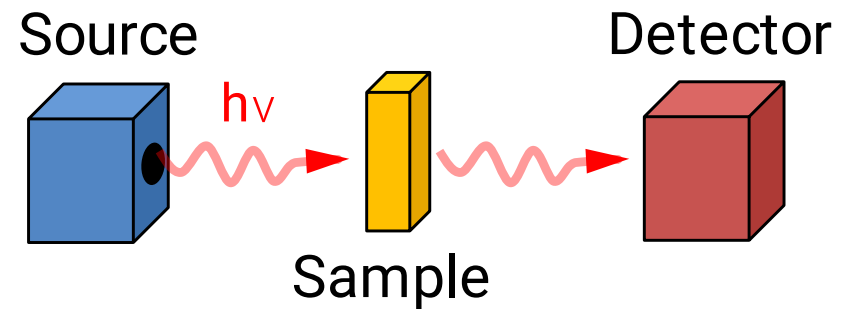
$$P_0 \approx T_{t(\text{without sample})} - R_{d(\text{with sample})}$$

$$P \approx T_{t(\text{with sample})}$$

$$A \approx \log \left(\frac{T_{t(\text{without sample})} - R_{d(\text{with sample})}}{T_{t(\text{with sample})}} \right)$$

Outline

- 1) Beer's Law
- 2) Absorption Spectrum
- 3) Instrument Components
 - Light sources
 - Monochrometers
 - Detectors
 - Other components
 - The sample
- 4) Instrument Architectures
- 5) Applications
- 6) Limitations



HPLC

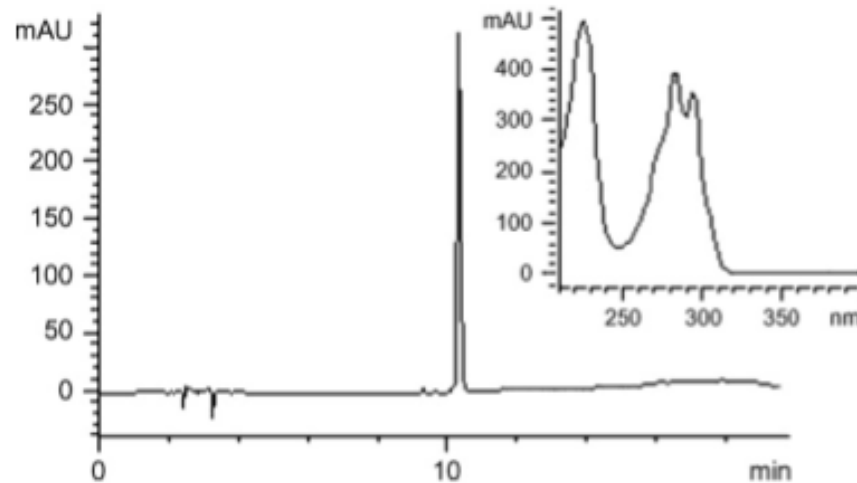
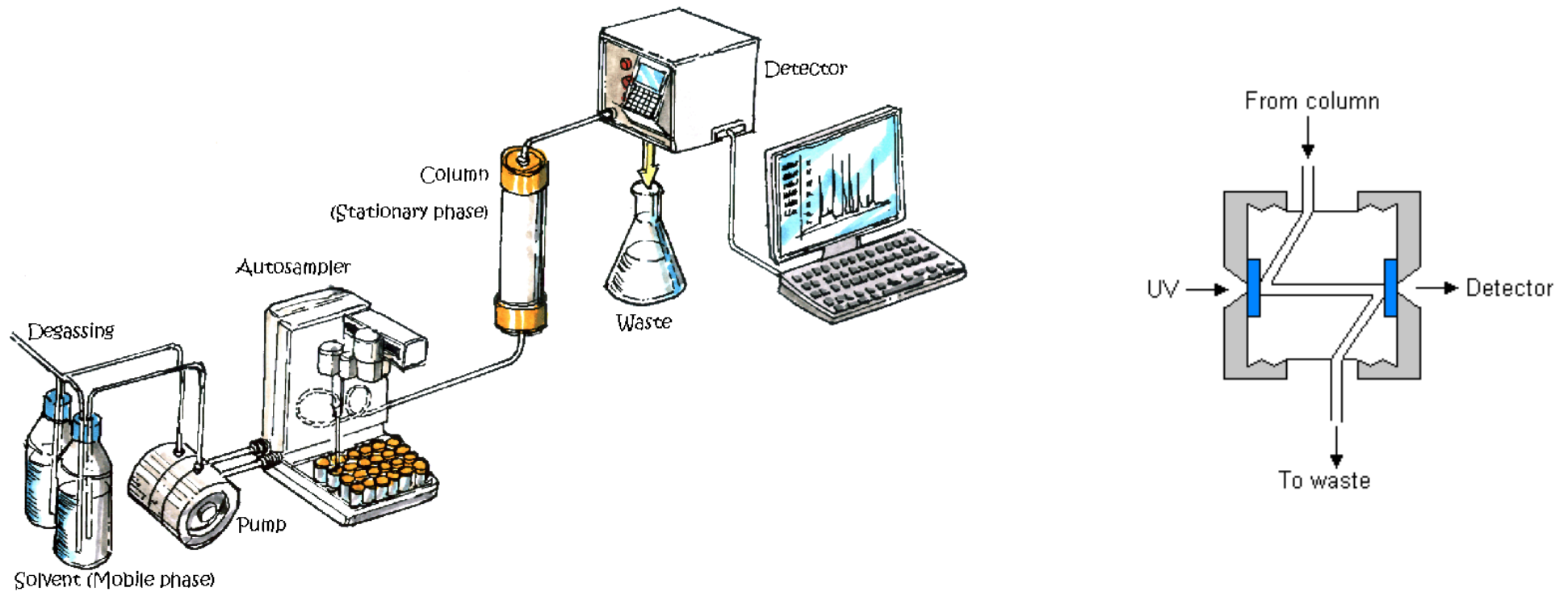
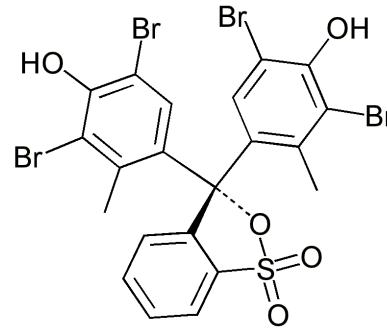


Fig. 3. HPLC chromatogram and UV-vis spectra (inset) of Bromo-Dragonfly. The HPLC trace was detected at 210 nm.

pK_a Determination

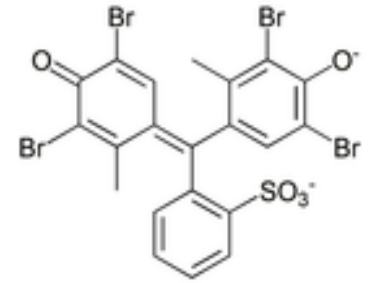
Titration of bromocresol

- 1) Bromocresol Green in H₂O
- 2) Titrate with base
- 3) Monitor pH
- 4) Monitor Absorption Change
- 5) Graph absorbance vs pH
- 6) Inflection point = pK_a

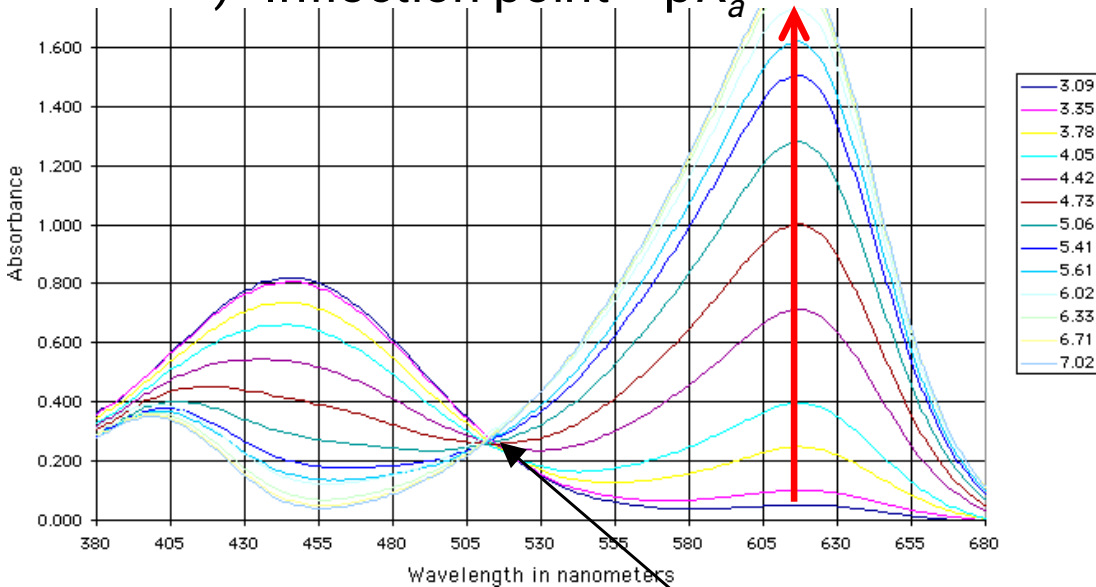


yellow

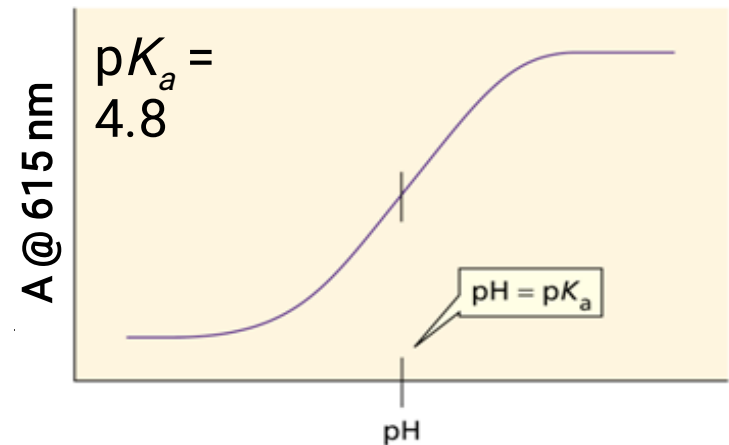
- H⁺
+ H⁺



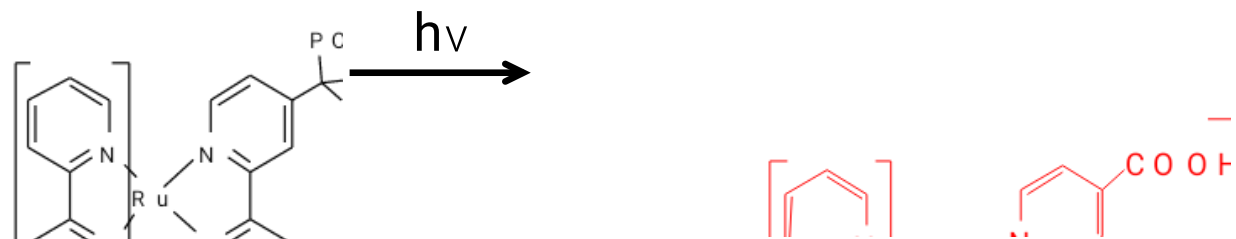
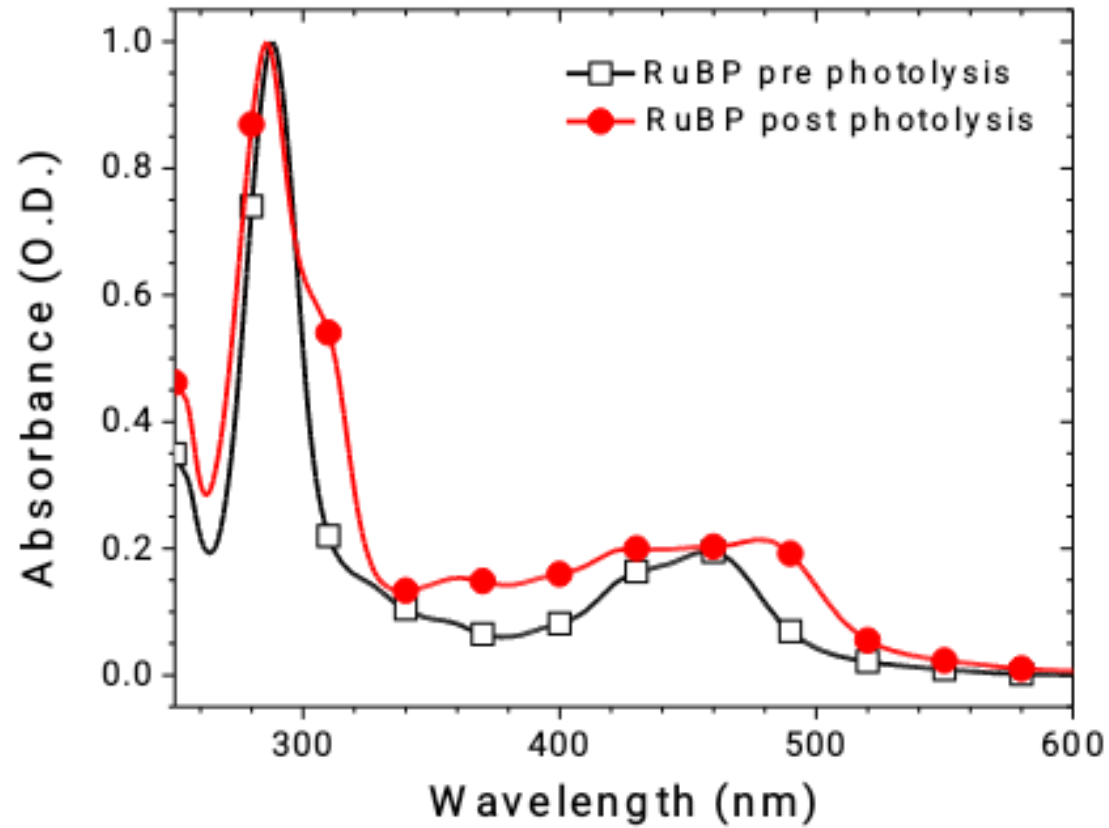
blue



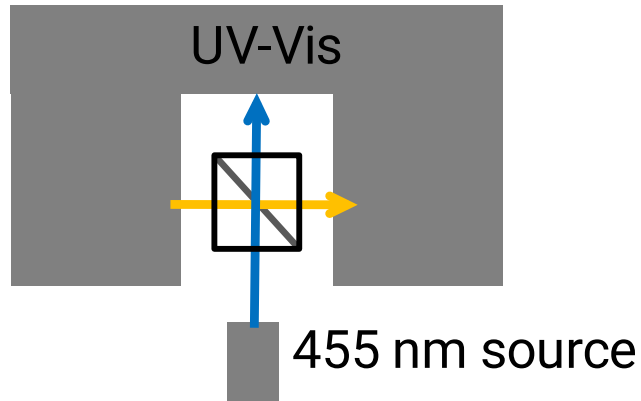
Isosbestic point



Reaction Kinetics

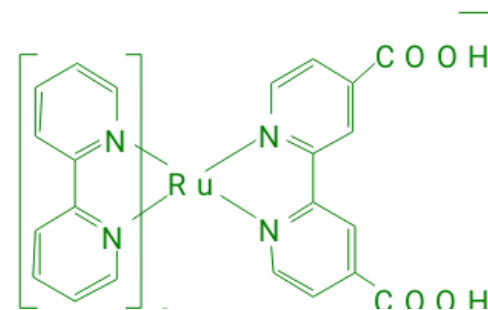
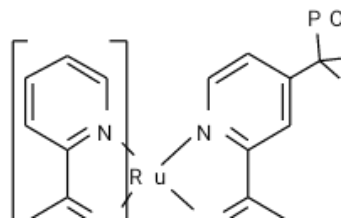
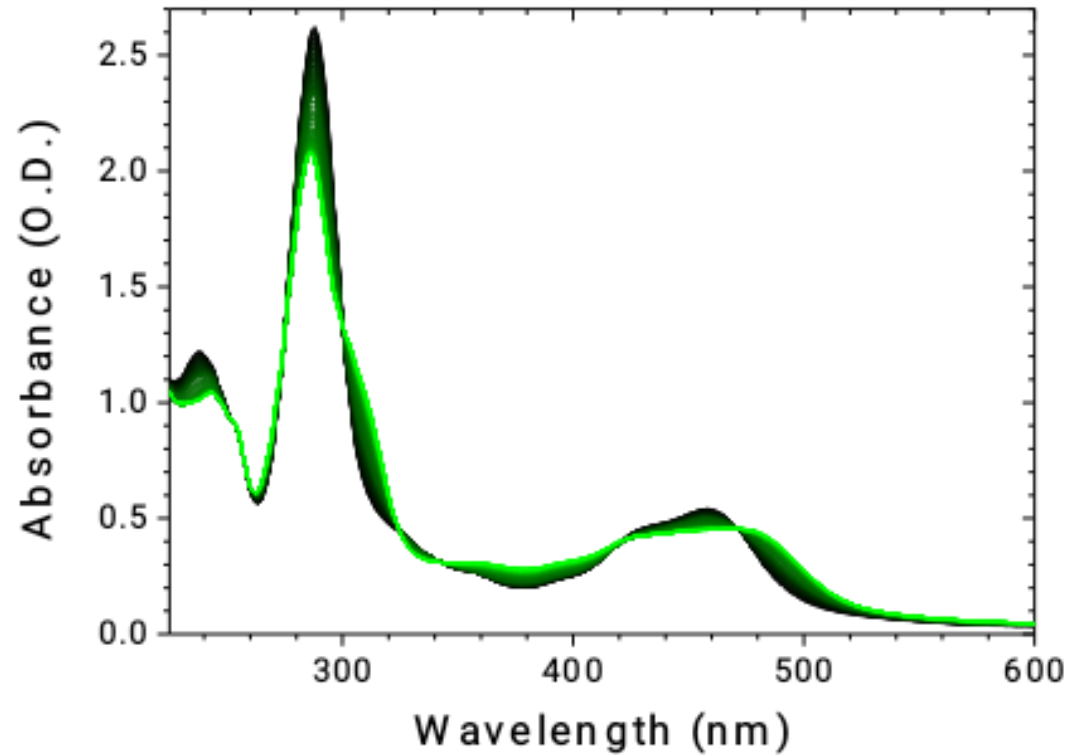


Real Time Monitoring

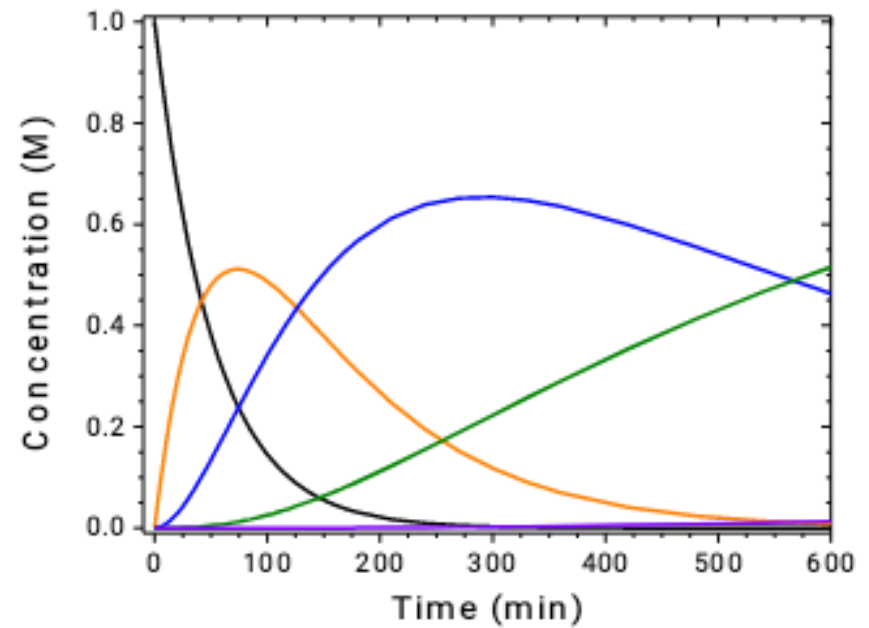
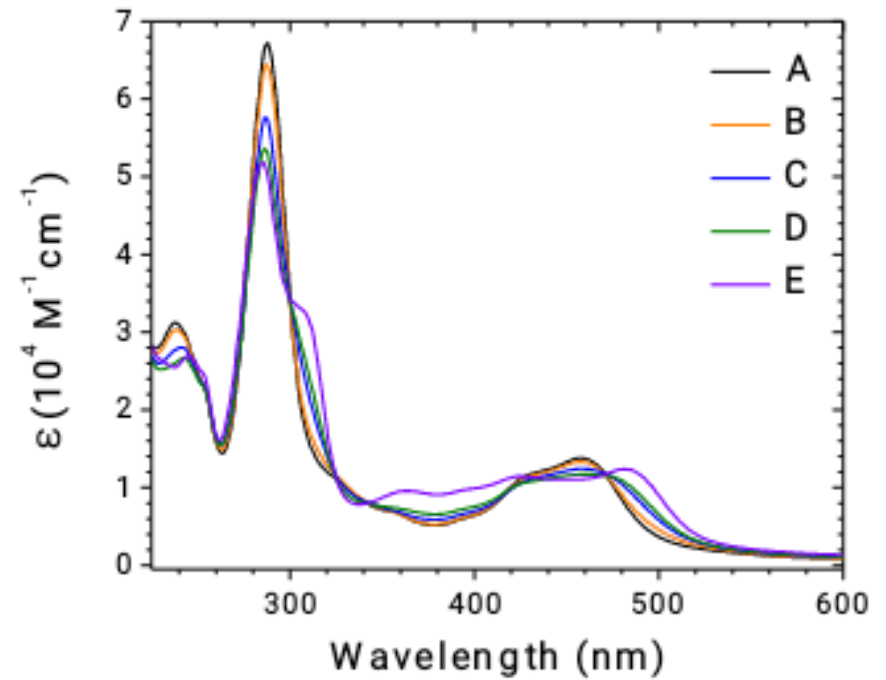
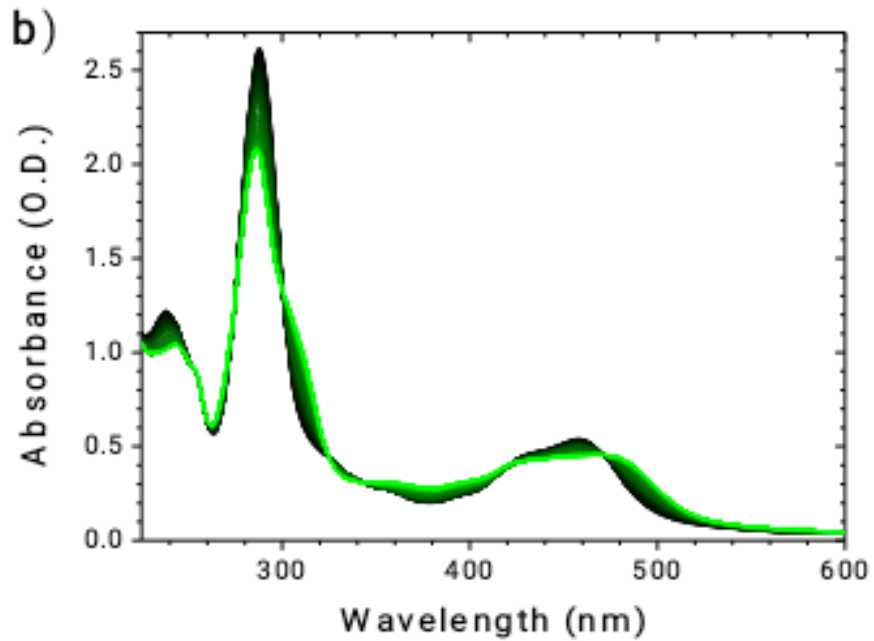


3mL of 40 μ M RuBP in pH 1, atm

Monitor: Every 5 min for 180 min
Every 30 min for 180 min
Every 60 min for 3420 min



Spectral Fitting

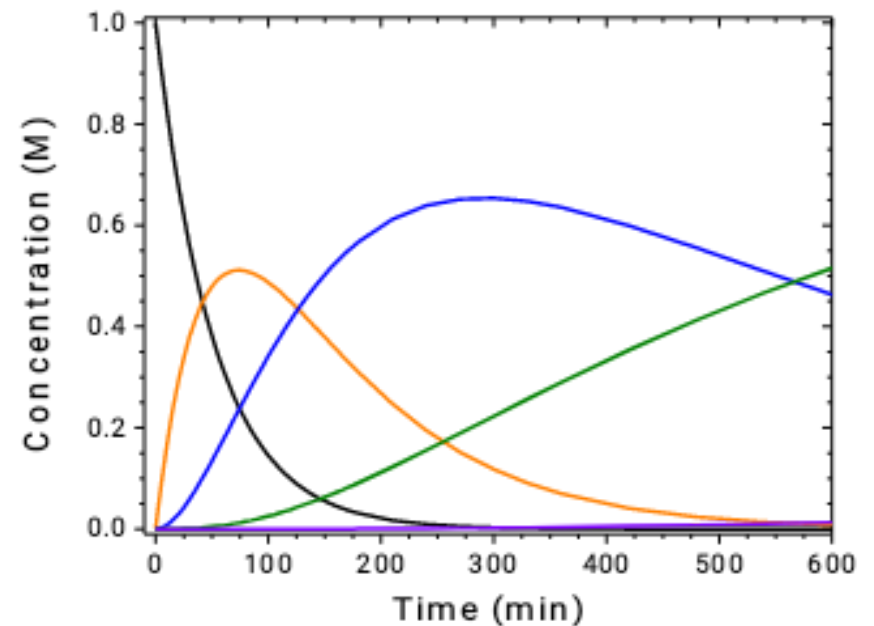
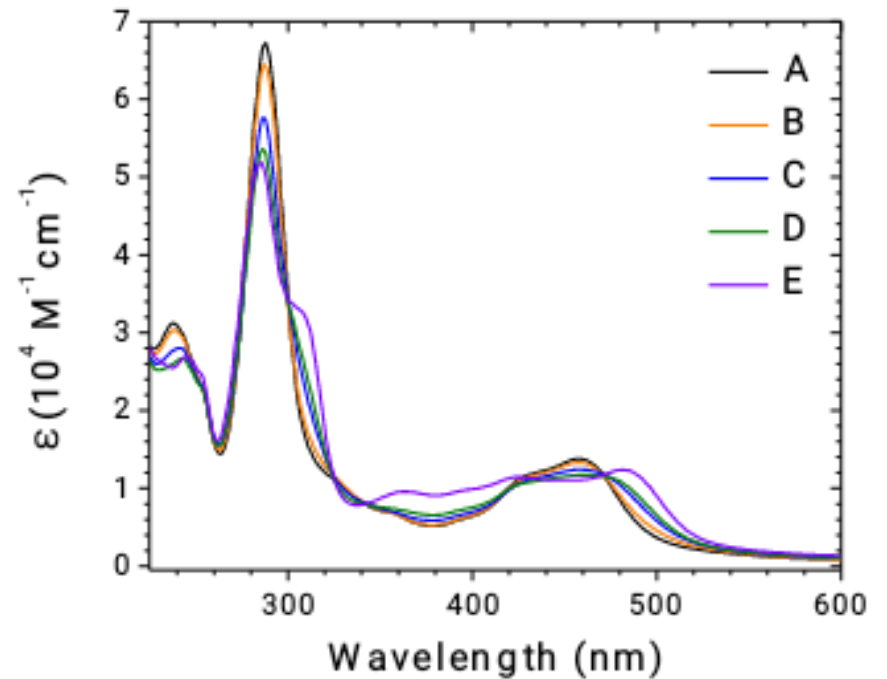


Spectral Fitting

Table 1. Reaction rate constants for the photodecomposition of RuBP (error in parentheses).^a

Solvent	$k_{A \rightarrow B}$ (10^{-4} s^{-1})	$k_{B \rightarrow C}$ (10^{-4} s^{-1})	$k_{C \rightarrow D}$ (10^{-5} s^{-1})	$k_{D \rightarrow E}$ (10^{-6} s^{-1})
H ₂ O	2.8 (0.06)	1.3 (0.07)	3.4 (0.07)	4.0 (0.6)
D ₂ O	8.3 (0.08)	1.1 (0.02)	2.9 (0.07)	4.8 (1.1)
0.1 M HClO ₄	3.2 (0.3)	1.5 (0.06)	2.9 (0.09)	1.6 (0.4)
0.1 M HClO ₄ ^b	16.4 (1.4)	2.9 (0.02)	4.9 (0.04)	2.9 (0.3)

a) In atmosphere with 455 nm (50 mW/cm²) irradiation unless otherwise noted. b) Bubbled with pure O₂.



Potential Complications

With the Sample

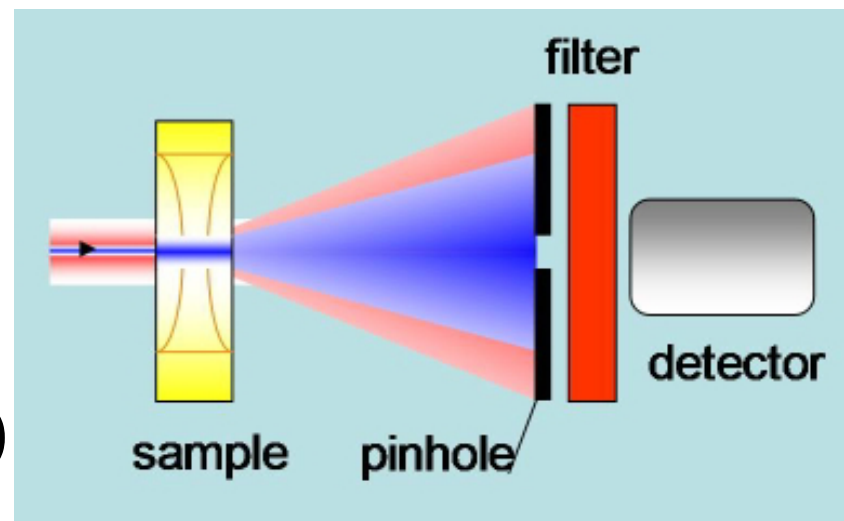
- Photo Reaction/Decomposition
- Concentration too high
 - non-linear ($A > 2$)
 - Aggregation
 - Refractive index change
- Air bubble generation

With the Cuvette + Solvent

- Cuvette non-uniformity
- Sample holder mobility
- Lensing (abs + heat)
- Temperature (line broadening)

With the Instrument

- Lamp Stability
- Room Lighting
- Noise



Absorption End

Any Questions?